

Mitochondrial Cytochrome b Sequence Variation in Three Sturgeon Species (*A. stellatus* Pallas, 1771, *A. gueldenstaedtii* Brandt, 1833, *H. huso* Linnaeus, 1758) from the Black Sea Coasts of Turkey

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E-mail: yciftci@odu.edu.tr	Accepted 15 April 2013

Abstract

It has been assumed that sturgeons originated in the Tethys Sea, and the Black Sea is their most important diversification center. Hence Turkey has had a significant place throughout history. Although there are some genetic studies about sturgeon species that inhabit the Black Sea, these studies have not included samples from Turkey. In this study, the phylogenetic relationship, cladistic positions, and genetic variations were determined from the Cyt-b (1141bp) mtDNA of 3 sturgeon species (n = 28), collected from Turkish coast of the Black Sea between 2005 and 2007. Nineteen haplotypes, and haplotype diversity ranging from 81.8% to 96.4%, were detected from Cyt-b sequences. Studied A+T rates were found between 52.6 and 53.8% for all species. Ti/Tv rates were estimated for each species: *A. gueldenstaedtii* (9:1), *A. stellatus* (7:2), and *H. huso* (6:0). Nucleotide diversity and nucleotide differences calculated for three species varied from 0.167-0.378% and 1.89-4.2, respectively. Genetic distances were calculated (1.258-5.288%). Each species was separated by phylogenetic reconstruction with a high bootstrap value (100%). All *A. gueldenstaedtii* samples replaced in the Black Sea Lineage Group and were separated into 2 clades (A and B). Similarly, *A. stellatus* samples were separated into 2 clades, but *H. huso* samples were not separated.

Keywords: Sturgeon, A. stellatus, A. gueldenstaedtii, H. huso Turkey, Black Sea, mtDNA, Cyt-b, sequence analysis, phylogenetics.

Karadeniz'in Türkiye Kıyılarında Dağılım Gösteren Üç Mersin Balığı Türünün (Acipenser stellatus, Acipenser gueldenstaedtii, Huso huso) Mitokondriyal Sitokrom b Sekans Varyasyonu

Özet

Mersin balıklarının Tethis Denizi kökenli olduğu ve farklılaşma merkezinin Karadeniz olduğu dü şünüldüğünde Türkiye çok önemli bir yere sahiptir. Karadeniz'deki mersin balığı türleri hakkında yapılmış çalışmalar bulunmasına rağmen Türkiye kıyılarından örnekleme yapılmamıştır. 2005-2007 yılları arasında Türkiye kıyılarından örneklenen 3 türe ait (*Acipenser stellatus*, *A. gueldenstaedtii*, *H. huso*) 28 örnekte Cyt-b geni (1141bp) sekans verilerinden genetik mesafeler ve filogenetik dendogramlar oluşturulmuş ve kladistik analizler yapılmıştır. Cyt-b geninde 19 haplotip ve %81,8-96,4 arasında değişen haplotip çeşitliliği belirlenmiştir. A+T oranı tüm türler için %52,6 ile 53,8 arasında bulunmuştur. Ti/Tv oranı *A. gueldenstaedtii* 'de (9:1), *A. stellatus* 'ta (7:2) ve *Huso huso* için (6:0) olarak bulunmuştur. Nükleotid çeşitlili ği ve nükleotid farklılığının sırasıyla %0,167-%0,378 ve 1,89–4,2 arasında değiştiği belirlenmiştir. Türler arası mesafe Kimura-2 parametresine göre hesaplanmıştır (%1,258–5,288). Filogenetik ağaçlar sonucunda %100'lük seç-bağla değeri ile türler ayrılmıştır. *A. gueldenstaedtii* örneklerinin hepsi Karadeniz soy grubu içerisinde yer aldığı ve 2 farklı gruba (A ve B) ayrıldığı belirlenmiştir. Benzer olarak *A. stellatus* örneklerinin de 2 gruba ayrıldığı tespit edilmiştir. Buna karşın, *H. huso* örneklerinde ayrılma görülmemiştir.

Anahtar Kelimeler: Mersin Balığı, A. stellatus, A. gueldenstaedtii, Huso huso, Türkiye, Karadeniz, mtDNA, Sitokrom b, sekans analizi, filogenetik

Introduction

Throughout history, sturgeon fish has had great value in different aspects. The high cultural,

commercial, ecological, and zoological value of sturgeons has promoted an interest in sturgeon biology including their systematics and evolution (Bemis *et al.*, 1997). More information is needed

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about the ecology, genetics, environmental physiology, toxicology, and behavior of sturgeons to understand the environmental impacts on different species. The *Acipenseriformes* order (sturgeons and paddlefish) includes 27 species in four currently recognized genera in a Holarctic distribution. This is an ancient assemblage with recognizable acipenserid fossils from the Upper Cretaceous and fossil relatives extending the origin of Acipenseriformes into the Lower Jurassic (Grande and Bemis, 1991; Findeis, 1997).

These species are mainly distributed in the water systems; such as rivers, lakes, and sea water (brackish water) of China, Central Asia, Lake Aral, the Caspian Sea, the Black Sea, and in North America (Grande and Bemis, 1991; Bemis et al., 1997). There are six sturgeon species inhabiting in the the Danube river basin, The Black Sea including the Turkish seas (Geldiay and Balik, 1996). However, due to anthropogenic impacts, Atlantic sturgeon (Acipenser sturio), Sterlet (Acipenser ruthenus) and Ship sturgeon (Acipenser nudiventris) have nowadays almost disappeared from the region, while population of Beluga (Huso huso), Russion sturgeon (Acipenser gueldenstaedtii) and stellate sturgeon (Acipenser stellatus) are experiencing severe decline (Williot et al., 2002).

Sturgeon fish are slow growing and mature very late in life that may take 20 years (depending on species) and, even then, do not spawn every year. Thus, they are particularly susceptible to exploitation and to many other risks, including pollution and habitat fragmentation. Fishermen easily catch adult sturgeon because of their large body and recognized spawning migration routes. Their dependence on shallow spawning areas and fast flowing rivers leads to the effects of exposure to environmental factors, as well (Bemis and Kynard, 1997). Today, almost all sturgeon species categorized by the International Union for Conservation of Nature (IUCN) as endangered, vulnerable, or least concern (Birstein *et al.*, 1997b).

Meristic and morphological similarities among a number of species of sturgeons living within the same basin can be defined as a certain species of Acipenser, particularly during the juvenile period. For this reason, it is impossible to rely on literature for the type of comparisons that must be made (Bemis *et al.*, 1997; Birstein et al., 1997b). Although many diverse characteristics and methods have been used to analyze the genetic structure in exploited and endangered species like sturgeons (for example, ecological parasite distribution, approaches, tagging, physiological and behavioral traits, morphometrics and meristics, calcified structures, cytogenetics, immunogenetics and blood pigments, and the use of molecular genetic techniques in fisheries), researches on the genetics of this species have increased dramatically over the last three decades, largely due to the increased availability of techniques and increased

awareness of the value of genetic data. Nowadays, there are many different types of DNA markers used in molecular genetics of sturgeon, to identify stock discrimination of commercially important species and to determine species and their origin including: (Chikhachev, 1983; Ivanenkov allozyme and Kamshilin 1991; Kuz'min, 1991; Pourkazemi, 1996; Kuz'min, 2002), DNA sequence data (DeSalle and Birstein, 1996; Birstein et al., 1998), PCR-Single Strand Confirmation Polymorphism (SSCP) (Rehbein, 1997; Ludwig et al., 2000), RAPD (Comincini et al., 1998; Gharaei et al., 2005), Species-specific PCR (DeSalle and Birstein, 1996; Birstein et al., 1998; Mugue et al., 2006), PCR-RFLPs (Wolf et al., 1999; Ludwig et al., 2002), (Congiu et al., 2001), Microsatellites AFLP (Jenneckens et al., 2001; Zhu et al., 2002; Moghim et al., 2012), and SNP (Timoshkina et al., 2011).

