



Effect of the Integration between Plankton Natural Productivity and Environmental Assessment of Irrigation Water, El-Mahmoudia Canal, on Aquaculture Potential of *Oreochromis niloticus*

Mohamed Ashour^{1,*}, Hamdy Abo-Taleb², Mohamed Abou-Mahmoud¹, M. M. M. El-Feky¹

¹ National Institute of Oceanography and Fisheries, Alexandria Branch, Egypt.

² Al-Azhar University, Faculty of Science, Cairo, Egypt.

* Corresponding Author: Tel.: +20.128 3184088
E-mail: microalgae_egypt@yahoo.com

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Abstract

Aquaculture ecosystem assessment is a major key for successful aquaculture Planning. There are many aquaculture farms constructed in Egypt, but because of the non-sufficient studies about the environmental conditions of aquaculture ecosystem assessment, many projects achieved negative results and had low economical returns. The present study was conducted to evaluate the aquaculture ecosystem assessment of irrigation water of El-Mahmoudia Canal, Alexandria, Egypt. The result indicated that temperature ($24.65 \pm 0.4^\circ\text{C}$), DO ($5.38 \pm 0.20\text{mg/l}$), COD ($14.40 \pm 4.5\text{mg/l}$) and BOD ($2.86 \pm 0.20\text{mg/l}$) were in the optimum range for *O. niloticus*. Chlorophyll *a*, *b* and carotenoids were 10.449 ± 0.953 , 4.685 ± 0.704 and $2.374 \pm 0.054\ \mu\text{g/g}$, respectively. The biochemical composition of plankton showed that protein, lipid and carbohydrate were $37.7 \pm 3.39\%$, $16.33 \pm 0.85\%$, and $11.23 \pm 2.63\%$, respectively. Moreover, SFA, MUFA and PUFA were $70.29 \pm 4.07\%$, $11.172 \pm 0.132\%$, and $18.55 \pm 0.525\%$ of total fatty acids, respectively. Zooplankton community showed that rotifers and copepods configured the main bulk of zooplankton community (95.9% to the total zooplankton community). These conditions are considered the ideal recommended conditions for cultivation of *Oreochromis niloticus* that started with 100,000 fingerlings (distributed as 4.76 fish/m^3 of earth pond) for six months with 82% survival rate with final yield of 21.3 ton/1.6 hectare.

Keywords: Aquaculture ecosystem, Tilapia, phytoplankton, zooplankton, fatty acids.

Introduction

Aquaculture is a global important industry that supplies essential food to a growing world population, with an essential role in providing a cheap animal protein. In recent years, aquaculture sector has been greatly developing and is considered one of the most promising industries in Egypt. Subsequently, aquaculture acts as the most important tool for decreasing the current gap between consumption and fish production in Egypt. Despite the pressure on water, Egypt has the largest aquaculture industry in Africa with a market value of over \$2.18 billion, and globally, Egypt ranks the 8th in fish farming production and the 1st among African countries. Widely geographic distribution of *Oreochromis niloticus* is nominated to be a major freshwater aquaculture species in several developing countries (Ble, Etchian, Alla, Adingra, Niamke, & Diopoh, 2012). Tilapia, especially *Oreochromis niloticus*, comprises more than 75% of Egyptian aquaculture production, the production in 2012 estimated about 768,752 ton. Furthermore, after China, Egypt ranks

the 2nd in the production of *Oreochromis niloticus*, which represent the main farmed fish in Egypt (Soliman & Yacout, 2016). There are many aquaculture farms constructed in Egypt, but because of the non-sufficient studies about the environmental conditions of aquaculture ecosystem, many projects achieved negative results and had low economical returns.

The aquaculture production is directly affected by aquaculture ecosystems. Nutrient availability, water quality, soil grain size analysis, and plankton (community, productivity and nutritional composition) are the main ecosystem components affecting the cultured fish in earthen ponds (Ssanyu, Rasowo, Auma, & Ndunguru, 2011). The enhancement of nutrient availability in the aquaculture ecosystems is controlled by added fertilizers (El-Otify, 2015). Fertilization in fish earthen ponds accumulates nutrients at the bottom of ponds. The main source of the important nutrients (nitrogen and phosphorus) in earthen fish ponds are the added fertilizers, fish excretion and unconsumed feed. Moreover, decomposition of organic matter

produces ammonia, which diffuses from sediment to the water column. Consequentially, nutrient availability is easily incorporated into the plankton (mainly phytoplankton and sequentially zooplankton) community (Case *et al.*, 2008), productivity (Chib, Masud, & Proch, 2015), biochemical composition (Mitra, Mukhopadhyay, & Ayyappan, 2007) and ultimately into the fish biomass (Tavares, Millan, & Santeiro, 2010). Zooplankton takes a central position between the autotrophs and other heterotrophs maintaining an essential link in the food chain sustainability. In aquaculture conditions, cultured species derive an essential part of their dietary nutrient needs from available plankton as a live food exist in water column, which they are a valuable source of protein, carbohydrate, lipid, fatty acids, and vitamins (Mitra *et al.*, 2007).

Egypt is a country which has limited water resources and that reflects the quality and quantity of available water for aquaculture. (Soliman & Yacout, 2016). Alexandria, the main coastal city of Egypt, is limited in aquaculture activity because of many considerations, although this coastal city contains all different water resources of marine, freshwater, brackish water, Lake Maryout and underground water. El-Mahmoudia Canal is considered the main freshwater source of Alexandria City, Egypt. However, this study may be considered the first study that investigates the aquaculture potential and aquaculture ecosystem of El-Mahmoudia Canal.

The present study was conducted to evaluate the feasibility, natural plankton productivity and environmental assessment of utilizing the irrigation water of El-Mahmoudia Canal, Alexandria, Egypt, as a supplemented water source for culture of Nile Tilapia *Oreochromis niloticus*.

Materials and Methods

Before the beginning of culture, water quality, nutrient availability, soil grain size analysis, phytoplankton pigments, zooplankton community, total plankton productivity and nutritional composition (protein, carbohydrate, lipid and fatty acid profile) were measured in irrigation water of El-Mahmoudia Canal, Alexandria, which is used as culture water for *O. niloticus* in earthen pond.

Water Quality

Water quality parameters, like temperature (C°), pH (pocket digital pH meter), turbidity (*NTU*, Turbidity Meter bench HACH), electrical conductivity (EC, E.C-Meter portable HACH), total dissolved solids (TDS, Salinometer Oyster, inspected 82738, Extech instruments, Germany) and dissolved oxygen (DO, DO- meter HQ30d flexi meter HACH) were measured. Biological oxygen demand (BOD) and chemical oxygen demand (COD) were measured according to Pawlowski (1994). Moreover,

spectrophotometric determination of nitrate (NO₃), nitrite (NO₂) free ammonia (NH₃) and phosphate (PO₄) were measured according to Grasshoff (1976).

Grain Size Analyses

Grain size analyses of soil were conducted according to ASTM D421-85 (1998).

Plankton Investigations

Phytoplankton

Chlorophyll was measured as an indicator for Phytoplankton biomass. Chlorophyll-*a*, chlorophyll-*b* and total carotenoids were determined according to Dere, Gunes, and Sivaci (1998).

Plankton Productivity

One ton of irrigation water of El-Mahmoudia Canal, Alexandria, was filtered (three replicates) by plankton net (55 µm). The obtained biomass was washed twice with distilled water, dried at 70 °C for 24 h, cooled in desiccator and weighted to determine the plankton productivity in dry weight form (Olguin, Galicia, Angulo-Guerrero, & Hernandez, 2001).

