Combined Effect of Papain and Vitamin-C Levels on Growth Performance of Freshwater Giant Prawn, *Macrobrachium rosenbergii*

S.M. Manush¹, P.P. Srivastava^{1,2,*}, M.P.S. Kohli³, K.K. Jain¹, S. Ayyappan⁴, S.Y. Metar¹

¹Central Institute of Fisheries Education (Deemed University), Department of Nutrition and Biochemistry, Indian Council of Agricultural Research, Fisheries University Road, Seven Bungalows, Versova, Mumbai - 400 061 (M.S), India.

²National Bureau of Fish Genetic Resources, Molecular Biology and Biotechnology Division, Canal Ring Road, Teli Bagh, Lucknow -226 002, India.

³Central Institute of Fisheries Education (Deemed University), Indian Council of Agricultural Research, Fisheries University Road, Seven Bungalows, Versova, Mumbai - 400 061 (M.S), India.

⁴Director General, Indian Council of Agricultural Research, Krishi Bhawan, New Delhi-110 002, India.

* Corresponding Author: Tel.: +91.22 26361446; Fax: +91.22 26361446;	Received 16 July 2012
E-mail: ppsicar@gmail.com	Accepted 22 July 2013

Abstract

Present study was conducted to evaluate the combined effect of proteolytic enzyme, papain and vitamin-C on freshwater giant prawn, *Macrobrachium rosenbergii*. Post-larvae of *Macrobrachium rosenbergii* of average weight 9.5-16.3 mg initial weight were fed with eight different diets supplemented with papain 0.0%, 0.108%, 0.3%, 0.5%, 0.0%, 0.108%, 0.3%, 0.5% and vitamin-C 0.0%, 0.0%, 0.0%, 0.2%, 0.2%, 0.2% and 0.2% for feeds F1 to F8 respectively. Before the start of the experiment, prawns were reared using papain and vitamin-C free diet in order to deplete their vitamin-C stores. The same diet formulation was used in all treatments containing soybean meal, fish meal, shrimp meal and wheat flour as the major ingredients. Feeding level was kept same for all the groups of prawn and fed to satiation. Protease specific activity after one hour of feeding at the end of the experiment was estimated, which have shown a higher value in comparison to the protease specific activity one hour before feeding at the beginning of the experiment. The results obtained after feeding trial demonstrated improved growth (P<0.05) at higher concentration (F7 and F8). However, the growth of post-larval stages of prawn fed only vitamin-C or papain showed lesser growth than those of the combined effect. Survival was recorded between 46-56% in all the treatments. No growth reduction or deficiency signs were observed. This finding is the first report of its kind where a combined effect of papain and vitamin-C showing a synergistic effect on the growth performance. Further standardization and commercialization of this finding, through economically viable methods, is needed to improve the growth performance of this premium commodity.

Keywords: Macrobrachium rosenbergii, papain, vitamin-C, protease, growth.

Introduction

In intensive prawn culture operations, one of the foremost requirements is the availability of properly formulated practical feeds for different growth stages of prawn. The efficacy of the diet not only depends on the nutrient composition and nutrient balance, but also on the effective utilization of the nutrients by the animal. In the utilization of dietary nutrients, the digestive enzymes play a vital role in catalyzing the hydrolytic reactions splitting the macromolecules into simple absorbable form of molecules. The activity of these biocatalysts is regulated by many physical, chemical and biological factors and thus a shift from these optimum conditions necessary for these affect their enzymes may activity, thereby correspondingly modify the digestibility of nutrients supplied to the animals (Hemambika and Paul Raj, 1993). There are several reports on the use of exogenous proteolytic enzyme in post-larval stages of

Penaeus monodon. (Hong-Yung and Hsian-Fu, 1990, Fang and Lee, 1992). It is very well reported that proteolytic enzyme of exogenous origin plays an important role in feed digestibility in fishes like utilizing trypsin (Dabrowski and Glogowski, 1977a), papain (Srivastava et al., 1994) in common carp diet. Protease activity in the digestive tract is a key determinant of the digestibility and assimilation efficiency of ingested proteins. Dabrowski (1984) suggested that incompletely found digestive tract contain insufficient digestive enzymes to completely digest dry feeds. According to several authors, enzymes from live animals contributed to the digestion process by autolysis or zymogen activation (Janacarik, 1964; Dabrowski and Glogowski, 1977a; Kolkovski et al., 1993; Person-Le Ruyet et al., 1993). In fish and Invertebrates, some mammalian digestive protease-like enzymes have been described (Ikeda et al., 1986; Gildberg, 1998).

