

## A Comparative Study on the Toxicity of a Synthetic Pyrethroid, Deltamethrin and a Neem Based Pesticide, Azadirachtin to *Poecilia reticulata* Peters 1859 (Cyprinodontiformes: Poeciliidae)

S. Israel Stalin<sup>1</sup>, S. Kiruba<sup>1</sup>, S. Sam Manohar Das<sup>1,\*</sup>

<sup>1</sup> Scott Christian College (Autonomous), Department of PG studies and Research Centre in Zoology, Nagercoil 629 003, Tamil Nadu, India.

\* Corresponding Author: Tel.: +91.4652 231807; Fax: +91.4652 229800;  
E-mail: sambiocontrol@gmail.com

Received 12 February 2007  
Accepted 25 September 2007

### Abstract

Addition of pollutants changes the natural qualities of water. Pesticides in agricultural runoff affect fish and other aquatic organisms. Fish are common indicators of water pollution status. This study was done to check whether plant based pesticides are less toxic to fish and other non-target organisms, compared to chemical pesticides. Bioassays of a natural pesticide of plant origin (Azadirachtin) and a synthetic pyrethroid, deltamethrin were separately done on a freshwater teleost, *Poecilia reticulata* Peters 1859. Toxicity estimations were done following static bioassay and probit analyses models. The 96 h LC<sub>50</sub> of deltamethrin is 0.0019 and azadirachtin is 0.011 mg/L. The plant based pesticide; azadirachtin is less toxic to fish compared to deltamethrin, a synthetic pyrethroid. Plant based pesticides contain easily biodegradable molecules which are more target specific than the highly persistent broad-spectrum synthetic chemical moieties. Use of plant based pesticides is less disastrous and more ecofriendly. This study is done to compare the non-target toxicity of a natural pesticide of plant origin (Azadirachtin) with a synthetic pyrethroid, deltamethrin on a fresh water teleost, *Poecilia reticulata* Peters 1859.

**Key words:** deltamethrin, azadirachtin, *Poecilia reticulata*, non-target toxicity, 96h LC<sub>50</sub>.

### Introduction

Increased use of chemical pesticide results in the excess inflow of toxic chemicals, mainly into the aquatic ecosystem (Baskaran *et al.*, 1989; Kalavathy *et al.*, 2001). The aquatic flora and fauna are affected by the toxic substances which eventually enter in to their systems or bring about external damages (Pant and Singh, 1983; Hodson, 1988; Johal and Dua, 1995). Several species of fish are susceptible to deleterious effects when exposed to heavy metals, pesticides and other environmental stressors (Khangrat *et al.*, 1988; Areechon and Plump, 1990).

Recent emphasis is on the use of natural pesticides, which are usually of plant origin. Azadirachtin derived from neem (*Azadirachta indica* A. Juss) is a very effective and extensively used pesticide. Pesticides based on azadirachtin may have direct adverse effects on aquatic organisms and their toxicity depends on various factors. It has been reported that neem extracts in aquatic environments are lethal to benthic populations and drastically decrease the number of organisms in the food web and nutrient cycling process (Goktepe *et al.*, 2002; El-Shazly *et al.*, 2000). Even though *Azadirachtin* is not likely to accumulate or cause long-term side effects, toxicity to fish is only moderate and may not kill fish under normal use (Miller and Uetz, 1998).