Plankton Nutritional Composition

Dried plankton samples were kept under -20°C until the analysis of biochemical composition, as total proteins, total carbohydrates, total lipids and fatty acids (three replicates for each analysis). Total proteins were extracted according to Rauch (1981) and determined according to Hartree (1972). Total carbohydrates were extracted according to Myklestad and Haug (1972) and determined according to Dubois, Giles, Hamilton, Rebers, and Smith (1956). Total lipid was calculated according to Bligh and Dyer (1959). Preparation of fatty acids methyl ester from total lipids was performed according to the procedure of Radwan (1978). Identification of fatty acids fractions were performed on gas chromatography instrument, model HP (Hewlett Packard) 7890 GC equipped with a flame ionization detector; column (HP-5, 30 m, 0.32 mm ID, 0.25 µm film thickness), detector (FID) with detector temperature (250 °C), injected temperature (220 °C, injection volume 2 µl, splitless mode), and nitrogen carrier gas with gas flow 1 ml/min.

Zooplankton Community

Collections of samples were carried out by standard plankton net (No. 25) of 55 µm mesh size which lowered vertically till near bottom then pushed up to the water surface. Zooplankton organisms were transferred from the plankton net into small glass bottles, preserved in 5% neutralized formalin solution

and then the sample volume was adjusted to 100 ml. The samples were examined under a binocular research microscope. The identification was undertaken to species levels. For estimation of standing crop, subsamples of 5 ml were transferred to a counting chamber (Bogorov chamber) using a plunger pipette (three times). Identification of zooplankton community was carried out according to Sars (1911, 1918 and 1926), Gurney (1927 and 1931), Edmondson (1959), Berzins (1960), Hardig and Smith (1974), Hutchinson (1967), Dussart (1969) and Bick (1972). All zooplankton organisms were counted to species level and the standing crop was calculated and estimated as organisms per cubic meter, according to the following formula:

$$N = n (v/V) * 1000$$

Where, N : Total number of zooplankton per cubic meter. n : Average number of zooplankton in 1 ml of the sample. v : Volume of zooplankton concentrate (ml). V : Volume of total water filtered (L).

Oreochromis Niloticus Culture

100,000 fingerlings of *Oreochromis niloticus*, with mean body weight 3.77 g obtained from tilapia hatchery in Kafer El-sheekh, Egypt, were transported and acclimatized into the earthen fish pond in Mr. Ehab Abdel-Fatah fish Farm exist Alexandria City (31° 12' 30.07" N and 29° 58' 41.66" E), Egypt, (Figure 1). The experiment was set in three connected earthen ponds with total area about 4 acre (about 1.6 hectare). The depth of ponds ranges from 1.2 to 1.4 m

with a total volume of water of 21,010 m³.

O. niloticus were randomly distributed with stocking density 4.76 fish/m³. The culture of *O. niloticus* in ponds was conducted for 210 days. Ponds preparations included the basal fertilization of cow dung at 10 ton/1.6 ha, followed by chicken manure (3 ton/1.6 ha/month) in burlap bags 50 kg/bag. *O. niloticus* were fed 20 tons supplementary diet containing 25% crude protein (diet composition are shown in Table 1). The feeding rate was 3% of fish body weight, for twice a day, six days a week, depending on fish biomass. Growth performance parameters in term of weight gain (TWG), average daily gain (ADG), specific growth rate (SGR), feed conversion ratio (FCR) and Protein efficiency ratio (PER) were calculated according to Bahnasawy, Abdel-Baky, and Abd-Allah (2003) as the following:

WG (kg) = Final Weight (kg) – Initial Weight (kg);

ADG (g/d) = Weight Gain (g) / Time (days);

SGR (%/d) = 100 X (Ln Final Weight (g) – Ln Initial Weight (g))/Time (days);

FCR = Food Consumed (kg) / Weight Gain (kg);

PER = Live Weight Gain (g) / Protein Consumed (g).

Results

Water Quality

Table 2 shows the mean values of different water quality parameters recorded of irrigation water of El-Mahmoudia Canal, as a water source for cultured *O. niloticus*. The results indicated that the



Figure 1. Location of Ehab fish Farm, El-Mahmoudia Canal, Alexandria, Egypt.

Table 1. Composition of supplemental pelleted diets (25% crude protein)

Ingredients	%
Fish Meal	20
Soy Bean	15
Yellow Corn	30
Wheat bran	30
Oil	3
Vitamin & Mineral	1
Baker Yeast	1

Table 2. Water quality parameters of irrigation water of El-Mahmoudia Canal

Parameter	Means±SD
Temperature (°C)	24.65±0.4
Dissolved Oxygen (DO, mg/l)	5.38±0.20
pH (pH)	7.51±0.10
Turbidity (NTU)	9.75±1.10
Chemical Oxygen Demand (COD, mg/l)	14.40±4.5
Biological Oxygen Demand (BOD, mg/l)	2.86±0.20
Alkalinity (mg CaCO ₃ /l)	162.8±12.7
Total Dissolved Solids (TDS, mg/l)	340.0±27.4
Electrical Conductivity (EC, µS/cm)	566±32.1
Nitrate (NO ₃ , mg/l)	1.96±0.60
Nitrite (NO ₂ , mg/l)	0.50±0.10
Ammonia, free (NH ₃ , mg/l)	0.84±0.20
Phosphate (PO ₄ , mg/l)	0.59±0.20

recorded water quality parameters, including temperature (24.65±0.4 °C), DO (5.38±0.20 mg/l), pH (7.51±0.1), turbidity (9.75±1.10 NTU), COD (14.40±4.5 mg/l), BOD (2.86±0.20 mg/l), alkalinity (162.8±12.7 mg/l), TDS (340.0±27.4 mg/l), and EC (566±32.1 µS/cm). As well as, the values of NO₃ (1.96±0.60 mg/l), NO₂ (0.50±0.10 mg/l) and NH₃ (0.84±0.20 mg/l), and PO₄ (0.59±0.20 mg/l) were recorded in irrigation water of El-Mahmoudia Canal.

Soil Grain Size Analysis

The grain size analysis of sediment sample that collected from the farm location area indicated that the sediment type was mostly fine silt. It composed of the 50.4% fine silt, 31.3% medium silt, while 13.7% was very fine silt and 4.5 % clay (Table 3 and Figure 2). The relation between particles diameter and class weight proportions showed that more than 70% of the sediment particles diameter was between 5 and 8 µm (Figure 3). The sediment cumulative mass retained proportions were negatively correlated with the particles size, they decreased gradually with the increasing of particles size and reached their maximum values at the sediments with particle diameter more than 10 µm (Figure 4).

Plankton Investigations

Plankton Productivity and Nutritional Composition

The results in Table 4 showed that chlorophyll *a*, *b* and total carotenoids were 10.449±0.953,

4.685±0.704 and 2.374±0.054 µg/g fresh wet of total plankton, respectively. As well as, the present study found that the plankton productivity (in dry weight form) was 4.281±0.216 g dw/ton of El-Mahmoudia Canal irrigation water, as shown in Table 4. Furthermore, the nutritional composition of total plankton (total protein, total lipid and total carbohydrate) were 47.7±3.39%, 16.33±0.85%, and 11.23±2.63%, respectively, as shown in Table 4. Moreover, fatty acids profiles were shown in Table 5. The contents of total saturated fatty acids (SFA), monounsaturated fatty acids (MUFA) and polyunsaturated fatty acids (PUFA) were 70.29±4.07%, 11.172±0.132 %, and 18.55±0.525 % of total fatty acids, respectively. The predominant fatty acid in SFA was 16:0 (28.38%), while the predominant in MUFA were 18:1n-9 (3.47%) and 16:1n-7 (3.31%). The predominant fatty acids in PUFA were DHA C22:6n-3 (10.40 %) followed by C18:2n-6 (7.896 %) and LNA 18:3 n-3 (0.25%), as shown in Table 5.