For identification of phylogenetic relationship among sturgeon species, mtDNA is preferred as it is the rapid rate of evolution, the maternal mode of inheritance and the relatively small size of mtDNA make the analysis of this molecule one of the methods of choice for many studies (Ferguson et al., 1995). Furthermore, the low effective size of mtDNA compared to the nuclear markers makes it more likely to provide a population-specific marker, resulting in generally greater genetic differentiation than nuclear markers (Ferguson et al., 1995). In the course of evolution, the variation ratio of the Cytb gene sequence is higher than that of other functional areas. This gene has also been one of the most frequently utilized segments of mtDNA because it is easy to align and it has been characterized in many vertebrates (Stepien and Kocher, 1997; Kocher et al., 1989; Zardoya and Doadrio, 1999; Orti et al., 1994; Brito et al., 1997). In the case of fish, the cytochrome b gene has been used to address many phylogenetic questions, from relationships among closely related species to deep phylogenetic questions such as the relationships among Acipenser gueldenstaedtii, A. baerii and A. persicus, so the Cytb gene is considered to be a good marker to research on genetic differentiation and phylogenetic relationship.

This study characterized the mtDNA variation in the 3' end of the Cyt-b gene in three sturgeon species found at the Turkish coast of the Black Sea. Genetic structure was assessed by evaluating the pattern of variation in mtDNA sequences within this selected geographic range.

This is the first molecular study based on an endangered sturgeon species from the Turkish cost of the Black Sea. Thus, it may shed light on the mode of diversification of Black Sea fauna of the area. Second, mutual monophyly at the Cyt-b level of Turkish and other Black Sea countries' sturgeons was preliminarily tested in this study. To this aim, 4 species of sturgeon from several downstream areas and seashores of the Turkish coast of Black Sea were included in the study.

Materials and Methods

Sample Collection and DNA Extraction

Between 2005-2007, on the Black Sea coast of Turkey, A. gueldenstaedtii (12), A. baerii-like (1) A. stellatus (7) and H. huso (8) species were sampled (Table 1). Sampling for sturgeon species was carried out at seven sites in the Black sea area ranging from Rize to Sakarya. Samples from the cultured Russian sturgeon whose eggs were transferred from Russia to Istanbul University were also considered. There is one A. baerii-like haplotype in the A. gueldenstaedtii samples. Although the situation of fish with this haplotype is controversial (Bristein et al., 2000), it was evaluated in this study to make comparison with other studies. A non-destructive sampling method was used in this study by taking caudal fin tissue from the fish. Caudal fine tissues about 1-1.5 cm² were preserved immediately in 98% ethanol and stored at an ambient temperature in the field and -20°C in the laboratory.

Total genomic DNA was extracted from 15-20 mg of tissue by the DNA extraction kit (PROMEGA SV Wizard kit) using the following the manufacturer's protocol or standard proteinase K, phenol/chloroform extraction (Sambrook *et al.*, 1989). The DNA concentration and quality was assessed by a UV/visible spectrophotometer (BIO-RAD, The SmartSpec Plus) at 260/280 nm. Furthermore, the extracted DNA was examined on 1% agarose gels stained with EB and stored at -20°C until needed.

PCR Amplification and Sequencing

Approximately 1.227 bp of mitochondrial segment, including the Cyt-b gene, were amplified with Cyt-f (5'-TGACTTGAAAAACCACCGTTGTTA-3') (position 14.387 – 14.410 in *Acipenser stellatus* mitochondrion, complete genome) designed for brown trout (Bernatchez and Donzman, 1993) and Cyt-r (5'-CTTCGGTTTACAAGACCG-3') primers (position 15.597-15.614 in *Acipenser stellatus* mitochondrion, complete genome) (Ludwig *et al.*, 2000). Each 50µl reaction contained 200-250 ng DNA, each primer, PCR Master Mix (2X) (Promega) (Reaction buffer, pH 8.5), and ddH₂O. The 2X PCR Master Mix (Promega), which is a premixed and

ready-to-use solution, contains Taq DNA Polymerase (50 unit/ml), dNTPs (400 µM dATP, 400µM dGTP, 400µM dCTP, 400 µM dTTP), MgCl₂ (3mM) and reaction buffers. Samples were amplified in a PTC-200 gradient thermal cycler (MJ Research, Waltham, Massachusetts, USA) using the following protocol: after a preliminary denaturation at 94°C for 1 min, each of the 35 cycles consisted of denaturation at 94°C for 1 min, annealing at 47°C for 30 s, and primer extension at 72°C for 1.5 min. To improve the subsequent reamplification and sequencing, a final cycle was added; including an extension at 72°C for 5 min. Negative controls of DNA extraction and negative PCR control were subjected to amplification. The success of the PCR reaction was visualized using 1.5-2.0% agarose gel electrophoresis with a molecular-size ladder, and stained with ethidium bromide (EB) solution to determine the size and quality of fragments under UV light. DNA bands in gels were viewed under UV light and the images were recorded.

The PCR products were purified before sequencing using Wizard® SV Gel and the PCR Clean-Up System, following the manufacturer's instruction. The purified products were then sent to MACROGEN (Macrogen Inc., Seoul, Korea) for sequencing, using both forward and reverse strands.

Nucleotide sequences of the Cyt-b segment of mtDNA were established for 28 sturgeons. Differing variants of the sequenced regions were deposited in GenBank/NCBI under accession numbers the KC130090-KC130117. The following Cyt-b gene sequences from the related sturgeon species were retrieved from the GenBank and used in the present study (AF238688, AF238694, AF238669, AJ249692, AF238699, AJ563386, AF238679. AJ277594, AJ563395, AJ563394. AF238700, AF238705, (AF238620. AJ277596) Α. gueldenstaedtii, A. persicus, (AJ245833, AJ245834, AF238615) AF238660) A. naccarii (AF238658, AF238653, AF238631, AF238640, 11102385) Α. baerii. (AJ249693, AY846685, AY846696, AY846689) A. stellatus, (AJ245828) A. brevestrum, (AJ245832) A. nudiventris, (AJ249694) A. ruthenus, (AY510085) A. dabryanus, (AJ245830) A. medirostris, (AJ245839) A. sturio, (AJ245840) H. huso, (AJ252187) H. dauricus, (NC005834) Psephurus (NC004419) gladius, Polyodon spathula. For comparative analysis, 37 Cyt-

Table 1. Sampling location of sturgeon species used in these studies [number of sample (sampling no)]

Samling locations	A. gueldenstaedtii	A. baerii/like	A. stellatus	H. huso	Total
Rize	-	-	-	1 (25)	1
Trabzon	-	-	3 (26-27-28)	3 (20-29-69)	6
Giresun	4 (40-44-53-54)	1 (46)	2 (48-60)	3 (18-56-59)	10
Samsun	5 (37-61-67-91-62)	-	2 (32-66)	1 (76)	8
Sakarya	1 (63)	-	-	-	1
Hatchery orginated samples from Russia	2 (2-11)	-	-	-	2
Total	12	1	7	8	28

b sequences from the GenBank were combined with the sequences of the present study and used to infer the phylogenetic relationship of sturgeons in the Turkish coast of the Black Sea. For the analysis, one representative of a closely related species, *Polyodon spathula*, was used as the outgroup.

The genetic structure was assessed by evaluating the pattern of variation in mtDNA sequences within this selected geographic range.

Statistical Analysis

Genetic differences among sturgeons and proximal matrix relationships were tested using the statistical packages BIOEDIT (Hall, 1999), DnaSP 4.0 (Rozas and Rozas, 1999), MEGA 3.0 (Kumar et al., 2004), Arlequin (Schneider et al., 2000), and PAUP (Swofford, 2003). Nucleotide sequences from all specimens were compared and aligned among themselves and in respect to other sequences present in databases using the ClustalW computer software package (EMBL, Heidelberg, Germany). Minimal visual adjustments were made to the completed alignments using MEGA 3.0 (Kumar et al., 2004) and the BioEdit Sequence Alignment Editor (Hall, 1999). The protein sequence was predicted from the Cyt-b gene sequences using the BioEdit Sequence Alignment Editor (Hall, 1999). Haplotype numbers and haplotype diversity (h), nucleotide diversity (Pi, π) and nucleotide differences (Tajima, 1983) were computed using DnaSP 4.0 (Rozas and Rozas, 1999) software.