Pesticides containing bioactive compounds from the neem plant, *Azadirachta indica* Juss are reported to be target specific and comparatively less toxic. Plants are virtually inexhaustible sources of structurally diverse and biologically active substances (Istvan,

2000). Some plants contain compounds of various classes that have insecticidal, piscicidal and molluscicidal properties. Unlike synthetic chemical pesticides, which leave harmful residues in the aquatic environment (Koesomadinata, 1980; Cagauan, 1990; Cagaun and Arce, 1992), botanical insecticides are believed to be more environmentally friendlier because they are easily biodegraded and leave no residues in the environment. Most of the pesticides, both plant based and chemical, applied in the various agro ecosystems reach water bodies through runoff affecting fish, the most abundant aquatic organism, and a variety of other fauna. Persistent chemical molecules with long half-life periods found in chemical pesticides pose a threat to fish and also to the human population consuming the affected fish. The purpose of this study is to establish whether azadirachtin has any non-target effect on fish and to compare its toxic impact with deltamethrin, a powerful chemical pesticide. Based on the results, the use of the less toxic pesticide could be promoted among the agriculturists. It is possible to substitute chemical pesticides with pesticides of plant origin. This needs an extensive study on the properties of the two types of pesticides, especially a study of their toxic effect on aquatic organisms (Lockwood, 1976; Woin, 1988; Glow and Godzi, 1994; Christina, 2004). Fish are considered to be indicators of water pollution (von Westernhagen, 1988). Guppies are common fresh water fish capable of tolerating wide fluctuations in water quality and hence have been selected for this work. Any impact on such hardy species is considered to be ultimate, since other

susceptible species succumb at concentrations, much lower than that could be tolerated by this species. This study is designed to compare the toxicity of a synthetic pyrethroid, deltamethrin and a neem derivative, azadirachtin in the aquatic environment using *Poecilia reticulata* Peters as the test species.

## Materials and methods

### Test Animals

Healthy, unsexed *P. reticulata* (Guppy) (1.12±0.5 g) were brought to the laboratory from the freshwater pond of the Zoology Research Laboratory, Scott Christian College (Autonomous), Nagercoil. The fish were acclimatized to the laboratory environment for about 5 days. They were fed with commercial food pellets and only healthy fish were used in the experiments. Toxicant was prepared in wide ranging concentrations and the fish were exposed in one litre glass aquaria. This range finding test gave an idea as to the effective concentration of the toxicant required to bring about mortality in the test species and 15 different concentrations were set up. The test water used for dilution was drawn from the campus well.

### Experimental Design

Two different types of toxicants were used; deltamethrin and juerken (Azadirachtin). Stock solutions were prepared by adding 0.5 ml insecticide with 999.5 ml water and stored in glass bottles. Working stock solution (0.1%) of the toxicant was prepared and stored in glass bottles. Proper dilutions of the stock solutions were made to make a series of test solutions of varying concentrations. *P. reticulata* is exposed to different concentrations of deltamethrin ranging from 0.0014 to 0.0028 mg/L and azadirachtin, ranging from 0.002 to 0.030 mg/L.

To each concentration, 10 healthy fish were introduced. The mortality was recorded after 6, 12, 18, 24, 36, 48, 72, 96 and 120 h of exposure. The mortality of a single fish was recorded as 10 percent mortality. The toxicity studies were conducted following the static bioassay method (Sprague, 1973).

The toxicant was renewed everyday. The dead fish were carefully removed to avoid contamination. The mortality of exposed fish was observed at different intervals.

### Calculation of LC<sub>50</sub> Values

The results of the static bioassay were analysed using linear regression probit analysis (Finney, 1971) using the statistical package, (POLO-PC, LeOra software, 1987) The LC<sub>50</sub> values were calculated for 24, 48, 72, 96, and 120 h of exposure to deltamethrin and azadirachtin separately. The upper and lower fiducial limits were also calculated.

### Results

The LC<sub>50</sub> values for 24, 48, 72, 96 and 120 h of exposure to deltamethrin are 0.0024, 0.0021, 0.0020, 0.0019 and 0.0018 and to azadirachtin 0.0020, 0.017, 0.014, 0.011 and 0.010 mg/L, respectively (Table 1). The toxicity curves (Figure 1 A and B) follow the regular pattern with asymptotes close to 72 hours. Based on probit analysis, regression equation for delta methrin (96 h exposure) is  $Y = 17.72x - 17.33$  (Table 2) and azadirachtin (96 h exposure) is  $Y = 4.24x + 0.43$  (Table 3).