Zooplankton Community

In the present study, zooplankton community showed in Table 6 and Table 7 found that there were 57 forms of different organisms, including the mature and immature forms. The most diversified groups were Rotifera, Copepoda, Protozoa and Cladocera. Rotifers were represented by 27 species in order to several forms of rotifers metamorphoses which represent 49.8% of the total recorded species. There were 7 Copepod species and their immature forms (Copepodite stages and Nauplius larvae). Cladocerans

Table 3. Grain size analysis of studied sample

SAMPLE STATISTICS						
SAMPLE IDENTITY: 5			ANALYST & DATE: Abou-Mahmoud, 26-7-2016			
SAMPLE TYPE: Unimodal, Moderately Well Sorted			TEXTURAL GROUP: Mud			
SEDIMENT NAME: Fine Silt						
	μm	ϕ	GRAIN SIZE DISTRIBUTION			
MODE 1:	8.250	6.922	GRAVEL: 0.0%	COARSE SAND: 0.0%		
MODE 2:			SAND: 0.0%	MEDIUM SAND: 0.0%		
MODE 3:			MUD: 100.0%	FINE SAND: 0.0%		
D ₁₀ :	2.848	6.771		V FINE SAND: 0.0%		
MEDIAN or D ₅₀ :	6.753	7.210	V COARSE GRAVEL: 0.0%	V COARSE SILT: 0.0%		
D ₉₀ :	9.157	8.456	COARSE GRAVEL: 0.0%	COARSE SILT: 0.0%		
(D ₉₀ / D ₁₀):	3.215	1.249	MEDIUM GRAVEL: 0.0%	MEDIUM SILT: 31.3%		
(D ₉₀ - D ₁₀):	6.309	1.685	FINE GRAVEL: 0.0%	FINE SILT: 50.4%		
(D ₇₅ / D ₂₅):	1.737	1.115	V FINE GRAVEL: 0.0%	V FINE SILT: 13.7%		
(D ₇₅ - D ₂₅):	3.461	0.797	V COARSE SAND: 0.0%	CLAY: 4.5%		
	METHOD OF MOMENTS			FOLK & WARD METHOD		
	Arithmetic	Geometric	Logarithmic	Geometric	Logarithmic	Description
	μm	μm	ϕ	μm	ϕ	
MEAN (\bar{x}):	6.374	5.773	7.426	5.974	7.387	Fine Silt
SORTING (σ):	2.350	1.640	0.713	1.577	0.657	Moderately Well Sorted
SKEWNESS (Sk):	-0.413	-1.510	1.551	-0.473	0.473	Very Fine Skewed
KURTOSIS (K):	2.389	5.422	5.507	1.165	1.165	Leptokurtic

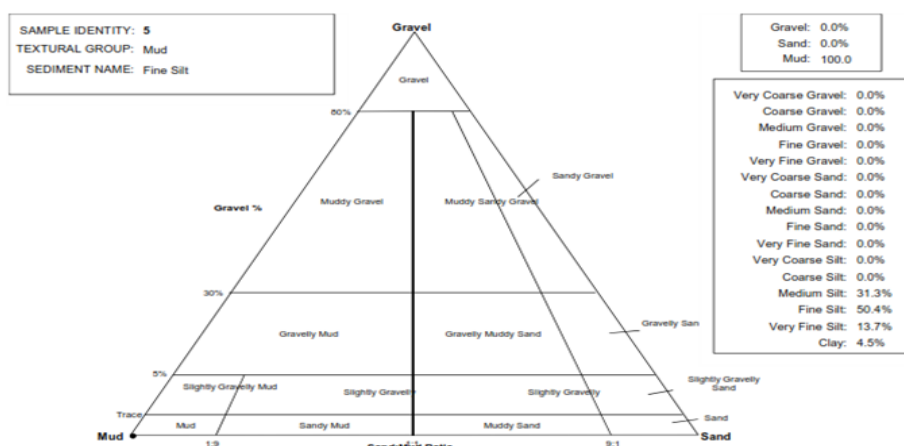


Figure 2. Grain size analysis of studied sample.

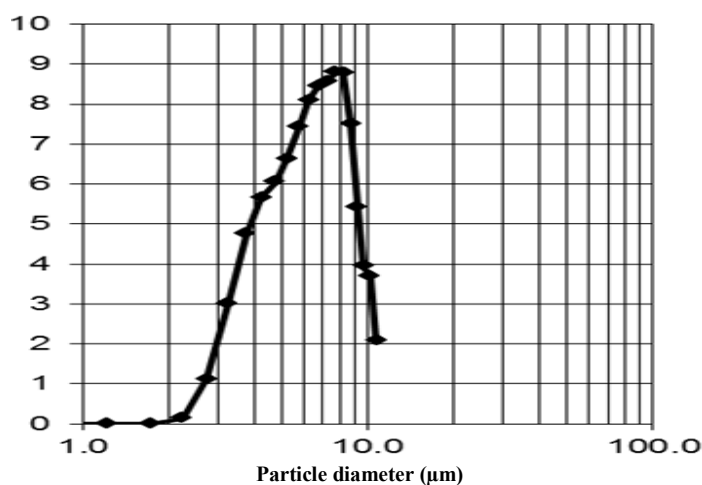


Figure 3. The relation between particles diameter and class weight proportions.

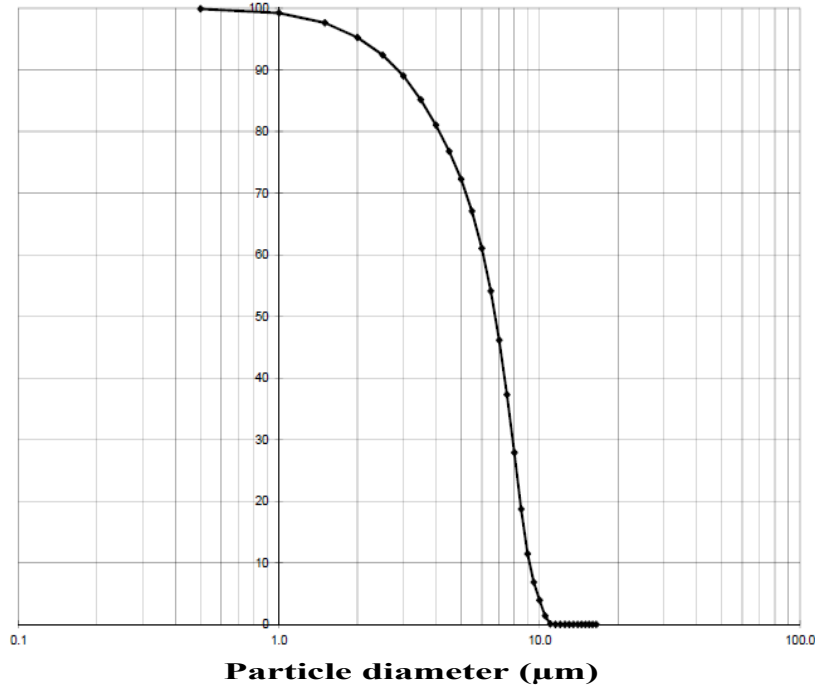


Figure 4. Cumulative frequency curve.