All matrices were analyzed with two approaches: Bayesian inference and parsimony. Bayesian analyses were performed in MrBayes 3.1 (Huelsenbeck and Ronquist, 2001; Ronquist and Huelsenbeck, 2003). For each single-gene dataset, the best-performing evolutionary model was identified under two different model selection criteria: Akaike information criterion (AIC) (Akaike, 1974) and the Bayesian information criterion (BIC) (Schwartz, 1978). These calculations were made in the Modeltest 3.7 (Posada and Crandall, 1998) and MrModeltest v.2.2 (Nylander, 2004) software using the graphical interface MrMTgui (1.0) (Nuin, 2008) and PAUP* ver. 4.0b10 for Windows (Swofford, 2003; Nylander, 2004). Phylogenetic trees were constructed using maximum parsimony (MP; minimum heuristic search factor of 2), neighbor-joining (NJ) and the unweighted pair group method (UPGMA), by MEGA version 3 (Kumar et al., 2004) based on Kimura twoparameter distances (Kimura, 1980). The robustness of the inferred UPGMA, MP and ML trees were bootstrapping with tested by 1000 pseudo replications. These trees were determined by applying the 50% majority rule (strict consensus) for bootstrap replicates. In order to investigate the genetic differentiation among the three sturgeon species, an analysis of the pairwise Kimura-2 parameter distances between species was computed using the Arlequin 3.1 software (Schneider et al., 2000).

Results

Alignment and Base Composition of the Cyt-b Gene

The 1141 bp region of the 3' end of the mtDNA Cyt-b gene was aligned for 28 specimens; 19 haplotypes were identified with 100 segregating sites including 7 single variable sites and 93 parsimony informative sites (Figure 1). Haplotypes indicated minimal homoplasy because no more than two different nucleotides were identified at each segregating site. Haplotypes were not shared between species and were found to be specific to the species. While five haplotypes (H1, H2, H3, H4, H5) were observed in A. gueldenstaedtii (n = 12), only one haplotype (H6) was shared with one specimen of A. baerii, similar to the one from Giresun. Six (H7, H8, H9, H10, H11 and H12) and 7 haplotypes (H13, H14, H15, H16, H17, H18 and H19) were found in A.stellatus (n=7) and H. huso (n=8), respectively. Frequency distribution of haplotypes within and between species is given in Figure 1. H2 was the most frequent haplotype (33.3%) identified in A. gueldenstaedtii and shared by 4 specimens.

For all the sequences of three sturgeon species, the average nucleotide frequencies of thymine (T), cytosine (C), adenine (A), and guanine (G) were 26.6%, 31.9%, 26.5%, and 15.2%, respectively, and varied between 14.9 and 32.0%. The content of A+T (53.1%) was higher than that of G+C (46.9%) (Table 2). At the third position, strong compositional biases against G (only 7.5%) were observed. At all levels of sequence divergence, the ratio of unambiguous transitions to transversions in Cyt-b were estimated for each species; A. gueldenstaedtii (9:1), A. stellatus (7:2), and H. huso (6:0) and Ti's outnumbered Tv's, even in comparisons between the ingroup and the outgroup (Table 2). Meanwhile, the ratio of transitions/transversions (R) was 7.3. There was no significant difference in base compositional bias between ingroup and outgroup species. A total of twenty-five intraspecific variable sites were found among the 1141 bp of the Cyt-b gene in the 28 cases examined. Nucleotide diversity (Pit) and nucleotide differences (k) calculated for three species varied from 0.167% to 0.378% and from 1.89 to 4.2, respectively. Most of the mutation events were transitions (88%, including 63.6% CT and 36.4% AG). Transversions were mostly AT (66.7%) and AC (33.3%).

Genetic Distances

Pairwise genetic distances among and within three sturgeon species and the outgroup, *Polyodon spathula*, were estimated by MEGA (Kumar *et al.*, 2004) using the K2P method with gamma correction

	Nucleotide position			
Haplotype	11111111111 11111111112222222223333333344444444455566666666666	A.g A.b A.s H.h		
Hap_1	${\tt tacgcatctacgtactttcactacttcgaaaggccccctccct$	3		
Hap_2	TCAT.TT	4		
Hap_3	T	3		
Hap_4	T	1		
Hap_5	т.	1		
Hap_6		1		
Hap_7	CGTATCG.CTCCA.GATTCTCCCCATAT.TTT.TCC.CGCGAG.GTG.ATGCG.AG.CA.GC.G.	1		
Hap_8	CGTA.TCTCG.CTCCAATTCTCCCCATAT.TTT.TCC.C.CGAG.GTG.ATGCG.AG.CA.GC.GC	1		
Hap_9	CGTA.TCTCG.CTCCAATTCTCCCCATTAT.TTT.TCC.C.CGAG.GTG.ATGCG.AG.CA.GC.GC	1		
Hap_10	CGTACG.CTCCA.GATTCTCCCCATAT.TTT.TCC.C.CGAG.GTG.ATGCG.AG.CA.GC.GC	1		
Hap_11	CGTA.TCTCG.CTCCA.GATTCTCCCCATAT.TTT.TCC.C.CGG.GTG.ATGCG.AG.CA.GC.GC	1		
Hap_12	CGTA.TCTCG.CTCCA.GATTCTCCCCATAT.TTT.TCC.C.CGAG.GTG.ATGCG.AG.CA.GC.GC	2		
Hap_13	ATTCGC.T.CGTC.TCCTAGAATTTCT.C.CA.A.TAACTTGT.TTT.CGATGGGGAGAAGCT	1		
Hap_14	ATTCGC.T.CGTC.TCCTAG.GAATTTCT.C.CA.A.TAACTTGT.TTT.CGATGGGGAGAAAGCT	1		
Hap_15	ATTCGC.T.CC.GTC.TCCTAG.GAATTTCT.C.CA.A.TAACTTGT.TTT.CGATGGGGAGAAGCT	2		
Hap_16	ATTCGC.T.CGTC.TCCTAG.GAATTTCT.C.CA.A.TAACT.GT.TTT.CGATGGGGAGAAGCT	1		
Hap_17	ATTCGC.T.CGTC.TCCTAG.GAATTTCT.C.CA.A.TAACTTGT.TTT.C.ATGGGGAGAA.GGCT	1		
Hap_18	ATTCGC.T.CGTC.TCCTAG.GAATTTCT.C.CA.A.TAACTTGT.TTT.CGATGGGGAGAAGCT	1		
Hap_19	ATTCGC.T.CGTC.TCCTAG.GAATTTCT.C.CA.A.TAACTTGT.TTT.CGATGGGGAGAA.GGCT	1		

Figure 1. Variable nucleotide sites in haplotypes of the mtDNA cytochrome b gene and haplotypes frequecies. Numbers above the sequence refer to variable nucleotide positions (Acc. No: KC130090–KC130117). Only variable nucleotide positions are shown, and dots represent identity with the first sequence.

Table 2. Frequency of base composition in 4 sturgeon species for 1141 bp of cyt b sequence, the ratio of transitions/transversions (R), Nucleotide diversity (Pit) and nucleotide differences (k). A-T percentages are also shown

Species	Т	С	А	G	%AT	Ti:Tv	Pit	k
A. gueldenstaedtii	26.9	32.0	26.3	15.0	53.2	9:1	0.00378	4.212
A.baerii/like	27.0	31.4	26.8	14.9	53.8	-	-	-
A. stellatus	26.3	32.0	26.3	15.4	52.6	7:2	0.00299	3.333
H. huso	26.3	32.0	26.3	15.4	52.6	6:0	0.00167	1.857
Average	26.6	31.9	26.5	15.2	53.1			

(Table 3). The genetic distance was lowest between *A.baerii*-like - *A. gueldenstaedtii* (1.63%) and highest between *A. stellatus* - *H. huso* (27.9%). The intraspecies sequence divergence ranged from 0.42% (*A. gueldenstaedtii*) to 0.17% (*H. huso*).