### Discussion

Newer biological pesticides are developed to replace deleterious chemical pesticides. Even though chemical pesticides are target specific and effective, their impact on the environment is mostly deleterious. Plant based pesticides contain active principles with low half-life period and their effects on the environment are not too detrimental (Sharma *et al.*, 1995). In the present study, the pesticide containing azadirachtin is less toxic to fish compared to deltamethrin. The 96 h LC<sub>50</sub> of deltamethrin is 0.0019 mg/L. whereas azadirachtin is much higher 0.011 mg/L indicating the less toxic nature of the plant based pesticide. Svobodova (2003) reported that the 96 h LC<sub>50</sub> of deltamethrin to *Cyprinus carpio* juveniles was 0.00145 mg/L. Boran *et al.* (2007)

**Table 1.** LC<sub>50</sub> values (mg/L) with their fiducial limits

Hours	Deltamethrin			Azadirachtin		
	LCL	LC <sub>50</sub>	UCL	LCL	LC <sub>50</sub>	UCL
24	0.0022	0.0024	0.0025	0.018	0.02	0.023
48	0.002	0.0021	0.0022	0.015	0.017	0.019
72	0.0019	0.002	0.0021	0.012	0.014	0.016
96	0.0018	0.0019	0.002	0.009	0.011	0.014
120	0.0017	0.0018	0.0019	0.008	0.010	0.012

Note:

LCL = Lower Confidence Limit

UCL = Upper Confidence Limit

LC<sub>50</sub> = Lethal Concentration for 50 percent of the exposed fish

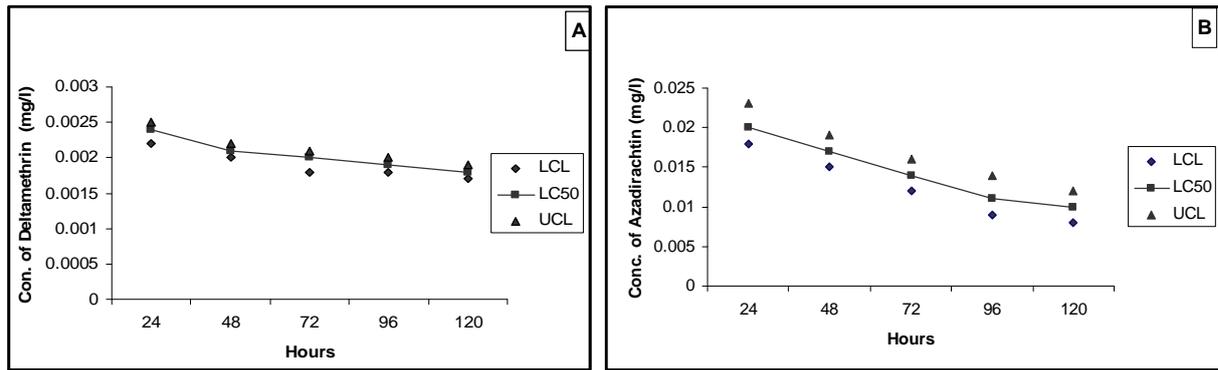


Figure 1. Toxicity curve showing the response of *P. reticulata* A. Deltamethrin B. Azadirachtin.

Table 2. Log-dose/ probit regression line analysis of the response of *Poecilia reticulata* exposed to deltamethrin for 96 h