Table 4. Pigments ($\mu\text{g/g}$), biochemical composition (% of dw) and productivity (g dw/ton) of total plankton presented in irrigation water of El-Mahmoudia Canal

Pigments of Phytoplankton			Biochemical composition			Plankton productivity
<i>Chlorophyll a</i>	<i>Chlorophyll b</i>	<i>Carotene</i>	<i>Protein</i>	<i>Lipid</i>	<i>Carbohydrate</i>	(Dry weight)
10.449 \pm 0.953	4.685 \pm 0.704	2.374 \pm 0.054	47.7 \pm 3.39	16.33 \pm 0.85	11.23 \pm 2.63	4.281 \pm 0.216

Table 5. Fatty acid profile (g/100g lipid) of mixed zooplankton presented in irrigation water of El-Mahmoudia Canal

Fatty Acids	Fatty acid %
C8:0	0.08 \pm 0.006
C10:0	0.23 \pm 0.002
C12:0	0.34 \pm 0.004
C13:0	0.73 \pm 0.002
C14:0	3.74 \pm 0.226
C15:0	2.57 \pm 0.125
C16:0	28.38 \pm 2.98
C17:0	3.08 \pm 0.002
C18:0	13.43 \pm 0.663
C20:0	14.57 \pm 0.261
C21:0	3.14 \pm 0.179
* Σ SFA	70.29 \pm 4.07
C14:1n-5	0.30 \pm 0.018
C15:1	1.32 \pm 0.027
C16:1n-7	3.31 \pm 0.048
C17:1	2.75 \pm 0.009
C18:1n-9	3.47 \pm 0.103
* Σ MUFA	11.17 \pm 0.132
C18:2n-6	7.896 \pm 0.513
C18:3n-3	0.25 \pm 0.014
C22:6n-3	10.40 \pm 1.053
* Σ PUFA	18.55 \pm 0.525

SFA: saturated fatty acids

MUFA: Monounsaturated fatty acids

PUFA: Polyunsaturated fatty acids

and Protozoa were expressed by 5 and 6 species, respectively. On the other hand, there were two Molluscan forms (Lamellibranch and Gastropod larvae). Moreover, two types of insect larvae (Chironomus and non Chironomus) were observed. Also, in addition to fish eggs, one of Diplostraca and Ostracod species were observed, as presented in Table 6 and 7.

Regarding to the density, the total zooplankton number was 33565 organism/m³. Rotifers and copepods configured the main bulk of zooplankton and they figured together 95.9% to the total zooplankton numbers (32189 org./m³). Rotifers were

the highest population recorded during the present study, and estimated by 16716 org./m³ and represent 49.8% of total zooplankton community. Genus *Brachionus* expressed by 8 species formed about 11148 org./m³ (33.2% of total zooplankton and 67% of total rotifer community). Copepods were the second highest population recorded after rotifers, estimated by 15473 org./m³ and constituted 46.1% of the total zooplankton counts. There were two species of the genus *Acanthocyclops* formed 40% of the total copepods community and 18% of total zooplankton community. In conclusion, 10 species (8 rotifers and 2 copepods) formed more than 50% of the total

Table 6. Different zooplankton groups recorded in irrigation water, El-Mahmoudia Canal

Groups	Composition		
	No.	Org./m ³	%
<i>Rotifera</i>	28	16716	49.80
<i>Copepoda</i>	9	15473	46.10
<i>Protozoa</i>	6	297	0.88
<i>Cladocera</i>	5	367	1.09
<i>Mollusca</i>	3	146	0.43
<i>Insecta</i>	2	92	0.27
<i>Diplostraca</i>	1	65	0.19
<i>Nematoda</i>	1	32	0.10
<i>Ostracoda</i>	1	372	1.11
<i>Fish eggs</i>	1	5	0.01
Total	57	33565	100

Table 7. List of the recorded zooplankton organisms (community) at the study area, irrigation water, El-Mahmoudia Canal

ROTIFERA	COPEPODA	MOLLUSCA
<i>Ascomorpha saltan saltans</i> Bartsch, 1870	<i>Acanthocyclops americanus</i> Marsh, 1893	<i>Pleurocera acuta</i> Rafinesque, 1831
<i>Asplanchna priodonta</i> Gosse, 1850	<i>A. vernalis</i> Fischer, 1853	Gastropoda veligers
<i>Brachionus angularis</i> , Gosse, 1851	<i>Diacyclops bicuspidatus odessanus</i> Shmankevich, 1875	Lamellibranch veligar
<i>B. budapestinensis</i> Daday, 1885	<i>Halicyclops magniceps</i> Lilljeborg, 1853	
<i>B. calyciflorus</i> Pallas, 1766	<i>Mesocyclops leuckarti</i> Claus, 1857	DIPLOSTRACA
<i>B. caudatus</i> Barrois and Daday, 1894	<i>Schizopera clandestina</i> Klie, 1924	<i>Oxyurella tenuicaudis</i> G.O.Sars, 1862
<i>B. leydigii</i> Cohn, 1862	<i>Thermocyclops crassus</i> Fisher, 1853	
<i>B. plicatilis</i> Müller, 1786	Copepod stages	INSECTA
<i>B. quadridentatus</i> Hermann, 1783	Nauplius larvae	<i>Chironomus</i> larvae Meigen, 1803
<i>B. urceolaris</i> Müller, 1773		Insects larvae
<i>Diplois daviesia</i> Gosse, 1886	PROTOZOA	
<i>Filinia longiseta</i> Ehrenberg, 1834	<i>Centropixis aculata</i> Ehrenberg, 1832	
<i>Keratella cochlearis</i> Gosse, 1851	<i>C. ecornis</i> Ehrenberg, 1841	NEMATODA
<i>K. quadrata</i> Müller, 1786	<i>Difflogiaglobulosa</i> Dujardin, 1837	Nematode larvae
<i>K. tropica</i> Apstein, 1907	<i>Difflogia</i> sp. Lamarck, 1816	
<i>Lecane luna</i> Müller, 1776	<i>Eugelina acus</i> (O.F. Müller, 1786) Ehrenberg, 1830	OSTRACODA
<i>Lepadella ovalis</i> O.F.Müller, 1896	<i>Euplotes</i> sp. O.F.Müller, 1786	Ostracod species
<i>Monostyla bulla</i> Gosse		
<i>Philodina</i> sp. Ehrenberg, 1830	CLADOCERA	EGGS
<i>polyarthra vulgaris</i> Carlin, 1943	<i>Diaphanosoma excisum</i> G.O.Sars, 1885	
<i>Pomopholyx sulcata</i> Hudson, 1885	<i>Bosmina longirostris</i> O.F.Müller, 1878	
<i>Rotaria rotatoria</i> Pallas, 1766	<i>Conochilus unicornis</i> Rousselet, 1892	
<i>Synchaeta oblonga</i> Ehrenberg, 1832	<i>Lynceus bukobensis</i> O.F.Müller, 1776	
<i>S. okai</i> Sudzuki, 1964	<i>Moina micrura</i> Kurz, 1874	
<i>Testudinella patina</i> Hermann, 1783		
<i>Trichocerca cylindrica</i> Imhof, 1891		
<i>T. Semilis</i> Wierzejski, 1893		
Metamorphoses of rotifers		

zooplankton community. Ostracoda, Cladocera and Protozoa were estimated by 372, 367 and 297 org./m³ and represented about 1.11%, 1.09% and 0.88% of total zooplankton community, respectively. Diplostraca was represented by one species expressed by 65 org./m³ (0.19% total zooplankton community). Molluscan was estimated by 146 org./m³ (0.43% of total zooplankton community), while insects, represented by several larvae and the chironomus larvae, were 92 org./m³ (0.27% of total zooplankton community). On the other hand, Nematoda, which were considered a pollution bio-indicator, was estimated by 32 org./m³ (0.1% of the total zooplankton community). Finally, eggs estimated by 5 egg/m³ (0.01 org./m³ of the total zooplankton community), as shown in Figure 5.

Oreochromis niloticus Culture

The mean values of growth performance, survival rate and feed utilization efficiency of *O. niloticus* cultured in earthen pond using irrigation water of El-Mahmoudia Canal, Alexandria, were summarized in Table 8. In the present study, we used 100,000 fry of *O. niloticus* with average of initial

individual body weight 3.77 g with stocking density 4.76 fish/m³ and the optioned survival rate was 82%. Therefore, the initial total weight and final number of fish was 370 kg and 82000 fish, respectively. Growth performance data presented in Table 8 showed that WG, ADG and SGR were 20930 kg, 99.67 g/d, and 1.93 % d⁻¹, respectively. As well as, feed utilization data showed that FCR and PER was 0.96 and 4.19, respectively, as shown in Table 8. The economic return of *O. niloticus* cultured in irrigation water, El-Mahmoudia Canal, Alexandria, Egypt was presented in Table 9. The final production yield of *O. niloticus* was about 21.3 ton/1.6 ha (about 4 acre) with three different marketing size; (1) grade A: 6.3 ton (weight mean 350 g/fish), grade B: 11 tons (weight mean 250g/fish) and (3) grade C: 4 ton (weight mean 200 g/fish), however, the economic analysis of the current experiment are presented in Table 9.