Phylogenetic Analysis

Prior to the maximum likelihood phylogenetic analysis, the MODELTEST version 3.7 (Posada and Crandall, 1998) was used in conjunction with PAUP* 4.0 beta 10 (Swofford, 2003) in order to determine the appropriate substitution model of DNA evolution that best fit the dataset. The combined Cyt-b data set included 28 sequences that belong to three species of the family *Acipenseridae*. For the analysis, one representative of closely related to Acipenserid *Polyodon spathula* was used as the outgroup. Among 56 different models, a best-fit model TrN+G ((Tamura and Nei, 1993) + gamma distribution), selected by both AIC and BIC in the Modeltest Version 3.7, are as follows: base frequencies 0.2733 (A); 0.3279 (C); 0.1402 (G); and 0.2585 (T), proportion of invariable sites = 0.000, and shape parameter (alpha) = 0.0544. A small alpha value indicated that most of the sites evolve very slowly. The transition/transvertion ratio (27.4749) also showed that there was a bias towards transition. At this point, no bias was found between base frequencies of the cases after checking the sequence with the PAUP* program ($\chi 2 = 7$, sd = 81, P=1.00).

molecular phylogenetic trees were The constructed based on the mtDNA Cyt-b gene sequences of three sturgeon species and Polyodon spathula was selected as an outgroup to determine the root of the tree. The dataset was analyzed with the Bayesian inference algorithm (MrBayes; Huelsenbeck and Ronquist, 2001), and UPGMA (Unweighted Pair Group Method with Arithmetic Means), MP (Maximum-parsimony), and ML (Maximumlikelihood) algorithms in PAUP* 4.0 beta 10 (Swofford, 2003) to resolve the phylogenetic relationships. The robustness of the tree was corroborated with bootstrap analyses (Bootstrap value =1.000). All phylogenetic trees obtained are widely based on the same topology with A. gueldenstaedtii and *A.baerii*-like *A. stellatus*, and *H. huso* appearing as monophyletic clades. Bootstrap support ranges from 50 to 100% under all tree-building methods. Bootstrap proportions are given for each method of analysis and shown for each node.

The results showed that the Black Sea sturgeons from the Turkish coast were grouped into two assemblages: *Acipenser* and *Huso*. The *Acipenser* group is represented by the species *A. gueldenstaedtii*, *A.baerii*-like and *A. stellatus*, and forms a separate branch in the UPGMA, MP and ML trees.

Results of the phylogenetic trees for each species shown in Figure 2 indicated the presence of a clear grouping within the Black Sea sturgeon species

(A. gueldenstaedtii) supported by high bootstrap values (60 - 96%). Group I could be clearly divided into two clusters. The first included the hatcheryoriginated individuals (2 and 11) grouped within the individuals from the Samsun samples (61and 62), and the second contained the other three (40, 44, 54) from Giresun grouped with one of the Sakarya samples (63). Group II contains four cases (37, 53, 67 and 91), three of which (37, 67 and 91) were from Samsun, and one of which (53) was from Giresun. A sample 46 completely separated from number *A*. gueldenstaedtii samples in dendograms were found closer to the A.baerii-like sample according to the results of the BLASTn, in which a nucleotide

Table 3. Kimura 2-parameter pairwise distances (gamma corrected) based on cytochrome-*b* sequence data (outgroup was included). Italic values are intra- species distances (α : 0.05)

Species	А	В	С	D	Е	
A. gueldenstaedtii (A)	0.0042					
A. stellatus (B)	0.1722	0.0032				
H. huso (C)	0.1782	0.2793	0.0017			
A.baerii/like (D)	0.0163	0.1700	0.1259	-		
P. spathula (E)	4.3372	4.8568	4.6423	4.3696	-	



Figure 2. Phylogenetic tree of the genus *Acipenser* and *Huso* based on mtDNA cytochrome b gene sequences. Numbers on branches represent the bootstrap values obtained for 1000 replications corresponding to the UPGMA (the unweighted pair group method), MP (maximum parsimony) and ML (maximum likelihood) methods. Support values under 50% are not represented and given order of UPGMA/MP/ML. Identification numbers of individuals are followed by sampling sites and shared haplotypes.

sequence is compared to the contents of a nucleotide sequence database. The sequences of homologous mtDNA regions were used, sequenced by other authors, and deposited in the GenBank/NCBI under accession numbers given in the methods section. Since all sequences were used in the comparative analysis, Group II samples were separated from GenBank samples belonging to the Black Sea and the adjacent area, but Group I together and two subgroups were included in the GenBank samples (Figure 3). The strong phylogenetic split into two groups corresponds to the main geographic areas. Group II seemed to be almost exclusive to the Turkish Black Sea waters while Group I might be entirely exclusive to other areas in the Black Sea. All trees (UPGMA, MP and ML trees) revealed that the *A. stellatus* was composed of two clusters. One of them was supported (>50% bootstrap) and contained representatives of cases 26 and 32. The second was a larger assemblage that consists of five individuals (27, 28, 48, 60 and 66). When analyzing the trees for *H. huso*, such a separation was not seen within the species. Samples



Figure 3. UPGMA (Unweighted Pair Group Method with Arithmetic Mean) dendrogram based on Kimura two-parameter distances (Kimura, 1980), showing relationship between related sturgeon species retrieved from GenBank and three Black sea species used in the present study, using Cytochrome b sequences. The robustness of the inferred UPGMA tree was tested by bootstrapping with 1000 pseudoreplications.

(20 and 25; 29 and 59) were closer to each other but were not separated from the others.

Discussion

Characteristics of the Cyt-b Sequences

There are phylogenetic and population studies regarding the Eurasia species in the Black Sea, such as *Acipenser gueldenstaedtii*, *A. stellatus* and the *H. huso* species together with other sturgeon species (Ludwig *et al.*, 2000; Bristein *et al.*, 1998a, b; Doukakis, 2000; Jennecknes *et al.*, 2000; Stabile *et al.*, 1996; Brown *et al.*, 1992). On the other hand, there have been no studies conducted thus far regarding the sturgeon species in Turkey based on either molecular or morphometric analysis. This study compared samples obtained from the Black Sea coast of Turkey both within the samples and with other studies by means of a sequence analysis of the mtDNA Cyt-b gene.

Approximately 1227 base pair (bp) fragments of the mitochondrial Cyt-b were amplified and then sequenced from 28 samples belonging to 4 species: *A.* gueldenstaedtii (12), *A. baerii*/like (1), *A. stellatus* (7) and *H. huso* (8). Complete sequences of 1141 bp Cytb were obtained after correction and alignment. Among the 28 sequences of the 4 sturgeon species, 19 distinct haplotypes were detected: 5 in *A.* gueldenstaedtii; 1 in *A.baerii*-like; 6 in *A. stellatus*; and 7 in *H. huso*. In general, one dominant haplotype (H2) was shared by 4 specimens in *A. gueldenstaedtii*. All haplotypes were unique to their own species.

Haplotype diversity in bony fish is over 80% (Grant and Bowen, 1998). Similar results were also obtained from other studies conducted with sturgeon species (Stabile *et al.*, 1996; Birstein and Doukakis, 2000; Doukakis *et al.*, 2005). In this study, high haplotype diversity, between 81.8% - 96.4% for each of the three species, was observed. Despite the low number of samples for the species studied, the results obtained from this study are the same as other studies.

Furthermore, in this study, one *A. baerii*-like haplotype (Hap6) was observed within the samples obtained from the Black Sea and classified morphologically as *A. gueldenstaedtii*. Similarly, in the studies conducted on the Volga River (Jennecknes *et al.*, 2000), the Sea of Azov (Voynova *et al.*, 2008) and the Eurasian species (Doukakis, 2000), *A. baerii*-like mitotypes were observed within the samples of *A. gueldenstaedtii* species. According to Ludwig *et al.* (2002), natural sturgeon hybrids are often rare. However, possible reason for hybrids results could be intended release programs, habitat alterations or unintended escapes from hatcheries.

Different researchers indicated that Cyt-b (53-57%) and D-loop (70-80%) regions have a high A+T nucleotide content (Apostolidis *et al.*, 1997), and an A+T nucleotide content with a 68% ratio is a general value accepted for a control region (Meyer, 1993). The current study revealed that the A+T nucleotide ratio is 52.6% for *A. stellatus* and *H. huso*, while it is 53.2% and 53.8% for *A. gueldenstaedtii* and *A.baerii*-like, respectively, suggesting that average level of genetic diversity for studied species.