S. No	Dose (mg/L)	No	Mor %	Log dose	Emp. Pro	Exp. Pro	Work Pro	Wt. Coef	Weight w	Wx	Wy	Y
1	0.0016	10	10	1.2	3.72	3.66	3.72	0.34	3.36	4.05	12.51	3.62
2	0.0017	10	20	1.23	4.16	4.13	4.16	0.47	4.71	5.8	19.6	4.09
3	0.0018	10	40	1.26	4.75	4.57	4.75	0.6	6.01	7.55	28.54	4.53
4	0.0019	10	50	1.28	5	4.99	5.01	0.64	6.37	8.15	31.88	4.94
5	0.002	10	60	1.3	5.25	5.38	5.25	0.6	6.01	7.82	31.56	5.34
6	0.0021	10	70	1.32	5.52	5.76	5.5	0.5	5.03	6.65	27.64	5.71
7	0.0022	10	80	1.34	5.84	6.12	5.8	0.14	4.05	5.44	23.3	6.07
8	0.0023	10	90	1.36	6.28	6.46	6.24	0.27	3.69	3.66	16.78	6.41
9	0.0024	10	100	1.38	7.33	6.78	7.13	0.18	1.8	2.49	20.84	6.74

#### STATISTICS

SW=40.030 SWX=51.605 X Bar=1.389 SWY=204.856 Y Bar=5.118

SWX\*X=66.622 SWY\*Y=1073.500 SWXY=265.604 b Value=17.724

Regression equation  $y=17.724x-17.73$

If  $y=5.0$  then  $x=1.283$  this corresponds to dose of 0.0019

Variance=0.0001 Chi-square=1.02 (with 7 Deg. of freedom p)

Lower Limit 1.2629 log dose 1.2825 Upper Limit 1.3021

Table 3. Log-dose/ probit regression line analysis of the response of *Poecilia reticulata* exposed to Azadirachtin for 96 h

S.No	Dose (mg/L)	No	Mor. %	Log dose	Emp. Pro	Exp. Pro	Work Pro	Wt. Coef.	Weight. w	Wx	Wy	Y
1	0.006	10	20	0.78	4.16	3.83	4.24	0.37	3.7	2.88	15.69	3.74
2	0.008	10	30	0.9	4.48	4.36	4.48	0.56	5.58	5.04	25	4.27
3	0.01	10	40	1	4.75	4.77	4.74	0.63	6.27	6.27	29.74	4.68
4	0.012	10	50	1.08	5	5.11	5	0.63	6.34	6.84	31.7	5.01
5	0.014	10	50	1.15	5	5.39	4.98	0.6	6.01	6.89	29.93	5.3
6	0.016	10	50	1.2	5	5.38	5.98	0.56	5.58	6.72	29.13	5.54
7	0.018	10	70	1.26	5.52	5.85	5.46	0.47	4.71	5.91	25.73	5.76
8	0.02	10	80	1.3	5.84	6.05	5.87	0.44	4.39	5.71	25.57	5.95
9	0.022	10	90	1.34	6.28	6.22	6.28	0.37	3.7	4.97	23.22	6.13
10	0.024	10	100	1.38	7.33	6.39	6.87	0.3	3.02	4.17	20.76	6.29

#### STATISTICS

SW=49.300 SWX=55.413 X Bar=1.124 SWY=256.457 Y Bar=5.202

SWX\*X=63.741 SWY\*Y=1356.993 SWXY=293.575 b Value=4.246

Regression equation  $y=4.246x+0.43$

If  $y=5.0$  then  $x=1.076$  this corresponds to dose of 0.011

Variance=0.0016 Chi-square=3.50 (with 8 deg. of freedom p)

Lower Limit 0.9971 log Dose 1.0764 Upper Limit 1.1557

reported that 96 h LC<sub>50</sub> of carbosulphan to guppies was 0.122 mg/L, methiocarb 1.256 mg/L and carbaryl 1.383 mg/L. Cagauan *et al.* (2004) showed that the lethal concentration of neem to Nile tilapia *Oreochromis niloticus* L. was 12.4 ml/L and mosquito fish *Gambusia affinis* Baird and Girard was 8.31 ml/L and the corresponding 96 h LC<sub>50</sub> values were 2.57 and 3.0 ml/L.