Discussion

Water Quality

In aquaculture, fish productivity mainly depends on aquaculture ecosystems, especially water quality.

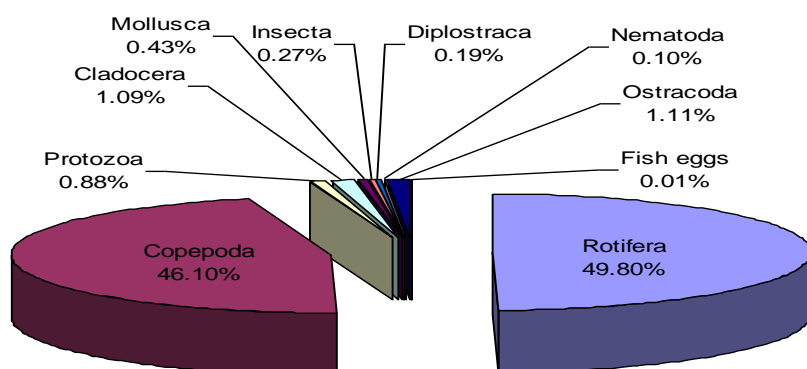


Figure 5. Percentage of different groups of total zooplankton community in irrigation water, El-Mahmoudia Canal.

Table 8. Growth performance and feed utilization of *O. niloticus* cultured in earthen pond using irrigation water of El-Mahmoudia Canal

Parameters	Value
Initial number of fish (fish)	100000
Initial mean individual weight (g)	3.77
Initial total weight (kg)	370
Survival ratio (%)	82
Final number of fish (fish)	82000
Final total weight (kg)	21300
WG (weight gain, kg)	20930
ADG (average daily gain, g/d)	99.67
SGR (Specific growth rate, % d ⁻¹)	1.93
Total feed given (kg)	20000
Protein consumed (kg)	5000
FCR (Feed conversion ratio)	0.96
PER (Protein efficiency ratio)	4.19

Table 9. Economic return of *O. niloticus* in irrigation water, El-Mahmoudia Canal

Item	Price(\$)
<i>Costs:</i>	
<i>O. niloticus</i> fry	357
Diets used	6571
Fertilizers	557
Working cost	3486
<i>Total</i>	10971
<i>Gross Return:</i>	
Fish of grade A: 6.3 ton (mean weight 350 g/fish)	10800
Fish of grade B: 11 ton (mean weight 250 g/fish)	15714
Fish of grade C: 4 ton (mean weight 200 g/fish)	5143
<i>Total</i>	26514
<i>Net return</i>	15543

Poor water quality causes decrease of fish productivity, increase of production costs and risk of diseases. In addition, it has negative effects on the environment, human health, laborers and consumers. *O. niloticus* is one of the most important cultured freshwater fish over the world. *O. niloticus* can tolerate a wider range of environmental conditions such as DO, pH, NO₃, NO₂ and ammonia levels. Therefore, *O. niloticus* can easily culture and adaptable to a wide range of environmental conditions (El-Sayed, 2006). The results of the present study indicated that the recorded water quality parameters, including temperature, DO, pH, COD, BOD, turbidity and alkalinity were in the optimum range for *O. niloticus* (Ross, 2000). Despite the high relative value of TDS (340.0±27.4 mg/l), PO₄ (0.59±0.20 mg/l) and EC (566±32.1 µS/cm), these recorded values have no negative effects on the cultured *O. niloticus* (Meade, 1989; El-Sayed, 2006).

The total ammonia nitrogen (TAN) exists in water in two forms: ammonia (NH₃, or unionized ammonia) and ammonium (NH₄⁺, or ionized ammonia). NH₃ is more toxic than 300 - 400 time of NH₄⁺. Only the NH₃ form goes freely through fish gills and it is therefore considered the TAN's most toxic form. However, it was demonstrated that the toxicity of ammonia relatively depends on pH and temperature (Silva, Lima, Vale, & Marcelo, 2013). In aquaculture, high doses of ammonia either from excretion or from external pollution can cause reduction in fish growth or even death. The resistance of fish for toxicity of ammonia differs between species, however, rainbow trout *O. mykiss* was found to be the most sensitive to ammonia (LC50 values ranging between 0.068- 0.62 ppm), whereas Common carp *C. carpio* (LC50 values ranging between 0.43-2.1 ppt) and channel catfish *I. punctatus* (LC50 values ranging between 1-3.8 ppm) were the most resistant (Arthur, Corlis, Allen, & Hedtke, 1987). On the other hand, many authors cited that the *O. niloticus* can withstand up to 7.1 ppm of ammonia, while the optimum concentration for growth is <0.05 ppm (El-Shafey, 1998). Many authors reported that tilapia can

be counted as more tolerant species to ammonia and can withstand a very polluted environment, feeding on animal manure and even sewage sludge (Wong, 1989; EPA, 1998; Semra, 2014; Silva et al., 2013).

In the present study, the recorded values of NO₃ (1.96±0.60 mg/l), NO₂ (0.50±0.10 mg/l) and NH₃ (0.84±0.20 mg/l) were high in irrigation water of El-Mahmoudia Canal, however, Bahnasawy et al. (2003) studied the aquaculture ecosystem to evaluate the using of irrigation water of El-Salam canal (near Damietta City, Egypt) to culture *O. niloticus* in earthen ponds, and he found that the values of NO₃, NO₂ and NH₃ were 0.12-0.78, 0.06-0.60, and 0.142-0.570, respectively in irrigation water of El-Salam canal. However, during ponds preparations, we enhanced the oxygen content in pond culture water by aeration with two paddles for each pond for eight hours per day for two weeks. As a result of this process, the value of DO was increased to be 8.63±0.20 mg/l, and the values of NO₃ (1.38±0.24 mg/l), NO₂ (0.34±0.07 mg/l) and NH₃ (0.15±0.04 mg/l) in pond culture water at the beginning of *O. niloticus* culture were in the recommended limit for *O. niloticus* culture (Lawson, 1995 and Bahnasawy et al., 2003).

Plankton

Tilapia ecosystems and productivities are indirectly affected by the changes of water quality conditions, which can regulate the plankton productivity, biochemical composition and community in such tilapia earthen pond. These conditions fundamentally change due to the availability of nutrient concentrations. These conditions mainly depend on the initial and the following of fertilization (El-Otify, 2015). Plankton are relevant to the changes in the environmental conditions. Several studies carried out in freshwater ecosystems have established that the growth, composition and density of phytoplankton may be controlled by the limitation of nutrients, light and the abundance and composition of zooplankton

community (Basualto *et al.*, 2006).

Tilapia production may be increased by supplying of organic fertilizers in fish earthen ponds, as a result of increasing the primary live feeds productivity. Primary live feeds exist in earthen ponds can increase the quality and survival of tilapia larvae, fingerlings and fry. Therefore, using the manures, as a low cost fertilization technique, in ponds preparation is the most effective way to increase live feed production in earthen ponds and thereby increase the final fish production (Chib *et al.*, 2015).

In the present study, in order to obtain an optimal natural productivity level, the fertilization program composed of cow dung (during the preparation of ponds before start *O. niloticus* culture) and chicken manure (during *O. niloticus* culture) were used. To determine the possible success or failure of *O. niloticus* propagation system in earthen ponds, the management of plankton is very important (Mischke & Zimba, 2004). Ssanyu *et al.*, (2011) concluded that the relative status of plankton gives an indicator of water quality parameter and the possible success or failure of the aquaculture system of *O. niloticus*.