Vitamin-C is an indispensable nutrient required

[©] Published by Central Fisheries Research Institute (CFRI) Trabzon, Turkey in cooperation with Japan International Cooperation Agency (JICA), Japan

to maintain the physiological processes of different animals (Tolbert, 1979). Fishes and crustaceans are incapable of biosynthesis of ascorbic acid since they do not have the enzyme L-gulonolactone oxidase, which is responsible for synthesis of vitamin-C. (Wilson, 1973). In crustaceans, vitamin-C influences the alkaline phosphatase activity during synthesis of chitin (Paul Raj, 1997).

A dietary requirement for vitamin-C has been reported for several species of crustaceans (Tacon, 1987). Crustaceans fed diets deficient in vitamin-C develop melanized lesions distributed throughout the collagenous tissue underlying the exoskeleton, decolourization and abnormal colourization and mortality (Deshimaru and Kuroki, 1979; Lightner et al., 1979, Heinen, 1984; Shiguein and Itoh, 1988). Most aquatic animals including shrimps-prawn require a dietary source of vitamin-C to prevent the development of deficiency symptoms; such as melanized lesions throughout the collagenous tissue underlying the exoskeleton, reduced growth, poor wound healing capacity and eventually, mortality (Hunter et al., 1979; Margarelli et al., 1979; He and Lawrence, 1993; Shiau and Hsu, 1994). Till date, the ascorbic acid requirements of farmed species of shrimps have been studied only from juvenile stage onwards (Merchie et al., 1995). Quantitative estimates of vitamin-C requirement reported for crustaceans include 5000-10000 mg.kg⁻¹ of diet for Penaeus japonicus (Guary et al., 1976) 1000-2000 mg.kg⁻¹ diet for P. califorensis and P. stylirosris (Lightner et al., 1979) and 215 to 430 mg.kg⁻¹ diet for Penaeus japonicus (Shiguein and Itoh, 1988). Recommended dietary ascorbic acid (AA) levels for shrimp using ascorbic acid-polyphosphate and crystalline ascorbic acid are 20 and 120-130 mg AA.kg⁻¹ for the postlarvae of tiger shrimp, Penaeus monodon and white shrimp, Penaeus vannamei respectively (He and Lawrence, 1993; Lavens et al., 1998, 1999). Reported vitamin-C requirements for crustaceans are generally at least ten fold higher than those reported for various species of fish (Tacon, 1987). The estimated dietary requirement for Macrobrachium rosenbergii falls with in the range (60 -150 mg) vitamin-C activity per kg of diet reported for several species of fish, and found in tissue of clams and adult brine shrimp as calculated by Conklin (1998).

Considerable work has been done on the utilization of exogenous enzymes in fish and shellfish diet (Janacarik, 1964; Dabrowski and Glogowski, 1977a,b; Kolkovski *et al.*, 1993; Person-Le Ruyet *et al.*, 1993) and also on dietary supplementation of vitamin-C, which are evident from various reports. (Merchie *et al.*, 1997a; Blom *et al.*, 1999; Adham *et al.*, 2000), but none of the reports are available on the combined effect of supplemented proteolytic enzyme, papain and vitamin-C, neither in fish nor in crustaceans.

Thus, the present study has been undertaken to evaluate both the individual and combined effect of

dietary incorporation of papain and vitamin-C in postlarval stage of *Macrobrachium rosenbergii* in terms of growth, deposition of nutrients in terms of flesh and survival. Here, selected digestive enzymes activities in post-larvae before and after feeding the diets with and without dietary proteolytic enzyme, were assessed.

Materials and Methods

Experimental Setup

The experiment was carried out using freshwater giant prawn post-larvae with average size between 9.4±0.92 and 16.3±5.43 mg over a period of 40 days at Aquafeed laboratory, Department of Fish Nutrition and Biochemistry, Central Institute of Fisheries Education, Mumbai, India. It was set up in 8 distinct groups each with 3 replicates. Uniform sized plastic pools of 50 liters capacity were used for the experiment. All the pools were cleaned and filled with bore-well water up to 30 liters. Each of the pools was stocked with 25 prawns at 26 + 1°C for a period of 40 days. Aeration was provided throughout the period with 2HP air blower. Experimental pools were supplied with chlorine-free bore-well water with a flow rate 1 liter.min⁻¹ throughout the experimental period. Each of the pools was covered with perforated cover to prevent the animals from jumping out. Physico-chemical parameters were monitored and recorded. Poly-Vinyl-Chloride pipes were used as hideout for prawns. The post-larvae were starved for a day before taking the initial body weight. Weighing of prawns was carried out in an interval of 20 days to assess the growth during the experimental period of 40 days. Prawns were anesthesized with Tricaine methane- sulfonate (MS-222) @ 50 mg/L.