Both deltamethrin and azadirachtin, two established insecticides exhibit non-target toxicity to fish. Comparison of the different LC<sub>50</sub> values clearly indicates that the plant based pesticide is less toxic compared to the chemical one. To reduce the chemical load on the environment, it is suggested that use of plant based pesticides should be encouraged (Schmutterer, 1990). However, care should be taken to use even the plant based pesticide at moderate levels. Furthermore, plant based pesticides disintegrate easily into constituent elements without leaving any indelible impression in different regions of the environment (Khan and Ahmed, 2000).

The test fish in the present study is a hardy ornamental fish. Kharat *et al.* (2003) rated *P. reticulata* as a tolerant species along with *Oreochromis mossambicus* and *G. affinis*. *P. reticulata* is affected by the non-target action of both the chemical and biological pesticides. The impact of the biological pesticide is much less compared to the chemical one, (deltamethrin 96 h LC<sub>50</sub> – 0.0019) (Table 2), and (azadirachtin 96 h LC<sub>50</sub> – 0.011 mg/L) (Table 3). It is advocated that more and more plant products should be developed with proper and targeted action and this eventually helps in keeping the environment free from hazardous chemicals.

### Acknowledgement

The authors wish to thank Dr. S. Prasanna Kumar, Dean of science and HOD of Zoology and Dr. E. James R Daniel, Principal, Scott Christian College (Autonomous), Nagercoil, for the facilities provided.

### References

- Areechon, N. and Plump, J.A. 1990. Sub lethal effects of malathion on channel cat fish, *Ictalurus punctatus*. Bull. Environ. Contam. Toxicol., 44: 435-442.
- Baskaran, P., Palanichamy, S., Visalakshi, S. and Balasubramanian, M.P. 1989. Effects of mineral fertilizers on survival of the fish *Oreochromis mossambicus*. Environ. Ecol., 7: 463-465.
- Boran, M., Altinok, I., Capkin, E., Karacam, H. and Bicer, V. 2007. Acute toxicity of carbaryl, methiocarb, and carbosulfan to the rainbow trout *Oncorhynchus mykiss* and guppy (*Poecilia reticulata*). Turk J. Vet. Anim. Sci., 31 (1): 39-45.
- Cagauan, A.G. 1990. The impact of pesticides on rice fields vertebrates with emphasis on fish. In: P.L. Pingali and P.A. Roger (Eds.), Impact of pesticides on farmer health and the rice environment. International Rice Research Inst., Kluwer Academic Publ., Philippines: 203-248.
- Cagauan, A.G. and Arce, R.G. 1992. Overview of pesticides use in rice-fish farming in South East Asia. In: C.R. Dela cruz, C. Lightfoot, B. Coasta pierce, V.R. Carangal and M.P. Bimbao (Eds.), Rice-fish research and development in Asia. International Centre for Living Aquatic Resources Management (ICLARM) Conf. Proc., Philippines: 24: 217-233.
- Caguan, A.G., Galaites, M.C. and Fajardo, L.J. 2004. Evaluation of botanical piscicides on Nile tilapia *Oreochromis niloticus* L. and mosquito fish *Gambusia affinis* Baird and Girard. Proceedings on ISTA, 12-16 September. Manila, Phillipines: 179-187.
- Christina, A. 2004. Effects of the insecticides deltamethrin on benthic macro invertebrates – field and laboratory studies. Department of Environmental Assessment, Swedish University of Agricultural Sciences, Box 7050, SE 75007, Uppsala.
- El-Shazly, M.M. and El-Sharnoubai, E.D. 2000. Toxicity of a neem (*Azadirachta indica*) insecticide to certain aquatic organisms. Journal of the Egyptian Society of Parasitology, 30(1): 221-231.
- Finney, P.J. 1971. Probit analysis 3<sup>rd</sup> edition, Cambridge University Press, Cambridge, 333 pp.
- Golow, A.A. and Godzit, A. 1994. Acute toxicity of Deltamethrin and Dieldrin to *O. niloticus*. Bull. Environ. Contam. Toxicol., 52: 351-354.
- Göktepe, I. and Pihak, L.C. 2002. Comparative toxicity of two azadirachtin – based neem pesticides to *Daphnia pulex*. Environmental Toxicology and Chemistry, 21 (1): 31-36.
- Hodson, P.V. 1988. The effect of metal metabolism on uptake, disposition and toxicity in fish. Aquat. Toxicol., 11: 3-18.
- Istvan, U. 2000. Semi-natural products and related substances as alleged botanical pesticides. Pest Management Science, 56(8): 703-705.
- Johl, M.S. and Dua, A. 1995. Elemental lipidological and toxicological studies in *Channa punctatus* (Bloch) upon exposure to an organochlorine pesticide, endosulfan. Bull. Environ. Contam. Toxicol., 55(6): 916-921.
- Kalavathy, K., Sivakumar, A.A. and Chandran, R. 2001. Toxic effect of the pesticide Dimethoate on the fish *Sarotherodon mossambicus*. J. Ecol. Res. Bioconserv., 2(1-2): 27-32.
- Khan, M.F. and Ahmed, S.M. 2000. Toxicity of crude neem leaf extract against housefly *Musca domestica* L. adults as compared with DDVP, Dichlorvos. Turk. J. Zool., 24(4): 219-223.
- Khangarot, B.S., Ray, P.K. and Singh, K.P. 1988. Influence of copper treatment on the immune response in an air-breathing teleost *Saccobranchus fossilis*. Bull. Environ. Contam. Toxicol., 41: 222-226.
- Kharat, S., Dahanukar, N., Raut, R. and Mahabaleshwarkar, M. 2003. Long-term changes in fresh water fish species composition in North Western Ghats, Pune district. Current Science, 84(6): 816-820.
- Koesomadinata, S. 1980. Pesticide as a major constraint in integrated agriculture-aquaculture farming system. In: R.S.V. Pullin and Z.H. Shehadeh (Eds.), Integrated Agriculture - Aquaculture Farming Systems. (ICLARM) Conf. Proc., 4: 45-51.
- Lockwood, A.P.M. 1976. Effects of pollutants on aquatic organisms. Cambridge University Press, Cambridge, 193 pp.
- Miller, F. and Uetz, S. 1988. Evaluating bio rational pesticides for controlling arthropod pest and their phyto toxic effects on Green house crops. Hort. Technology, 8(2): 185-192.