Quantification of phytoplankton as a primary production in terms of chlorophyll-*a* is alternative, easy and quick to measure phytoplankton biomass in aquaculture (Kundu & Jana, 1994). In the present study, we evaluate the phytoplankton pigments (chlorophyll-*a*, chlorophyll-*b* and total carotenoids) as an indicator for phytoplankton productivity and biomass. The results showed that the irrigation water of El-Mahmoudia Canal, Alexandria, Egypt, is considered a rich source of nutrient, pigments, phytoplankton productivity and biomass, which means that zooplankton, and consequentially cultured *O. niloticus*, will derive significant part of their immense from available phytoplankton, especially with the concentration of total carotenoids (2.374 ± 0.054 $\mu\text{g/g}$). On the other hand, the high recorded phytoplankton biomass may be due to high recorded concentration of NO_3 (1.96 ± 0.60 mg/l) in irrigation water of El-Mahmoudia Canal. Nitrate is the major form of nitrogen that is easily used by phytoplankton (El-Otify, 2015).

Protein is the most expensive component in the diets for aquatic species. One of the most important problems in aquaculture industry over the world, and especially in Egypt and developing countries, is the availability, cost, adulteration and bad treatments of protein during diet manufacture (GAFRD, 2012). Despite the efforts in the formulation development of starter diets, live food still remains a stable option in terms of survival and growth compared to formulated diet alone (Verreth, Storch, & Segner, 1987). One of the most effective factors which can evaluate the quality of natural live food for *O. niloticus* in earthen pond is the nutritional value of available plankton (Ssanyu *et al.*, 2011; and Mitra *et al.*, 2007). In the present study, in consideration of aquaculture ecosystem assessment, the evaluation the biochemical

composition (protein, carbohydrate, lipid and fatty acid profile) of total plankton exist in irrigation water of El-Mahmoudia Canal, as a water source for *O. niloticus* culture were investigated. Mitra *et al.* (2007) studied the biochemical composition of zooplankton community grown in freshwater earthen ponds, in the term of nutritional implication in nursery rearing of fish larvae and early juveniles and he found that protein, lipid and carbohydrate were ranging from 73–79%, 10.8–14.5% and 3.0–4.8%, respectively. Moreover, he reported that the content of SFA, MUFA and PUFA ranged from 64–81%, 7–15% and 6–20% of total fatty acids, respectively. On the other hand, the present study found that the plankton productivity (in dry weight form) was 4.281 ± 0.216 g dw/ton of El-Mahmoudia Canal irrigation water. There are no previous plankton studies performed on El-Mahmoudia Canal in Alexandria, in consideration to aquaculture ecosystem, however, the average of plankton dry weight reported by Mitra *et al.* (2007) ranged from 1.2 to 4.2 mg/l . The variation in biochemical composition (protein, carbohydrate, lipid and fatty acids) and productivity of freshwater plankton between our study and that observed by Mitra *et al.* (2007) may be due to many reasons, such as (1) water quality parameters, (2) nutrient limitation, (3) phytoplankton (availability, species and biochemical composition) and (4) zooplankton community and species structure (Vijverberg & Frank, 1976; Proulex & Nove, 1985; Kibria, Nugegoda, Fairclough, Lam, & Bradby, 1999; Basualto *et al.*, 2006). In aquaculture, species derive a substantial part of their dietary needs from available plankton as a valuable source of protein, amino acids, lipid, fatty acids and vitamins (Evjemo, Reitan, & Olsen, 2003). Therefore, plankton seems to provide a good source of immune and exogenous enzymes for larvae, however, the nutritional quality of plankton varies and plays a major role in producing high quality of fish larvae, juveniles and fry (Meeren, Olsen, Hamre, & Fyhn, 2008).

Another interesting investigated point in the current plankton study is zooplankton community. Investigations of zooplankton community in fish farm systems are important tools to evaluate the success of aquaculture ecosystem (Tavares *et al.*, 2010; Ssanyu *et al.*, 2011). On the other hand, fish density and nutrient supply are the two major factors affecting plankton communities in aquaculture ponds (Mitra *et al.*, 2007). In the present study, rotifers and copepods configured the main bulk of zooplankton community and they figured together 95.9% to the total zooplankton numbers. Many authors found that rotifers and copepods configured the main bulk of zooplankton communities in marine (Case *et al.*, 2008) or freshwater ecosystem (Ssanyu *et al.*, 2011), regardless to the type of cultured water, cultured fish, water quality and geographical location. However, the structure and functioning of the zooplankton community, regarding to species compositions and

densities of each species, are affected by several factors, such as (1) water quality aspects, (2) nutrient limitation, (3) phytoplankton (availability, species and community structure) and (4) predator types and densities (Tavares *et al.*, 2010; Ssanyu *et al.*, 2011).

Oreochromis niloticus Culture

Many authors cited that growth performance and feed utilization efficiency of *O. niloticus* were affected by different environmental factors in regarding to aquaculture ecosystem, such as water quality and live food quality and quantity (Gjedrem, 1997; and Workagegn, Ababbo, Yimer, & Amare, 2014). The average of ADG, SGR investigated in our results was higher than that observed by Bahnasawy *et al.* (2003), who studied the growth performance of *O. niloticus* cultured in earthen pond (with supplemental feed 25% protein at 50% satiation), in regard to enhance the natural feed (phytoplankton and zooplankton), it was 0.90 g/d and 1.55 %/day, respectively. Moreover, Goda, EWafa, El-Haroun, and Chowdhury (2007) found that SGR of *O. niloticus* ranging from 1.74 to 1.97 %/day, while Hassaan, Wafa, Soltan, Goda, and Mogheth (2014) found that SGR ranging from 2.98 to 3.10 %/day. The higher or lower IDG and/or SGR of *O. niloticus* recorded by different authors may be due to different initial weight, stoking density, offered live food and water quality (Bahnasawy *et al.*, 2003; Workagegn *et al.*, 2014). Feed conversion ratio FCR recorded in our results (0.96) was higher than that observed (0.82) by Bahnasawy *et al.* (2003), while protein efficiency ratio PER observed in our study (4.2) was lower (4.87) than Bahnasawy *et al.* (2003). On the other hand, El-Sayed, El-Ghobashy, and Al-Amoudi (1996) found that the FCR ranged from 2.2 to 3.15 in *O. niloticus* fed on artificial diets twice daily, while Goda *et al.* (2007) found that FCR of *O. niloticus* ranged from 1.70 to 2.00. The differentiation in feed utilization efficiency between the present study and that obtained by other authors may be due to the different growth performance, offered live food, protein offered in supplemental diets and water quality (Bahnasawy *et al.*, 2003).

Conclusions

Aquaculture productivity mainly depends on aquaculture ecosystems. Aquaculture ecosystems contain various living communities which constitute the biotic load of aquaculture pond. Natural productivity has the ability to increase this biotic load. Environmental assessment of water quality and natural productivity is required in order to determine how much aquaculture ecosystem can generate or produce using natural resources.

Although aquaculture is one of the most promising industries in Egypt, this sector, because of the Egyptian law, is not allowed to use the irrigation

water. In general, Egyptian freshwater aquaculture sector depends on agricultural drainage channels and/or groundwater. Our study strongly recommends that the irrigation water of El-Mahmoudia Canal is very successful and stable for aquaculture ecosystem of *O. niloticus*.