Experimental Diet

Papain (extracted from Carica papaya), Betaine Hydrochloride and L (+) Ascorbic acid were procured from E-MERCK Darmstadt, Germany. Nachini (local name) was used as natural feed attractant. Details of the diet components were given in Table 1. Diet F1 was used as control. All the ingredients were dried in hot air oven at 80°C for 8 hours, powdered and sieved through a 60 µm mesh size nylon netting to get a powder. All the ground ingredients used for basal diet were analyzed for their crude protein content before preparing the feed. Accordingly, eight isonitrogenous diets were prepared to keep approximately 38% protein content and approximately 8% fat. All the ingredients were mixed thoroughly with required amount of water to make dough, which were passed through a twin-screw extruder having a die of 2 mm size (Basic Technology Pvt. Ltd, Kolkata, W. B.). Pellets obtained were dried at 60°C in a hot air oven overnight and stored in a cool place till their use. Feeding were done up to satiation and was adjusted

Ingredients				Inclusior	n rate (%)			
	F_1	F_2	F_3	F_4	F_5	F_6	F ₇	F ₈
Soybean meal	30.0	30.0	29.9	29.9	29.9	29.9	29.9	29.8
Fish meal	29.0	29.0	28.9	28.9	30.0	28.9	28.9	28.8
Acetes	15.0	15.0	15.0	14.8	16.0	15.0	14.8	14.8
Wheat flour	10.4	10.28	10.3	10.3	10.3	10.28	10.3	10.28
Cod liver oil	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0
Carrot powder	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0
Soya lecithin	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
Vitamin mixture*	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0
Nachini	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Sucrose	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Betaine	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1
Papain	-	0.108	0.3	0.5	-	0.108	0.3	0.5
Vitamin- C	-	-	-	-	0.2	0.2	0.2	0.2
Protein	38.3	37.9	39.1	38.2	37.8	38.5	40.1	39.2
Fat	8.21	8.88	7.11	6.14	7.94	7.76	6.96	7.04
Fibre	10.94	8.04	9.16	8.95	8.15	11.07	7.85	10.84
Moisture	8.3	7.8	7.6	7.1	8.0	8.3	7.5	8.0
Ash	15.34	15.25	13.73	12.33	15.75	13.32	12.56	14.72

Table 1. Composition of experimental diets (% DM basis)

*Composition of vitamin mineral mixture: Vitamin A: 20,00,000 I. U., Vitamin D_3 :4,00,000 I.U., Vitamin B_2 : 0.8 g, Vitamin E: 300 I.U., Vitamin K: 0.4 g, Calcium Pantothenate: 1.0 g, Nicotinamide: 4.0 g, Vitamin B_{12} : 2.4 g, Choline chloride: 64.0 g, Calcium: 300.0 g, Manganese: 11.0 g, Iodine: 0.4 g, Iron: 3.0 g, Zinc: 6.0 g, Copper: 0.8 g, Cobalt: 0.16 g (Source: Sarabhai Chemicals, Baroda, INDIA)

on daily observation of feed intake of post-larvae by visual estimation.

Crude fat in the feed was estimated by Soxtec system (Model HT2, 1045 extraction unit Foss Tecator, Sweden) using diethyl ether as a solvent (boiling point 55°C), dry weight by drying at 1050 C for 6 hours, ash by burning for 6 hours at 600°C, crude fibre by the method suggested by Bennik (1994), using Fibretec system (Model 1017 Hot Extraction, Foss Tecator, Sweden). Carcass protein was estimated by the method of Lowry *et al.* (1951). Bovine serum albumin was used as the standard. Nitrogen in the feed was estimated by Micro Kjeldahl method using Foss Tecator 2200 Kjeltec system and the crude protein is calculated as N x 6.25, as suggested by Chang (1994).

Digestibility of dietary protein by post-larvae was assessed by protease assay developed by Peterson (1977). Tyrosine calibration curve was used to evaluate the data. The results of the specific activity are given in µg of liberated tyrosine mg⁻¹ of dissolved protein.minute⁻¹. Lipase activity was found out by Cherry and Crandall (1932), using olive oil emulsion.

Data were processed for Analysis of Variance (ANOVA) and significance were tested for various growth parameters and survival by the method described by Snedocor and Cochran (1961).