In previous studies, the Cyt-b gene was found to be rich in C, A, and T nucleotide ratios, but low in G content (Cantatore *et al.*, 1994; Duokakis, 2000; Almodovar *et al.*, 2000). The nucleotide frequencies of the Cyt-b gene were shown for different living groups by Johns and Avise (1998). When compared with the values given in the Table 4, the Cyt-b gene region was found to be rich in the C ratio (32%) but low in the G ratio (15%) in the current study.

The mean base composition in Cyt-b sequences was similar to those previously reported for Actinopterygian fish and the other mitochondrial protein-coding genes (Johns and Avise, 1998) with low G content (15.2%). A high A+T bias may be a common phenomenon in fish and an A+T nucleotide bias, when present, tends to accumulate in hyper variable sites (Simon, 1991).

Nucleotide diversity was calculated as 0.378%, 0.299% and 0.167% for *A. gueldenstaedtii*, *A. stellatus* and *H. huso*, respectively. Nucleotide diversity values calculated for the studied gene region was found to be different between species and generally in agreement with the values observed in other fish.

Phylogenetic Analysis

In contrast to the known phylogenetic positions of Acipenseriforms, the phylogenetic relationships of the species in this order are still controversial (Mayden and Kuhajda, 1996; Birstein *et al.*, 1997a; Findeis, 1997; Birstein and DeSalle, 1998; Ludwig *et al.*, 2000).

In UPGMA, MP and ML dendrograms obtained as a result of the phylogenetic analysis prepared for *A. gueldenstaedtii* samples, it was observed that this species is divided into two groups as A (A1-A2) - B. When all *A. gueldenstaedtii* samples are compared with the samples taken from the GenBank, the samples in the "B" group are separated from the samples both in the "A" group (A1 and A2) and from the GenBank. On the other hand, the samples in the

Table 4. Nucleotide frequencies in the Cyt-b gene for different animals (modified from Johns and Avise, 1998)

Animals	Т	С	А	G	A+T
Birds	25.0	33.9	27.6	13.5	52.6
Mammals	28.8	28.4	29.6	13.3	58.4
Reptiles	28.6	27.6	27.7	16.1	56.3
Amphibians	32.7	24.3	27.0	16.0	59.7
Fishes	29.5	29.9	25.3	15.3	54.8
Sturgeons (current study)	26.83	31.45	26.52	15.18	53.36

"A" group are not separated from the samples taken from the GenBank. In the study conducted by Birstein et al. (2000), based on the phylogenetic analysis of sequences of three mitochondrial gene regions (Cyt-b, ND5 and D-loop), A. gueldenstaedtii was separated into two main groups as NPG (A. naccarii, A. persicus, and A. gueldenstaedtii) and BG (A. baerii and A. gueldenstaedtii). Similar to this study, according to Almodovor et al. (2000), A. gueldenstaedtii is separated into two distinct genetic forms which are "typical" and "cryptic" (hidden) forms. The typical form phylogenetically combines A. persicus with A. gueldenstaedtii and A. naccarii. As a result of this data, it was specified that A. persicus may be a subspecies of A. gueldenstaedtii rather than a separate species. While the cryptic form of A. gueldenstaedtii is genetically very close to A. baerii, it is separated from A. baerii in 432th position of control region with C-T transition. When two mitochondrial lineage groups within Α. gueldenstaedtii are considered, all A. gueldenstaedtii samples that were studied are within the group (NPG) that can called "The Black Sea Lineage Group". The "B" group, included in the Black Sea Lineage Group, within A. gueldenstaedtii samples and represented by the samples on the shores of Turkey, should be finalized by working with more samples.

As seen in either the generated sequence data or the dendograms obtained as a result of phylogenetic analysis drawn by the data taken from the GenBank (Figure 3), A. gueldenstaedtii and A. baerii species are relative to A. brevirostrum and are close to each other. Based on the classification conducted by Almodovor (2000), North American shortnose sturgeon (A. brevirostrum), generally found on the Atlantic coast, was observed to be closely related to the Russian sturgeon (A. gueldenstaedtii), which is found in the Black Sea - Caspian (Eurasia) region (Vlasenko et al., 1989), and the Siberian sturgeon (A. baerii), which is found in the Siberian rivers and the Lake Baikal (Ruban et al., 1997). These results support the phylogenetic relationship obtained by Bristein and Desalle (1998). On the other hand, Bristein and Desalle (1998) compared the 650 bp region of the Cyt-b gene and used the 150 bp of 12S rDNA combined with the 350 bp of 16S rDNA. Furthermore, in the study conducted by Bristein and Desalle (1998), despite the use of a longer Cyt-b region in contrast to the data of other researchers, A. naccarii, A. persicus, A. gueldenstaedtii, A. baerii and A. brevirostrum species were found to be of a separate lineage.

The specimen that was indicated as *A. gueldenstaedtii* in samplings, but was closer to *A. baerii* according to sequence results, was found to be separated from *A. gueldenstaedtii* as a result of the comparison with *A. gueldenstaedtii* samples. The presence of *A. baerii* in different river basins, which are indeed not a natural distribution area of this species, was indicated by Jenneckens *et al.* (2000). On

the other hand, in the study conducted by Jenneckens *et al.* (2000), it was reported that the haplotype belonging to this species was observed within *A. gueldenstaedtii* samples obtained from the Volga River, but not observed in the Sea of Azov and the South Caspian Sea. In contrast to these results, the results obtained in this study indicate this sample was used in the study conducted by Jenneckens *et al.* (2000) and supports the doubts of it being of Siberian sturgeon with a haplotype shown as *A. baerii* haplotype. The most likely reason of this result is genetic contamination of *A. gueldenstaedtii* with *A. baerii* or *A. baerii* hybrids.

A. stellatus samples were separated as a species based on the topologies obtained with a 100% bootstrap value. In addition, when compared to the samples taken from the GenBank, it was found to be separated into its own (Clade 1 and Clade 2) and closer to A. nudiventris. When the studies were considered in order to determine the differences between intra-species, no distinction between species at subspecies level has been identified in the protein study conducted by Kuz'min (1994). Similarly, in the study conducted on D-loop and ND5 genes performed by Doukakis (2000), a low GST value calculated for A. stellatus indicated that the subspecies determined for the Ural River, the Volga River, the Caspian Sea, the Black Sea, or the Sea of Azov are the same genetically.

In light of these studies, stocking was believed to be the main factor for the intra-species embodiment of *A. stellatus*. The ongoing stocking studies ensured the continuity of populations in the Caspian, the Azov, and the Black Seas (Doukakis, 2000). Yet it was unknown whether this process had any effect on the genetic structure due to the lack of genetic studies conducted prior to these studies. This effect can be only determined by studying museum samples (outdated).

Despite the H. huso species belonging to the Huso genus, it was found to be separated from the studied samples with a 100% high bootstrap value, and when compared to the sequence data obtained from the GenBank, it was considered closely related to A. ruthenus (Figure 3). Similarly, in previous molecular studies (Bristein et al., 1997a; Bristein and Desalle 1998; Doukakis, 2000), H. huso was separated and considered as a monophyletic lineage, but it is within the Acipenser genus and closely related to A. ruthenus. H. huso was thought as a subgenus of Acipenser by most nineteenth-century systematics and the genus status related to its late evolution is still controversial since then (Artyukhin, 1995; Bemis et al., 1997). In contrast to these results, while Huso is the basis for all of other Acipenseriforms according to Findeis (1997), it is sister taxa within Acipenser according to Artyukhin (1995) and Mayden and Kuhajda (1996). Artyukhin (1995) emphasized that Huso may be within the Acipenser genus and is closely related to A. ruthenus,

A. nudivetris and *A. shcrenckii*. In addition, although *H. huso* and *A. ruthenus* species have different life stories and show large variations in their morphological aspects, they were found to be closely related due to their ability to produce fertile offspring by means of easy hybridization (Bristein and DeSalle, 1998; Bristein *et al.*, 1997b).