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References

- Arthur, J.W., Corlis, W.W., Allen, K.N., & Hedtke, S.F. (1987). Seasonal toxicity of ammonia to five fish and nine invertebrate species. *Bulletin of Environmental Contamination and Toxicology*, 38(2), 324-331. <https://dx.doi.org/10.1007/BF01606682>
- ASTM D421-85 (1998). Standard Practice for Dry Preparation of Soil Samples for Particle-Size Analysis and Determination of Soil Constants. ASTM International, West Conshohocken, US, American Society for Testing and Materials PA Press., 2pp. <https://dx.doi.org/10.1520/D0421-85R98>
- Bahnasawy, M.H., Abdel-Baky, T.E., & Abd-Allah, G.A. (2003). Growth performance of Nile Tilapia (*Oreochromis niloticus*) fingerlings raised in an earthen pond. *Archives of Polish Fisheries*, 11(2), 277-285.
- Basualto, S., Tapia, J., Cruces, F., Bertran, C., Shzatter, R., Pena-Cortes, F., & Hannstein, E. (2006). The effect of physical and chemical parameters on the structure and composition of the phytoplankton community of Lake Budi (IX Region, Chile). *Journal of the Chilean Chemical Society*, 51(3), 993-999. <https://dx.doi.org/10.4067/s0717-97072006000300015>
- Berzins, B. (1960). "Rotatoria" I- VI J. Conseil international pour l' Exploration de la Mer, Fiches d'Identification du Zooplankton, sheets 84-89, *International Council for the Exploration of the Sea (ICES)*, 27 pp.
- Bick, H. (1972). Ciliated Protozoa. An illustrated guide to the species used as biological indicators in freshwater biology. World Health Organization, Geneva, Switzerland, WHO Press., 198 pp.
- Ble, C.M., Etchian, O.A., Alla, Y.L., Adingra, A.A., Niamke, S., & Diopoh, J.K. (2012). Seasonal effects of the water quality in ponds on nutritive potential and digestibility of tilapia *Oreochromis niloticus* in natural feeding condition (Mali, West Africa). *African Journal of Agriculture Research*, 7(9), 1436-1441. <https://dx.doi.org/10.5897/AJAR11.1759>
- Bligh, E.G., & Dyer, W.J. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, 37(8), 911-917. <https://dx.doi.org/10.1139/o59-099>
- Case, M., Enide, E.L., Sigrid, N., Eneida, E.S., Ralf, S., & Antonio, T.M.J. (2008). Plankton community as an indicator of water quality in tropical shrimp culture ponds. *Marine Pollution Bulletin*, 56(7), 1343-1352. <https://dx.doi.org/10.1016/j.marpolbul.2008.02.008>
- Chib, S., Masud, S., & Proch, A. (2015). Effect of Different Doses of Poultry Manure and Cow Dung on the

- Growth of Indian Major Carp, *Catla catla*. *Journal of Animal Research*, 5(4), 833-837.
<https://dx.doi.org/10.5958/2277-940X.2015.00138.2>
- Dere, S., Gunes, T., & Sivaci, R. (1998). Spectrophotometric Determination of Chlorophyll-a, b and Total Carotenoid Contents of Some Algae Species Using Different Solvents. *Turkish Journal of Botany*, 22(1), 13-17.
- Dubois, M.K.A., Giles, J.K., Hamilton, P.A., Rebers, L., & Smith, F. (1956). Colorimetric method for determination of sugar and related substances. *Analytical Chemistry*, 28(3), 350-356.
<https://dx.doi.org/10.1021/ac60111a017>
- Dussart, B. (1969). Les copépodes des eaux continentales d'Europe occidentale, II: Cyclopoïdes et Biologie. Collection Faunes et Flores Actuelles. Academic Science, Paris, N. Boubee & Cie Press., 292 pp.
- Edmondson, W.T. (1959). Freshwater biology, 2nd Edition. New York, London, Sydney, John Wiley & sons, incorporation Press, 1248 pp.
- El-Otify, A.M. (2015). Evaluation of the physicochemical and chlorophyll-a conditions of a subtropical aquaculture in Lake Nasser area, Egypt. *Beni-Suef University Journal of Basic and Applied Sciences*, 4(4), 327-337.
<https://dx.doi.org/10.1016/j.bjbas.2015.11.009>
- El-Sayed, A.F.M. (2006). Tilapia culture. Wallingford, UK, CABI publishing Press., 277 pp.
<https://dx.doi.org/10.1079/9780851990149.0034>
- El-Sayed, A.F.M., El-Ghobashy, A., & Al-Amoudi, M. (1996). Effects of pond depth and water temperature on the growth, mortality and body composition of Nile tilapia, *Oreochromis niloticus* (L.). *Aquaculture Research*, 27(9), 681-687.
<https://dx.doi.org/10.1046/j.1365-2109.1996.00776.x>
- El-Shafey, A.A.M. (1998). Effect of ammonia on respiratory functions of blood of *Tilapia zilli*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 121(4), 305-313.
[https://dx.doi.org/10.1016/s1095-6433\(98\)10129-0](https://dx.doi.org/10.1016/s1095-6433(98)10129-0)
- EPA (Environmental Protection Agency) (1998). Update of Ambient Water Quality Criteria for Ammonia (Report No. EPA-822-R-98-008). Washington, DC, US, EPA Press., 148 pp.
- Meeren, T.V.D., Olsen, R.E., Hamre, K., & Fyhn, H.J. (2008). Biochemical composition of copepods for evaluation of feed quality in production of juvenile marine fish. *Aquaculture*, 274(2-4), 375-397.
<https://dx.doi.org/10.1016/j.aquaculture.2007.11.041>
- Evjemo, J.O., Reitan, K.I., & Olsen, Y. (2003). Copepods as live food organisms in the larval rearing of halibut larvae (*Hippoglossus hippoglossus* L.) with special emphasis on the nutritional value. *Aquaculture*, 227(1-4), 191-210. [https://dx.doi.org/10.1016/S0044-8486\(03\)00503-9](https://dx.doi.org/10.1016/S0044-8486(03)00503-9)
- GAFRD (General Authority for Fish Resources Development) (2012). Fisheries Statistics Year Book 2012, Cairo, Egypt, GAFRD Press., 300 pp.
- Gjedrem, T. (1997). Selective Breeding to Improve Aquaculture Production. *World Aquaculture*, 28(1), 33-45.
- Goda, A.S., EWafa, M., El-Haroun, E.R., & Chowdhury, M.A.K. (2007). Growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (Linnaeus, 1758) and tilapia galilae *Sarotherodon galilaeus* (Linnaeus, 1758) fingerlings fed plant protein-based diets, *Aquaculture Research*, 38(8), 827-837.
<https://dx.doi.org/10.1111/j.1365-2109.2007.01731.x>
- Grasshoff, G. (1976). Method of seawater analysis. New York, US, Verlag Chemie, Weinheim and New York Press., 317 pp.
- Gurney, R. (1927). Report on the Crustacea:—Copepoda and Cladocera of the Plankton. *Journal of Zoology*, 22(2), 139-172.
<https://dx.doi.org/10.1111/j.1096-3642.1927.tb00328.x>
- Gurney, R. (1931). British fresh-water Copepoda. Volume I. Ray Society, London, Adlard and Son, Limited Press., 238 pp.
<https://dx.doi.org/10.5962/bhl.title.82138>
- Hardig, J.P., & Smith, W.A. (1974). A key to the British freshwater Cyclopoïd and Calanoid Copepods. Scientific Publications No. 18, Freshwater Biological Association, UK, Cumbria Press., 56 pp.
- Hartree, E.F. (1972). Determination of protein: A modification of the Lowry method that gives a linear photometric response. *Analytical Biochemistry*, 48(2), 422-427.
[https://dx.doi.org/10.1016/0003-2697\(72\)90094-2](https://dx.doi.org/10.1016/0003-2697(72)90094-2)
- Hassaan, M.S., Wafa, M.A., Soltan, M.A., Goda, A.S., & Mogheth, N.M.A. (2014). Effect of Dietary Organic Salts on Growth, Nutrient Digestibility, Mineral Absorption and Some Biochemical Indices of Nile Tilapia; *Oreochromis niloticus* L. Fingerlings. *World Applied Sciences Journal*, 29(1), 47-55.
<https://dx.doi.org/10.5829/idosi.wasj.2014.29.01.81237>
- Hutchinson, G.E. (1967). A Treatise on limnology: Vol. II. Introduction to lake biology and the limnoplankton. New York, US, John Wiley and Sons Press., 1115 pp.
- Kibria, G., Nugegoda, D., Fairclough, R., Lam, P., & Bradbv, A. (1999). Utilization of wastewater-grown zooplankton: nutritional quality of zooplankton and performance of silver perch *Bidyanus bidyanus* (Mitchell 1838) (Teraponidae) fed on wastewater-grown zooplankton. *Aquaculture Nutrition*, 5(4), 221-227.
<https://dx.doi.org/10.1046/j.1365-2095.1999.00108.x>
- Kundu, G., & Jana, B.B. (1994). Influence of an atypical summer on the primary productivity of phytoplankton in two Tropical Ponds. *Internationale Revue der gesamten Hydrobiologie und Hydrographie*, 79(2), 249-257.
<https://dx.doi.org/10.1002/iroh.19940790210>
- Lawson, T.B. (1995). Fundamentals of Aquacultural Engineering. Boston, US, Springer-Verlag, MA Press., 355 pp. <https://dx.doi.org/10.1007/978-1-4613-0479-1>
- Meade, J.W. (1989). Aquaculture Management. Boston, US, Springer-Verlag, MA Press., 175 pp.
<https://dx.doi.org/10.1007/978-1-4615-6470-6>
- Mischke, C.C., & Zimba, P.V. (2004). Plankton community responses in earthen channel catfish nursery ponds under various fertilization regimes. *Aquaculture*, 233(1-4), 219-235.
<https://dx.doi.org/10.1016/j.aquaculture.2003.09.044>
- Mitra, G., Mukhopadhyay, P.K., & Ayyappan, S. (2007). Biochemical composition of zooplankton community grown in freshwater earthen ponds: Nutritional implication in nursery rearing of fish larvae and early juveniles. *Aquaculture*, 272(1-4), 346-360.