Results and Discussion

Composition of Diet

The proximate composition of different experimental diets is given in the Table 1. The estimated crude protein of different experimental diets

varied from 37.8-40.1%. It is reported that required level of protein in post-larvae is a little higher than the grow-out stages. However, the optimum requirement of crude protein in the diet for Macrobrachium rosenbergii is found to be in the range 23.8-38.5% (Corbin et al., 1983), 30-35% (D'Abramo and Reed, 1988; Freuchtenicht et al., 1988). The ether extract in the experiment was found to be in the range of 6.14% to 8.88%, which is in the optimum range for Macrobrachium rosenbergii PL as suggested by D' Abramo and Sheen (1989) and Sebastian (1996). In the present study, the fibre content was in the range from 7.85% to 11.07% as suggested by Sebastian (1996). Fair et al. (1980) reported that the incorporation of crude fibre in to the diet up to 30 % was showing good result in terms of growth performance. The soluble carbohydrates of different experimental diets were in the range of 18.91% to 27.28%. For better growth performance in Macrobrachium rosenbergii, optimum range of soluble carbohydrate in the feed was found to be up to 40% (Briggs, 1991).

Enzyme Activity

The specific protease activity at the beginning of the experiment was found to be 0.279 IU. μ g⁻¹ protein (F4) to 0.4316 IU. μ g⁻¹ protein (F8) and 0.4279 IU. μ g⁻¹ protein (F1) to 0.7036 IU. μ g⁻¹ protein (F7) at the end of the experiment as shown in the Figure 1. The lipase activity at the beginning of experiment was found to be 0.005 IU. μ g⁻¹ lipid (F2 and F3) to 0 0.015 IU. μ g⁻¹ lipid (F6) and that at the end was found to be 0.0.15 IU. μ g⁻¹ lipid (F1 and F5) 0 0.015 IU μ g⁻¹ lipid (F6 and F8) as shown in Figure 2. The effect of exogenous

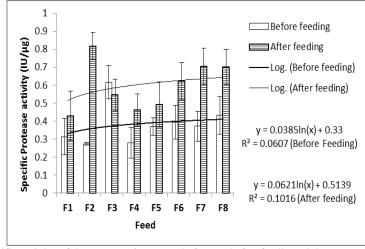


Figure.1. Protease specific activity of the *M. rosenbergii* PL before and after feeding trial.

digestive enzymes on the endogenous digestive enzyme activities of the shrimp, however, has been less clear and quantitative correlation between digestive enzyme activities and shrimp growth is also not conclusive (Dabrowski, 1979; Maugle et al., 1983 b). It is not clear whether shrimp growth could be positively correlated to the digestive enzyme positively. Many studies, however, have failed to support the correlation (Maugle et al., 1983b; Lee et al., 1984). The use of compounded diets to replace Artemia nauplii and other live food organisms for the larval and post-larval Penaeus monodon rearing has become popular. However, the quantity of exogenous enzymes received through them is decidedly lower. Maugle et al. (1983a,b) have reported that studies of diets with micro encapsulated amylase and bovine trypsin have shown increased growth in Penaeus japonicus juveniles. The present study concerns to evaluate the effect of incorporation of proteolytic enzyme, papain in Macrobrachium rosenbergii postlarval stage through diet on growth and survival and the effect of these feed supplements on the digestive enzyme activities of the post-larval prawn. Results reveal that there is an increase in growth performance as well as elevation in protease activity. Total protease activities, however, reflected differences associated with enzyme supplements, while the difference in lipase activity could not be correlated. The concentration of protease in the total tissue homogenate was elevated. This result has similarity with findings of Janacarik (1964); Dabrowski and Glogowski (1977a); Kolkovski et al. (1993) and Person-Le Ruyet et al. (1993) gives concurrence to the present findings. Contrary to this, (Kolkovski, et al., 2000) reported reduced growth while supplementing exogenous source of enzyme. The lipase activity in the tissue homogenate was found increased but it has no correlation in the body weight gain. It is known that dietary compositions have obvious influence on digestive enzyme activities. Lee (1984) concluded that the protein levels et al. influenced enzyme activity in Penaeus vannamei of all sizes, while the protein source had a greater influence on the enzyme activities in shrimp less than 10 g.