Divergence Times and Evolutionary Scenario

The possibility of inferring divergence times more accurately has been promoted by the idea that the accumulation of genetic change between lineages can be used as a molecular clock (Zuckerkandl and Pauling, 1965). Molecular sequence data allow an estimation of distances only. Under the assumption of a constant rate over time, the distance is a linear function of time. To convert distances into absolute times, an external time (called a calibration point) is used, and obtained from fossil data or geological events that mark the separation of the species. From a biological perspective, sturgeons are one of the oldest and most primitive existing Osteichthyan fishes (Ludwig, 2006). Acipenseriformes have existed since at least the Lower Jurassic period (approximately 200 MYBP), and all fossil and recent taxa arose in the Holarctic period. Phylogenetic relationships among Paleozoic and Early Mesozoic actinopterygians are problematic, but most researchers agree that Acipenseriformes are monophyletic and derived from some component of 'paleonisciform' fishes (Bemis et al., 1997). In order to estimate the divergence times among species from the mtDNA data set, published fossil data were used to calibrate the ages of the nodes on the ML tree. A number of previous clock studies have suggested that the estimated time for splitting between sturgeons and paddlefishes is 135-150 mya (Birstein et al., 1998a). Therefore, 135-150 mya was used as the range of dates for the C1 calibration node.

All available paleontological information indicates that Acipenseriforms originated in the Tethys basin (Birstein et al., 1997a). The geological events during the formation of the Tethys region are among the main factors playing a role in both the diversity of many species and the distribution of sturgeon fishes. The Tethys Sea sank, and the Black Sea, the Caspian Sea and the Aral Sea were formed approximately 120 mya, and as a result of these events, the sturgeon lineage originated (171 mya). Therefore, ongoing separation resulted in the North American A. oxyrinchus, and A. sturio from Europe. However, this scenario completely eliminates the other possibilities. Other reasons such as increased salinity in waters between continents should be considered. Another example for the distribution of sturgeon fish in the sea is the Vicariance Scenario, observed due to continental drift. The opening of the Atlantic Ocean between Europe and North America formed about 100 mya, and the difference between the two main lineages of sea sturgeon supports this event. Sea sturgeons are the anadromous fish, well adapted to the marine environment, and seen on both sides of the Atlantic Ocean. The adaptation of sturgeons to the marine environment is probably a primitive feature for these groups of fish. The loss or reduction in the ability to enter the marine environment is observed, depending on the formation of ecologically suitable freshwater locations.

Six different species were used for calibration, and the status of polyploidy was taken into account in the study conducted by Peng *et al.* (2007). In this way, the differentiation time of *H. huso* and *A. stellatus* species from *A. gueldenstaedtii* was calculated as 86.4 mya (48.2-132.6) and 59.7 mya (29.4-99.8), respectively. These values support the results obtained in this study. In other words, the distance from *A. gueldenstaedtii* to *H. huso* was calculated as 7.15-58.8 mya, while it was calculated as 6.89-58.1 mya from *A. gueldenstaedtii* to *A. stellatus* in this study.

According to previous studies on the molecular phylogenetic of sturgeon fish, two different mtDNA lineage groups in *A. gueldenstaedtii* complex and *A. baerii*, *A. persicus* and *A. naccarii* originated from one monophyletic. They are relatively younger and *A. gueldenstaedtii* is a more fundamental taxa. Two different mtDNA lineage groups in *A. gueldenstaedtii* indicate that the other three species may have originated from the populations of this species (Bristein and DeSalle, 1998).

Geological factors have a definite role in the structure of A. baerii intra-species. According to Berg (1928) in Doukakis (2000), ancestral A. baerii which belong to the BG group of A. gueldenstaedtii may have migrated from the Black Sea-Caspian region to the Siberian rivers and lakes system during the great ice age or by means of post-glacial melt (flood) (Duokakis, 2000). The connection of the lakes with the Siberian rivers occurred approximately 10,000-15,000 years ago (Arkhipov et al., 1998). It has been reported that the sea and the basins of Siberian Rivers were connected by way of a lake system during the last glacial period (Arkhipov et al., 1998). The last connection occurred approximately 18,000-20,000 years ago (Laukhin, 1997). From this relatively new connection to the present, there has not been enough time for change so that the nucleotide changes can be stabilized into different subspecies of A. baerii (Doukakis, 2000). In contrast to these geographic structures, according to the evaluation of the molecular clock indicated by Peng et al. (2007), A. baerii (24 mya) and A. naccarii (5.3 mya) appear to have diverged from A. gueldenstaedtii earlier than this period. These values support the results regarding the time of separation of A. baerii (0.5-12 mya) obtained in this study.

According to assessments of the researchers, the two species (*A. naccari* and *A. baerii*) originated from different genetic forms of *A. gueldenstaedtii* at approximately the same time. The differences in

genetic and morphological characters between species are very small due to the proximity of the past historical process.

Acknowledgments

This study was supported by Ministry of Agriculture and Rural Affairs, General Directorate of Agricultural Research (TAGEM), Project No: TAGEM/HAYSÜD/2006/. The authors would like to thank TAGEM for their financial support. The authors also wish to thank dearly departed Prof. Ibrahim OKUMUS for his scientific contributions and whose support was continuously felt by the authors.

References

- Akaike, H. 1974. A New Look at The Statistical Model Identification. IEEE Trans Autom Contr., 19, 716– 723. doi:10.1109/TAC.1974.1100705
- Almodovar, A. Machordom, A. and Suarez, J. 2000. Preliminary Results from Characterization of The Iberian Peninsula Sturgeon Based on Analysis of the mtDNA Cytochrome b. Boletín Instituto Español De Oceanografia, 16: 17–27.
- Apostolidis, A.P. Triantaphyllidis, C., Kouvatsi, A. and Economidis, P. S. 1997. Mitochondrial DNA Sequence Variation and Phylogeography Among *Salmo truta* L. (Greek Brown Trout) Populations. Mol. Ecol., 6: 531-542. doi: 10.1046/j.1365-294X.1997.d01-176.x
- Arkhipov, S. A., 1998. Stratigraphy and Palaeogeography of the Sartan glaciation in west Siberia, Quat. Int., 45, 29–42. doi:10.1016/S1040-6182(97)00004-9
- Artyukhin, E.N. 1995. On The Biogeography and Relationships within the Genus Acipenser. Sturg. Quart, 3: 6-8.
- Bemis, W.E. and Kynard, B. 1997. Sturgeon Rivers: An Introduction to Acipenseriform Biogeography and Life History. Env. Biol. Fish, 48: 167-183. doi: 10.1007/0-306-46854-9 8
- Bemis, W. E., Findeis, E. K. and Grande, L. 1997. An overview of Acipenseriformes. Env. Biol. Fish., 48, 25-71. doi: 10.1023/A:1007370213924
- Bernatchez, L. and Danzmann, R.G. 1993. Congruence in Control region Sequence and Restriction-Site Variation in Mitochondrial DNA of Brook Charr (*Salvelinus fontinalis* Mitchill). Mol. Biol. Evol., 10: 1002-1014.
- Birstein, V. J., Hanner, R. And Desalle, R. 1997a. Phylogeny of the Acipenseriformes: cytogenetic and moleculer approaches. In: Sturgeon Biodiversity and Conservation. (V.J. Birstein, J.R. Waldman and W Bemis eds.). Kluwer Academic Publisihers, Dordrecht, 127-155.
- Birstein, V. J., Bemis, W. E. and Waldman, J. R. 1997b. The Threatened Status of Acipenseriform Species: A Summary. Env. Biol. Fish, 48: 427-435. doi: 10.1023/A:1007382724251
- Birstein, V. J. and Desalle, R. 1998. Molecular Phylogeny of Acipenserinae. Mol. Phylogen. Evol, 9: 141-155. doi: 10.1006/mpev.1997.0443
- Birstein, V. J., Betts, J. and Desalle, R. 1998a. Molecular Identification of *Acipenser sturio* Specimens: A Warning Note for Recovery Plans. Biol. Conserv, 84:

97-101. doi: 10.1016/S0006-3207(97)00091-8

- Birstein, V. J., Doukakis, P., Sorkin, B. and Desalle, R. 1998b. Population Aggregation Analysis of Three Caviar-Producing Species of Sturgeons and Implications for the Species Identification of Black Caviar. Cons. Biol, 12: 766-775. doi: 10.1111/j.1523-1739.1998.97081.x
- Birstein, V. J. and Doukakis, P. 2000. Molecular analysis of Acipenser sturio L., 1758 and Acipenser oxyrinchus Mitchill, 1815: A review. Bol. Inst. Esp. Oceanogr. 16(1-4): 61-73.
- Birstein, V. J., Doukakis, P. and Desalle, R. 2000. Polyphyly of mtDNA lineages in the Russian sturgeon, *Acipenser gueldenstaedtii*: Forensic and Evolutionary implications. Cons. Gen., 1: 81-88. doi: 10.1023/A:1010141906100
- Brito, R.M., Briolay, J. Galtier, N. Bouvet, Y., Coelho, M.M. 1997. Phylogenetic Relationships within Genus*Leuciscus* (Pisces, Cyprinidae) in Portuguese Fresh Waters, Based on Mitochondrial DNA Cytochromeb Sequences. Molecular Phylogenetics and Evolution, 8(3): 435–442. doi: 10.1006/mpev.1997.0429.
- Brown, J. R., Beckenbach, A. T. and Smith, M. J. 1992. Influence of Pleistocene Glaciations and Human Intervention Upon Mitochondrial DNA Diversity in White Sturgeon (*Acipenser transmontanus*) Populations. Can. J. Fish. Aquat. Sci, 49: 358-367. doi: 10.1139/f92-041
- Cantatore, P., Roberti, M., Pesole, G., Ludovico, A., Milella, F., Gadaleta M. N. and Saccone, C. 1994. Evolutionary Analysis of Cytochrome b Sequences in Some Perciformes: Evidence for a Slower Rate of Evolution than in Mammals. J. Mol. Evol, 39: 589-597. doi: 10.1007/BF00160404
- Chikhachev, A. S. 1983. Monitoring the genetic structure of populations and hybridizations of valuable fish species in artificial culture. In Biological fundamentals of fish culture: problems of genetics and selection. Leningrad, Nauka, 91-102 pp.
- Comincini, S., Lanfredi, M., Rossi, R. and Fontana, F. 1998. Use of RAPD markers to determine the genetic relationships among sturgeons (Acipenseridae, Pisces). Fisheries Science 64: 35-38.
- Congiu, L., Dupanloup, I., Patarnello, T., 2001. Identification of interspecific hybrids by amplified fragment length polymorphism: the case of sturgeon. Molecular Ecology, 10: 2355-2359. doi:10.1046/j.0962-1083.2001.01368.x
- DeSalle, R. and Birstein, V.J. 1996. PCR identification of black caviar. Nature 381: 197-198. doi:10.1038/381197a0
- Doukakis, P. 2000. Systematics and Conservation Genetics of Sturgeons (Order: Acipenseriformes), phD Thesis, Yale University.
- Ferguson, A., Taggart, J.B., Prodohl, P. A., McMeel, O., Thompson, C., Stone, C., McGinnity, P., & Hynes, R. A. 1995. The application of molecular markers to the study and conservation of fish populations, with special reference to *Salmo*. Journal of Fish Biology, 47: 103-126. doi: 10.1111/j.1095-8649.1995.tb06048.x.
- Findeis, E.K. 1997. Osteology and Phylogenetic Interrelations of Sturgeons (Acipenseridae). Env. Biol. Fish., 48: 73-126. doi:10.1007/0-306-46854-9_5
- Geldiay, R. and Balık, S. 1996. Türkiye Tatlısu Balıkları. Ege Ünv. Su Ürünleri yayın No: 46, Ege Ünv. II.

Baskı. Basımevi, İzmir, 519 pp.

- Gharaei, A., Pourkazemi, M., Rezvani, S. and Mojazi Amiri, B. 2005. Genetic differences and resemblance between Acipenser persicus and Acipenser gueldenstaedtii by means of RAPD technique. Iranian Scientific Fisheries Journal, 14: 91-102
- Grande, L. and Bemis, W. E. 1991. Osteology and Phylogenetic Relationships of Fossil and Recent Paddlefishes (Polyodontidae) with Comments on the Interrelationships of Acipenseriformes. J. Vertebr. Paleo, 11: 1-121. doi:10.1080/02724634.1991. 10011424
- Grant, W.S. and Bowen, B.W. 1998. Shallow Population Histories in Deep Evolutionary Lineages of Marine Fishes: Insights from Sardines and Anchovies and Lessons for Conservation. J. Hered., 89: 415–426. doi:10.1093/jhered/89.5.415
- Hall, T.A. 1999. Bioedit: A User-Friendly Biological Sequence Alignment Editor and Nalysis Program For Windows 95/98/NT. Nucl. Acids. Symp. Ser, 41: 95-98.
- Huelsenbeck, J.P. and Ronquist, F. 2001. MrBayes: Bayesian inference of phylogenetic trees. Bioinformatics 17: 754–755. doi:10.1093/ bioinformatics/17.8.754
- Ivanenkov, V. V. and Kamshilin, I. N. 1991. On the possibility of using albumin fractions of sturgeons as genetic markers for population studies. Voprosy Ikhtiologii, 31: 234-232.
- Jenneckens, I., Meyer, J. N., Debus, L., Pitra C. and Ludwig A. 2000. Evidence of Mitochondrial DNA Clones of Siberian sturgeon, *Acipenser baerii*, within Russian sturgeon, *Acipenser gueldenstaedtii*. Ecol. Lett., 3: 503–508. doi:10.1111/j.1461-0248.2000. 00179.x
- Jenneckens, I., Meyer, J. N., Hörstgen-Schwark, G., May, B., et al. 2001. A fixed allele at microsatellite LS-39 is characteristic for the black caviar producer Acipenser stellatus. Journal of Applied Ichthyology, 17: 39-42. doi:10.1046/j.1439-0426.2001.00234.x
- Johns, G. C. and Avise, J. C. 1998. A comparative summary of genetic distances in the vertebrates from the mithocondrail cytochrome b Gene. Mol. Biol. Evol., 15: 1481-1490.
 - doi:10.1093/oxfordjournals.molbev.a025875
- Kimura, M. 1980. A Simple Method for Estimating Evolutionary Rates of Base Substitutions Through Comparative Studies of Nucleotide Sequences. J. Mol. Evol. 16, 111–120. doi:10.1007/BF01731581
- Kocher, T.D., Thomas, W.K., Meyer, A., Edwards, S.V., Paabo, S., Villablanca, F.X. and Wilson, A.C. 1989. Dynamics of mitochondrial DNA sequence evolution in animals. Proc. Natl. Acad. Sci. USA, 86: 6196-6200. doi: 10.1073/pnas.86.16.6196.
- Kumar, S., Tamura, K. and Nei, M. 2004. MEGA3: Integrated software for molecular evolutionary genetics analysis and sequence alignment. Briefings in Bioinformatics 5: 150-163. doi:10.1093/bib/5.2.150
- Kuz'min, Y. V. 1991. Comparative study of isozymes of muscle malate dehydrogenase of the Ob population of the Siberian sturgeon, *Acipenser baerii*, and the Don and Kama Sterlet, *A. ruthenus*. Voprosy Ikhtiologii, 31: 139-144.
- Kuz'min, Y. V. 1994. Comparative Analysis of the Fractional Composition of Sarcoplasmic Muscle Proteins of Different Representatives of Sturgeon

(Acipenseridae). J. Icht., 34: 111-124.