- Pant, J.C. and Singh, T. 1983. Inducement of metabolic dysfunction by carbamate and organophosphorus compounds in a fish, *Puntius conchonius*. *Pestle. Biochem. Physiol.*, 20: 294-298.
- Schmutterer, H. 1990. Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. *Ann. Rev. Entomol.*, 35: 271-297.
- Sharma, S.K., Dua, V.K. and Sharma, V.P. 1995. Field studies on the repellent action of neem oil. *Southeast Asian. J. Trop. Med. Pub. Helth.*, 26: 180-182.
- Sparague, J.B. 1973. The ABC's of pollutant bioassay using fish. In: J. Cairns Jr. and K.L. Dickson (Eds.), *Biological methods for the assessment of water quality*. American Society for Testing and Materials, Special Technical Publication 528, Philadelphia: 6-30
- Svobodovo, Z., Luskova, V., Drastichova, J., Svobodova, M. and Zlabek, V. 2003. Effect of deltamethrin on haematological indices of common carp *Cyprinus carpio* L. *Acta Vet. Brno.*, 72: 79-85.
- Von westernhagen, H. 1988. Sub lethal effects of pollutants on the fish egg and larvae. In: W.S. Hoar and D.J. Randall (Eds.), *Fish physiology, the physiology of developing fish*. Academic Press, San Diego: 253-346.
- Woin, P. 1988. Short-and long term effects of Pyrethroid insecticide fenvalerate on an invertebrate pond community. *Ecotoxicology and Environmental Safety*, 41: 137-156.