Growth Parameters

The body weight of experimental treatments recorded at 20 days intervals have been given in Figure 3. A trend of higher body weight gain was observed in F7 treatments compared to other treatments at the end of the experiment (60.75±1.93 mg). Comparative growth parameters of different experimental groups are as shown in Table 2. The absolute growth rate of different experimental treatments was found to be in the range of 9.8±0.3 mg and 48.8±5.15 mg. The highest and the lowest values were recoded in F7 and F2 respectively. The difference was found to be significant (P<0.05). Lobao et al. (1995) reported that the experiments in Macrobrachium rosenbergii with the diets containing vitamin-C @ 3 g.Kg⁻¹ have shown best growth performance. It is suggested that, on adding microbial enzyme in prawn diet (1 mg.g-1), the growth was found to increase by 13.87% (Zhong-Jhun, et al., 1994). The relation of feed intake and weight gain, i.e. FCR of different treatments were recorded in the range of 2.07 (F7 treatment) to 3.5 (F2 treatment), the difference was not statistically significant (P>0.05). The highest value of protein efficiency ratio of different experimental treatments (PER) was recorded in F2 treatment (0.45±0.04) while the lowest was observed in F3 treatment (0.15 ± 0.02) . The difference between the treatments was found to be significantly different (P<0.05). The survival (%) of the post-larvae of different treatments has been presented in Table.2 and Figure 4. The highest survival was recorded in F4 treatment (56%) whereas the lowest was recorded in F3, F7 and F8 treatment (46%), however, the survival of different groups did not differ significantly (P>0.05). Low survival reported in the experimental trial can be due to the stress during handling and weight measurement.

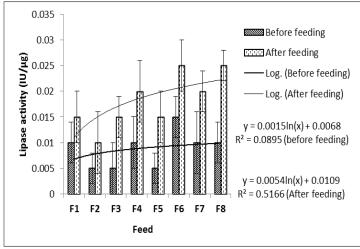


Figure 2. Lipase activity of the Macrobrachium rosenbergii PL before and after feeding trial.

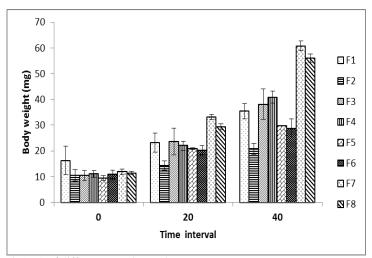


Figure 3. Body weight gain(mg) of different experimental groups.

Table 2. Comparative growth parameters for different experimental groups

	F_1	F_2	F_3	F_4	F_5	F ₆	F_7	F ₈
Per day increment (mg)	0.47±0.3 ^a	$0.24{\pm}0.009^{a}$	0.69±0.3 ^a	0.74 ± 0.15^{a}	0.50 ± 0.04^{a}	0.44 ± 0.08^{a}	1.22±0.1 ^b	1.1±0.09 ^b
Specific growth rate (%)	2.34±1.8 ^a	1.7±0.43 ^a	3.26±1.2 ^a	3.29±0.6 ^a	2.8±0.3ª	2.4±0.1 ^a	4.09±0.4 ^a	3.9±0.2 ^a
Absolute growth (mg)	19.1±5.2 ^a	9.8±0.3 ^a	27.7±13.5 ^a	29.8±6.2 ^a	$20.2{\pm}1.6^{a}$	17.8±3.3 ^a	48.8±5.15 ^b	44.6±3.8 ^b
FCR	$2.8{\pm}0.4^{a}$	3.5±0.1 ^a	2.12±0.01 ^a	2.34±0.1ª	$2.97{\pm}0.4^{a}$	3.1±0.6 ^a	2.07 ± 0.08^{a}	2.77±0.1 ^a
FER	0.36±0.0 ^a	$0.28{\pm}0.01^{a}$	$0.47{\pm}0.002^{b}$	$0.43{\pm}0.02^{b}$	0.35 ± 0.04^{ab}	0.33±0.06 ^a	0.48 ± 0.02^{b}	0.36±0.01 ^b
PER	0.23±0.09 ^a	$0.45{\pm}0.04^{b}$	$0.15{\pm}0.02^{a}$	$0.22{\pm}0.07^{a}$	$0.24{\pm}0.05^{a}$	0.26 ± 0.04	0.18±0.03 ^a	$0.26{\pm}0.0^{a}$
Survival	48±4.16 ^a	52±2.13 ^a	46±1.15 ^a	56±2.3ª	48±2.3ª	48±1.7 ^a	46±3.05 ^a	46±1.15 ^a

Superscripts in a row with different alphabets indicate significant difference (P<0.05).