- Kuz'min, Y. V. 2002. Allozyme Variation of Nonspecific Esterases in Russian Sturgeon Acipenser güldenstädti Br Comparative Analysis of the Fractional andt. Russian Journal of Genetics, 38(4): 408-414.
- Laukhin, S. A. 1997. The Late Pleistocene Glaciation in the Northern Chukchi Peninsular. Quaternary International, 41-42: 33-41. doi:10.1016/S1040-6182(96)00034-1
- Ludwig, A., May, B., Debus, L.and Jenneckens, I. 2000. Heteroplasmy in the mtDNA Control Region of Sturgeon (Acipenser, Huso, and Scaphirhynchus). Genetics, 156: 1933–1947.
- Ludwig, A., Debus, L. and Jenneckens, I. 2002. A molecular approach for trading control of black caviar. International Review of Hydrobiology, 87: 661-674.
- Ludwig, A., Sebastian, L., Lutz, D., Ralf, R. 2002. First evidence of hybridization between endangered sterlets (*Acipenser ruthenus*) and exotic Siberian sturgeons (*Acipenser baerii*) in the Danube River Arne. Biol Invasions, 11: 753-760. doi: 10.1007/s10530-008-9289-z
- Ludwig, A. 2006. A sturgeon view on conservation genetics. European Journal of Wildlife Research. 52:3-8. doi:10.1007/s10344-005-0006-2
- Mayden, R.L. and Kuhajda, B.R. 1996. Systematics Taxonomy and Conservation Status of the Endagered Alabama Sturgeon *Scaphirhynchus suttkusi* Williams and Clemmer (Actinopterygii, Acipenseridae). Copeia 2: 241-273. doi:10.2307/1446842
- Meyer, A. 1993. Evolution of Mitochondrial DNA in Fishes. Bioc. and Mol. Biol. Fish., 2: 1-38.
- Moghim, M., Heist, E.J., Tan S. G., Pourkazemi, M., Siraj, S.S., Panandam, J.M., Pourgholam, R., Kor, D., Laloei, F. and Taghavi, M. J. 2012. Isolation and characterization of microsatellite loci in the Persiansturgeon (*Acipenser persicus*, Borodine, 1897) and cross-speciesamplification in four commercial sturgeons from the Caspian Sea. Iranian Journal of Fisheries Sciences 11(3): 548-558.
- Mugue, N., Barmintseva, A. E., Rastorguev, S. M., Mugue, V. N. and Barmintsev, V. A. 2006. Sturgeon mtDNA control region polymorphism and its suitability for species identification. 2ndWorkshop. Identification of Acipenseriformes Species in Trade. Berlin, 29th Sep. - 1st Oct. 2006: 9-10.
- Nuin, P. 2008. MrMTgui: cross-interface for ModelTest and Mr Modeltest; "http://www.genedrift.org/mtgui.php".
- Nylander, J.A.A. 2004. MrModeltest v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.
- Orti, G., Bell, M.A., Reimchen, T.E. and Meyer, A. 1994. Global survey of mitochondrial DNA sequences in the threespine stickleback: evidence for recent migrations. Evolution, 48: 608-622. doi: 10.2307/2410473.
- Peng, Z., Ludwig, A., Wang, D., Diogo, R., Wie, Q. and He, S. 2007. Age and Biogeography of Major Clades in Sturgeons and Paddlefishes (Pisces: Acipenseriformes). Mol. Phyl. Evol., 42: 854-862. doi:10.1016/j.ympev.2006.09.008
- Posada, D. and Crandall, K. A. 1998. Modeltest: Testing The Model of DNA Substitution. Bioinformatics, 14: 817-818. doi:10.1093/bioinformatics/14.9.817
- Pourkazemi, M. 1996. Molecular and biochemical genetic analysis of sturgeon stocks from the South Caspian

Sea. Ph.D. thesis. University of Wales, Swansea.

- Rehbein, H. 1997. Fischartbestimmung von Caviar durch Protein- und DNA-Analyse. Informationen der Fischwirtschaft, 44: 27-30.
- Ronquist, F. and Huelsenbeck, J. P. 2003. Mr Bayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics, 19: 1572–1574. doi:10.1093/bioinformatics/btg180
- Rozas, J. and Rozas, R. 1999. DnaSP Version 3: An Integrated Program for Molecular Population Genetics and Molecular Evolution Analysis. Bioinformatics, 15: 174-175. doi:10.1093/bioinformatics/15.2.174
- Ruban, G. I. 1997. Species Structure, Contemporary Distributiona and Status of the Siberian sturgeon, *Acipenser baerii*. Environ. Biol. Fish., 48, 221–230. doi:10.1007/0-306-46854-9_12
- Sambrook, J., Fritsch, E. F. and Maniatis, T. 1989. Molecular Cloning: A Laboratory Manual, vol. I. 2nd edition, Cold Spring Harbor Laboratory Press.
- Schneider, S., Roessli, D., Excoffier, L. 2000. ARLEQUIN: Software for Population Genetics Data Analysis (version 2.000). http://lgb. unige.ch/arlequin/.
- Schwarz, G. 1978. "Estimating the Dimension of a Model". Annals of Statistics, 6: 461-464. doi:10.1214/aos/1176344136
- Simon, C. 1991. Molecular systematics at the species boundary: exploiting conserved and variable regions of the mitochondrial genome of animals via direct sequencing from amplified DNA. In: G.M. Hewitt, A.W.B. Johnston, J.P.W. Young, (Eds.), Molecular Techniques in Taxonomy. Springer-Verlag. New York: doi:10.1007/978-3-642-83962-7_4
- Stabile, J., Waldman, J. R., Parauka, F. and Wirgin, I. 1996. Stock Structure and Homing Fidelity of Gulf Sturgeon (*Acipenser oxyrinchus desotoi*) Based on Restriction Fragment Length Polymorphism and Sequence Analyses of Mitochondrial DNA. Genetics, 144: 767-775.
- Stepien, C.A. and Kocher, T.D. 1997. Molecules and morphology in studies of fish evolution. Pp. 1-11 in Molecular Systematics of Fishes, In: T.D. Kocher, C.A. Stepien (Eds), Academic Press. doi: 10.1016/B978-012417540-2/50002-6.
- Swofford, D.L. 2003. PAUP*: Phylogenetic Analysis Using Parsimony (* and Other Methods), Version 4.0b 10. Sinauer Associates, Sunderland, Massachusetts.

- Tajima, F. 1983 Evolutionary Relationship of DNA Sequences in Finite Populations. Genetics, 105: 437-460.
- Tamura, K. and Nei, M., 1993. Estimation of the Number of Nucleotide Substitutions in the Control Region of the Mitochonrial DNA in Human and Chimpanzees. Mol. Biol. and Evol., 10: 512-526.
- Timoshkina, N. N., Vodolazhskii, D. I. and Usatov, A. V. 2011. Molecular-genetic markers in studies of intraand interspecies polymorphism in sturgeon (Acipenseriformes). Russian Journal of Genetics: Applied Research, 1(2): 160-171. doi:10.1134/S2079059711020122
- Vlasenko, A. D., Pavlov, A.V., Sokolov, L. I. and Vasıl'ev, V. P. 1989. Acipenser gueldenstaedtii Brandt 1833. In: The Freshwater Fishes of Europe (Holcík J. Hrsg.), AULA-Verlag Gmbh, Wiesbaden: 294-344.
- Voynova, N.V., Mirzoyan, A.V, Timoshkina, N. N and Rynza, E. T. 2008. Occurrence of Non-notive Specimens of Caspian Origin within the Sea of Azov population of the Russian sturgeon (*Acipenser* gueldenstaedtii). J. App. Icht., 24: 50-51. doi:10.1111/j.1439-0426.2008.01090.x
- Williot, P., Arlati, G., Chebanov, M., Gulyas, T., Kasimov, R., Kirschbaum, F., Patriche, N., Pavlovskaya, L. P., Poliakova, L., Pourkazemi, M., Kim, Y., Zhuang, P. and Zholdasova, I. M. 2002. Status and management of Eurasian sturgeon: an overview. International Reviews in Hydrobiology, 87(5-6): 483-506.
- Wolf, C., Hübner, P. and Lüthy, J. 1999. Differentiation of sturgeon species by PCR-RFLP. Food Research International, 32: 699-705. doi:10.1016/S0963-9969(99)00150-7
- Zardoya, R., Doadrio, I.1999. Molecular Evidence on the Evolutionary and Biogeographical Patterns of European Cyprinids. Journal of Molecular Evolution, 49: 227-237. doi: 10.1007/PL00006545.
- Zhu, B., Zhou, F., Cao, H., Shao, Z., Zhao, N., May, B., and Chang, J. 2002. Analysis of genetic variation in the Chinese sturgeon, Acipenser sinensis: estimating the contribution of artificially produced larvae in a wild population. J. App. Icht., 18: 301–306. doi:10.1046/j.1439-0426.2002.00379.x
- Zuckerkandl, E. and Pauling, L.1965. Evolutionary divergence and convergence in proteins, Evolving Genes and Proteins, edited by V. Bryson and H.J. Vogel. Academic Press, New York, 97-166 pp.