- <https://dx.doi.org/10.1016/j.aquaculture.2007.08.026>
- Myklestad, S., & Haug, A. (1972). Production of carbohydrates by the marine *Chaetoceros affinis* var. *willei* (Gran.) Hustedt. I. Effect of the concentration of nutrients in the culture medium. *Journal of Experimental Marine Biology and Ecology*, 9(2), 125-136.
[https://dx.doi.org/10.1016/0022-0981\(72\)90041-x](https://dx.doi.org/10.1016/0022-0981(72)90041-x)
- Olguin, E., Galicia, S., Angulo-Guerrero, O., & Hernandez, E. (2001). The effect of low light flux and nitrogen deficiency on the chemical composition of *Spirulina* sp. (*Arthrospira*) grown on digested pig waste. *Bioresource Technology*, 77(1), 19-24.
[https://dx.doi.org/10.1016/S0960-8524\(00\)00142-5](https://dx.doi.org/10.1016/S0960-8524(00)00142-5)
- Pawlowski, L. (1994). Standard methods for the examination of water and wastewater, 18th edition. *Science of The Total Environment*, 142(3), 227-228.
[https://dx.doi.org/10.1016/0048-9697\(94\)90332-8](https://dx.doi.org/10.1016/0048-9697(94)90332-8)
- Proulx, D., & Nove, J. (1985). Growth of *Daphnia magna* on urban wastewater tertiary treated with *Scenedesmus* sp., *Aquacultural Engineering*, 4(2), 93-111.
[https://dx.doi.org/10.1016/0144-8609\(85\)90008-1](https://dx.doi.org/10.1016/0144-8609(85)90008-1)
- Radwan, S.S. (1978). Coupling of two dimensional thin layer chromatography for the quantitative analysis of lipid classes and their constituent fatty acids. *Journal of Chromatographic Science*, 16(11), 538-542.
<https://dx.doi.org/10.1093/chromsci/16.11.538>
- Rauch, T. (1981). The estimation of microalgal protein content and its meaning the evolution of algal biomass, comparison method for extracting for protein. *Hydrobiologia*, 78(3), 237-251.
<https://dx.doi.org/10.1007/bf00008520>
- Ross, L.G. (2000). Environmental physiology and energetics. In: Beveridge M.C.M., McAndrew B.J. (eds) Tilapias: Biology and Exploitation. *Fish and Fisheries Series*, 25, 89-128.
https://dx.doi.org/10.1007/978-94-011-4008-9_4
- Sars, G.O. (1911). An account of Crustacea of Norway, Copepoda, Harpacticoida. Bergen Museum, Skrifter, *Bergen*, 5, 1-449. <https://dx.doi.org/10.1038/088276a0>
- Sars, G.O. (1918). An account of Crustacea of Norway, Copepoda, Cyclopoida. Bergen Museum, Skrifter, *Bergen*, 6, 1-225. <https://dx.doi.org/10.1038/102304a0>
- Sars, G.O. (1926). Freshwater Ostracoda from Canada and Alaska. *Report of the Canadian Arctic Expedition, 1913-1918*, 7(1), 1-23.
<https://dx.doi.org/10.5962/bhl.title.64194>
- Semra, K.K. (2014). Acute toxicity of ammonia to blue tilapia, *Oreochromis aureus* in saline water. *African Journal of Biotechnology*, 13(14), 1550-1553.
<https://dx.doi.org/10.5897/AJB12.1547>
- Silva, F.J.R., Lima, F.R., Vale, D.A., & Marcelo, V.C. (2013). High levels of total ammonia nitrogen as NH_4^+ are stressful and harmful to the growth of Nile tilapia juveniles. *Acta Scientiarum Biological Sciences*, 35(4), 475-481.
<https://dx.doi.org/10.4025/actascibiolsci.v35i4.17291>
- Soliman, N.F., & Yacout, D.M. (2016). Aquaculture in Egypt: status, constraints and potentials. *Aquaculture International*, 24(5), 1201-1227.
<https://dx.doi.org/10.1007/s10499-016-9989-9>
- Ssanyu, G.A., Rasowo, J., Auma, E., & Ndunguru, M. (2011). Evaluation of Plankton Community Structure in Fish Refugia Acting as *Oreochromis niloticus* Propagation and Nursery Units for Rice/Fish Trials, Uganda. *Journal of Aquaculture Research & Development*, 2(4), 100-116.
<https://dx.doi.org/10.4172/2155-9546.1000116>
- Tavares, L.H.S., Millan, R.N., & Santeiro, R.M. (2010). Characterization of a plankton community in a fish farm. *Acta Limnologica Brasiliensia*, 22(1), 60-69.
<https://dx.doi.org/10.4322/actalb.02201008>
- Verreth, J., Storch, V., & Segner, H. (1987). A comparative study on the nutritional quality of decapsulated Artemia cysts, micro-encapsulated eng diets and enriched dry feeds for *Clarias gariepinus* (Burchell) larvae. *Aquaculture*, 63(1-4), 269-282.
[https://dx.doi.org/10.1016/0044-8486\(87\)90078-0](https://dx.doi.org/10.1016/0044-8486(87)90078-0)
- Vijverberg, J., & Frank, T.H. (1976). The chemical composition and energy contents of copepods and caladocerans in relation to their size. *Freshwater Biology*, 6(4), 333-345.
<https://dx.doi.org/10.1111/j.1365-2427.1976.tb01618.x>
- Wong, M.H. (1989). Toxicity test of landfill leachate using *Sarotherodon mossambicus* (Freshwater Fish), *Ecotoxicology and Environmental Safety*, 17(2), 149-156.
[https://dx.doi.org/10.1016/0147-6513\(89\)90033-X](https://dx.doi.org/10.1016/0147-6513(89)90033-X)
- Workagegn, K.B., Ababbo, E.D., Yimer, G.T., & Amare, T.A. (2014). Growth performance of the Nile Tilapia (*Oreochromis niloticus* L.) fed different types of diets formulated from varieties of feed ingredients. *Journal of Aquaculture Research & Development*, 5(3), 1-4.
<https://dx.doi.org/10.4172/2155-9546.1000235>