Requirement of Ascorbic Acid

According to Merchie *et al.* (1997b), higher levels of ascorbic acid levels are required in postlarvae during metamorphosis than in larval stages. However, vitamin-C requirement of juvenile freshwater prawns is much lower than previously reported for other crustaceans. The lower requirement is more likely attributed to the source of vitamin-C used rather than interspecific differences. Previous reports of vitamins-C requirements of crustacean species were based upon the studies that used sources that are less stable and more subject to leaching, circumstances likely leading to over estimates of the requirement. Moreover, the estimated dietary quantitative requirement for *Macrobrachium rosenbergii* falls in the range (60-150 mg Vitamin-C activity.kg⁻¹ diet) reported for several species of fish,

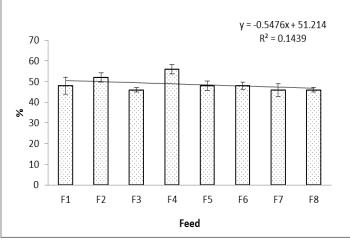


Figure. 4. Survival rate in percentage of different experimental groups during the feeding trial.

and found in tissue of clams and adult brine shrimp as calculated by Conklin (1998) from published values. In the present feeding trial, we have incorporated high dose of vitamin C to make sure the availability of ascorbic acid to the animal. D' Abramo *et al.* (1994) suggested that by adding high level of L- crystalline ascorbic acid in the diet would be sufficient enough to compensate for the assumed rapid loss due to high rates of leaching. The low requirement in comparison with the supplement suggest that even if 98% of the crystalline Vitamin-C were lost due to leaching prior to the time before consumption, a sufficient amount would remain to satisfy the requirement.

Field trials on this study in pond conditions are required for further standardization of this work. Later, this enzyme application in crude form in combination with vitamin-C can be suggested to the farmers for better feed conversion ratio of feeds used.

Acknowledgements

The authors are grateful to the Director, Central Institute of Fisheries Education, Mumbai for providing facilities for this work. Thanks are also due to Dr. R.S. Biradar and Mr. G.K. Rao for their help in statistical analysis.

References

- Adham, K.G., Hashem, H.O., Abu-Shabana, M.B. and Kamel, A.H., 2000. Vitamin-C deficiency in catfish *Clarias gariepinus*. Aquaculture Nutrition, 6: 129-139. doi: 10.1046/j.1365-2095.2000.00139.x
- Bennik, M. R., 1994. Fibre analysis. In: Introduction to chemical analysis of Foods - Suzanne, S. and Neilsen (Eds), 109 pp.
- Blom, J. H., Dabrowski, K. Rapp, J. D., Sakakura, Y. and Tsukamoto, 1999. Competition for space and food in Rainbow trout Onchorhynchus mykiss as related to ascorbic acid status, Aquaculture, 180: 79-87. doi: 10.1016/S0044-8486(99)00189-1
- Briggs, M. R. 1991. The performance of juvenile prawns, Macrobrachium rosenbergii, fed a range of

carbohydrate source in semi purified diets. Journal of World Aquaculture. Society., 22(3): 16 A.

- Chang, S.K.C., 1994. Protein analysis. In: Introduction to chemical analysis of Foods- Suzanne, S. and Neilsen (Eds) 207 p.p.
- Cherry and Crandal, 1932. Determination of serum lipase activity, American. Journal of Physiology., 100: 266 p.p.
- Conklin, L.P. 1998. Vitamin-C: A new pathway from old antioxidant, Trends in Plant science, Research news, 3(9): 300-301.
- Corbin, J. S., Fugimoto, M. N. and iwaiIwai, T. Y. 1983. Feeding practices and nutritional consideration for *Macrobrachium rosenbergii* culture in Hawaii. Handbook of Mariculture, 1: Crustaceans Aquaculture. CRC Press Inc. Boca Rocton. Florida, 391 p.p.
- D' Abramo, I.K. and Reed, L. 1988. Optimum dietary protein level for juvenile freshwater prawn *Macrobrachium rosenbergii*. 19th annual meeting of the World Aquaculture Society, Hanolulu, Hawaii, 27 p.p.
- D' Abramo, I. K. and Sheen, S.S. 1989. Essential fatty acid requirement of freshwater prawn *Macrobrachium rosenbergii*. 19th annual meeting of the world Aquaculture society, Los Angeles, California, 35 p.p.
- D' Abramo, L.R., Moncreiff, C.A., Holcomb, F.P., Montanez, J.L. and Buddangton, R.K. 1994. Vitamin-C requirement of Juvenile fresh water prawn, *Macrobrachium rosenbergii*. Aquaculture, 128: 269-275.
- Dabrowski, K.,1984. The feeding of fish larvae: present "state of the art" and perspectives, Reproduction, nutrition and development, Special publication, European Mariculture Society, 24: 807-833.
- Dabrowski, K., 1979, The role of proteolytic enzymes in fish digestion, In: E.Jaspers, and G.Persoone, (Eds) Cultivation of fish fry and its live food, Special publication European. Mariculture. Society. Special publication, 4:107-126.
- Dabrowski, K. and Glogowski, J. 1977a. Studies on the role of exogenous proteolytic enzymes in digestion process in fish. Hydrobiologia, 54(2): 129-134.
- Dabrowski, K. and Glogowski, J. 1977b. A study of application of proteolytic enzymes to fish food. Aquaculture, 12 : 349-360.

- Deshimaru, O. and Kuroki, K. 1979. Studies on a purified diet for prawn. VII. adequate dietary levels of ascorbic acid and Inositol. Bulletin. of Japanese. Society Science Fisheries., 42: 571-576.
- Fair, P.H., Fortner, A.R., Millikin, M.R. and Sick, L. V. 1980. Effects of dietary activity of the prawn, *Macrobrachium rosenbergii*, Proceedings. of World Mariculture Society., 11: 369-381.
- Fang, L.S. and Lee, B.N., 1992. Ontogenic change of digestive enzymes in *Penaeus monodon*. Comparative. Biochemistry and Physiology., 103B: 1033-1037.
- Freuchtenicht, G.W., Barak, I.E., Malech. S. R. and Stanley, R. W., 1988. The effect of protein level in feed on growth performance of freshwater prawn, *Macrobrachium rosenbergii* individually reared clear water flow through aquaria. Presented at the 19th Annual meeting of the World Aquaculture Society, January 2-9. Honolulu, Hawaii.
- Gildberg, A. 1998. Aspartic proteinases in fishes and aquatic invertebrates, Comparative. Biochemistry and Physiology., 91B: 425-435.
- Guary, M., Kanazawa, A., Tanaka, N. and Ceccaldi, H.J. 1976. Nutritional requirements of prawn VI: Requirements of ascorbic acid. MEM, Faculty Fisheries. Kagoshima University., 25(1): 53-57.
- He, H. and Lawrence, A.L. 1993. Vitamin-C requirements of the shrimp *Penaeus vannamei*, Aquaculture, 114: 305-316.
- Hemambika, M. and Paul Raj, R. 1993. studies on the digestive enzymes of the Indian white prawn *Penaeus indicus* H. Milne Edwards. Central Marine Fisheries Research Institute Special. Publication, 56: 88-94.
- Heinen, J.M. 1984. Nutritional studies on the giant asian prawn, *Macrobrachium rosenbergii*. PhD. Dissertation, Boston University, 124 pp.
- Hong-Yung, C. and Hsian-Fu, L. 1990. The effects of exogenous digestive enzymes on the growth of early post-larval *Penaeus monodon*, R.Hirano and I.Hanyu, (Eds). The Second Asian Fisheries Forum, Asian Fisheries Society, Manila, Philippines, 991 p.p.
- Hunter, B., Margarelli, P. C., Jr. Lightner, D.V. and Colvin, IL. B. 1979. Ascorbic acid dependent collagen formation in Penaeid shrimp, Comparative. Biochemistry and Physiology. 64: 381-385.
- Ikeda, T., Watanabe, S., Yago, N., Horiuchi, S. 1986. Comparative biochemistry of acid proteinase from animal origins. Comparative. Biochemistry and Physiology. 8: 725-730.
- Janacarik, A. 1964. Die Verdauung der hauptnahrstoffe beim karpfen. ZtZT. Fische. DerenHilfswiss.,12: 601-684.
- Kolkovski, S., Tandler, A., Kissil, G. and Gertler, A. 1993. The effect of dietary exogenous enzymes on ingestion, assimilation, growth and survival of Gilthead seabream ((*Sparus aurata*, Sparidae, Linnaeus) *Sparus auratus*) larvae. Fish Physiology and Biochemistry, 12: 203-209.
- Kolkovski, S., Yackey, C., Czesney, S. and Dabrowski, K. 2000. The effect of microdiet supplementation of dietary digestive enzymes and enzyme activity in Yellow Perch juveniles, North American Journal of Aquaculture, 62: 130-134.
- Lavens, P., Merchie, G., Ramos, X., Leon-Hing Kujan, A., Van Hauwaert, A., Pedrazzoli, A., Neils, H. and Dee Leenheer, A. (1998-1999). Supplementation of ascorbic acid 2-monophosphate during the early post

larval stages of shrimp *Penaeus vannamei*, Aquaculture Nutrition, 5 : 73-81.

- Lee, P.G., Smith, L.L. and Lawrence, A. L. 1984. Digestive protease of Penaeus vannamei Boone: relationship between enzyme activity, size and diet, Aquaculture, 42 : 225-239.
- Lightner, D. V., Hunter, B., Magarelli, P. C. Jr and Colvin, L.B. 1979. Ascorbic acid : nutritional requirement and role in wound repair in penaeid shrimp. Proceedings of World Mariculture Society. 10 : 513-528.
- Lobao, V.L., De-Marques, H.L. Roverso, E.A. and Pazzinatto, A.C. 1995. Weight increase in *Macrobrachium rosenbergii* in Indoors, using different kinds of diets. Boletim do Instituto de Pesca, Sao Paulo BOL-INST-PESCA, 22(2): 63-69.
- Lowry, O.H. Rosebrough, J.I., Farr, A.L. and Randall, R.J. 1951. Protein measurement with Folin- phenol reagent, Journal. of Biological Chemistry., 193: 265-275.
- Margarelli, P.C., Jr., Hunter, B., lightner, D.V. and Colvin, L.B. 1979. Black death an ascorbic acid deficiency disease in Penaeid shrimp, Comparative. Biochemistry and Physiology.Physiol. B, 63 : 1083-108.
- Maugle, P. D., Deshimaru, O., Katayama, T. and Simson, K. L., 1983a, . Effect of microencapsulated amylase and bovine trypsin dietary supplements on growth and metabolism of shrimp. Bulletins of Japanese Society. Science. Fisheries., 49: 65-77.
- Maugle, P.D., Deshimaru, O., Katayama, T., Nagatani, T and Simson, K.L. 1983b. The use of amylase supplements in shrimp diets. Journal of World Mariculture. Society., 14: 25-37.
- Merchie, G., Lavens, P., Dhert, P., Dehasque., De Nelis, H., Leenheer, A., and Soorgeloos, P. 1995. Variation in ascorbic acid content in different live food organisms. Aquaculture, 134:; 325-337.
- Merchie, G., Lavens, P., Vrreth, J., Ollevier, F., Nelis, H., Leenheer, D., Storch, V. and Sorgeloos, P., 1997a., The effect of supplemental ascorbic acid in enriched live food for Clarias gariepinus larvae at start feeding. Aquaculture, 151: 245-258.
- Merchie, G., Lavens, P. and Sorgeloos, P. (1997b). Optimization of dietary vitamin-C in fish and crustacean larvae: a review. Aquaculture, 155: 165-181.
- Paul Raj, R. 1997. Introduction: In Aquaculture feed, Handbook on Aquafarming, MPEDA Publication.
- Person-Le Ruyet, J., Alexander, J.C., Thebaud, L. and Mugnier, C. 1993. Marine fish larvae feeding: formulated and live preys. Journal of World Aquaculture Society, 24: 211-224.
- Peterson, H. 1977. A simplification of protease assay method of Lowry *et al.*, which is generally more applicable. Analytical Biochemistry, 83: 346-356.
- Sebastian, C. 1996. Giant Freshwater prawn farming. In: A manual on seed production and farming of giant freshwater prawn, *Macrobrachium rosenbergii*, MPEDA. India: 55-56.
- Shiau, S.Y. and Hsu, T.S. 1994. Vitamin-C requirement of grass shrimp Penaeus monodon, as determined with L- ascorbyl- 2 - mono phosphate. Aquaculture, 122: 347-357.
- Shigucin, K. and Itoh, S. 1988. Use of Mg-L- ascorbyl-2phosphate as a vitamin-C source in shrimp diets. Journal of World Mariculture Society., 19: 168-174.
- Snedecor, G.W. and Cochran, G. 1961. Statistical methods.

Oxford and IBH publishing Co., New Delhi, 593 p.p.

- Srivastava, P.P., Tagare, M.N., D.R. Rao, Jain, K.K. and Sinha, A. 1994. Studies on the application of proteolytic enzyme, papain, on dietary incorporation and its dose-dependent influence in the digestion of protein ingredients and in increasing feed digestibility in *Cyprinus carpio*. Second Asia Pacific Conference on Agricultural Biotechnology, Madras, 160: 243 p.p
- Tacon, A.G. J. 1987. The nutrition and feeding of farmed fish and shrimp - a training manual. The essential nutrients. GCP/RLA/ 075 ITA Field Document. Food and Agriculture Organization of United Nations,

Brazil, 117 pp.

- Tolbert, B.M. 1979. Ascorbic acid metabolism and physiological function. International. Journal of Vitamin Nutrition Research, 19: 127-142.
- Wilson, R.P. 1973. Absence of ascorbic acid synthesis in channel catfish Ictalurus punctatus and blue catfish, *Ictalurus frucatus*. Comparative Biochemistry and Physiology, 46: 635-638.
- Zhong-J., Wang, J. and Mongrui, M. 1994. On application of feed enzyme preparation agent to make prawns diet, Shadong-Fish-Quilu-Yuye, 11(3): 30-32.