

# Survival and Growth Rates of Juvenile Grass Carp *Ctenopharyngodon idella* Overwintered in Ponds and Recirculating Aquaculture Systems Including a Comparison of Production Economics

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## Abstract

The survival of grass carp juveniles overwintered in ponds and RAS were compared in this study. The grass carp,  $W = 2.49 \pm 0.34\text{g}$ , were distributed among five 350 L tanks connected to a RAS and/or into three (1500 m<sup>2</sup>) ponds (P group). After 165 days, the survival rate was significantly higher in the RAS group,  $97.8 \pm 0.5\%$ , compared to  $10.9 \pm 11.4\%$  in P group. The grass carp in a RAS during the first winter significantly improved survival and good conditions of juvenile for subsequent on-growing period. The RAS provided production profit compare to loss in P group.

*Keywords:* RAS, pond culture, overwintering, cyprinids, fish production

## Introduction

Grass carp *Ctenopharyngodon idella*, are widely cultured Chinese freshwater fish (Du *et al.*, 2006) the rearing of which began more than a thousand years ago (Fu, Shen, Xu & Li, 2016). They have been cultured in c. 40 countries worldwide (FAO 2016; Pípalová, 2006) and were introduced to Europe in 1963 as a low-cost biological alternative for pond maintenance (Krupauer, 1971), particularly for control of aquatic plants (Dibble & Kovalenko, 2009; Murphy *et al.*, 2002; Schoonbee, 1991; Silva, Cruz, Pitelli & Pitelli, 2014). The major producers of grass carp are China, Bangladesh, Taiwan, Iran, and the Russian Federation (FAO 2016).

Grass carp production is likely to increase, because of their ability to utilize plant protein sources to produce high quality meat commanding a good market price (Brožová, 2005; Lin, Zeng, Zhu & Song, 2012; Lin *et al.*, 2016). Currently, production is limited by inadequate and fluctuating production of juvenile stock for on-growing in ponds. In Central Europe, the primary challenge to commercial grass carp production is the generally low survival rate and growth in ponds during the first winter, especially in fish of total length 50–70 mm (Able & Curran, 2008).

Recirculating systems have been used for winter production of juvenile common carp in Germany (Rümmeler *et al.*, 2011). A combination of pond and intensive culture in a recirculating aquaculture system (RAS) is used for producing high-quality juvenile pikeperch, especially in regions with large pond areas (Blecha, Křišťan & Policar, 2016; Policar *et al.*, 2013; Policar *et al.*, 2016). Application of such technologies to grass carp can be

positive as limitation of low winter temperatures which are not well tolerated by this species. Nevertheless, the economic benefits of rearing an herbivorous fish, with high flesh quality and a potential developing market, suggest that RAS culture over winter, followed by pond culture under higher water temperatures would be profitable and effective. The aim of this study therefore was to investigate survival of 3-month-old grass carp in a RAS group compared with pond group overwintering.

## Materials and Methods

Thirty thousand juvenile grass carp were obtained from experimental ponds at the University of South Bohemia, Faculty of Fisheries and Protection of Waters (USB FFPW). Body weight (W) ( $2.49 \pm 0.34$  g) was measured to the nearest 0.01 g (PCB 1000-2, Kern, Germany), and total length (TL) ( $59.5 \pm 1.9$  mm) was measured with callipers to the nearest 0.1 mm (Polícar *et al.*, 2011a). At the end of October, the experimental fish were divided into two equal groups (P and RAS) for 163 – 165 day culture during wintering.

The P group was maintained under traditional pond monoculture in three ponds, each stocked with 5000 fish (3.3 fish for 1 m<sup>2</sup>). Ponds (1500 m<sup>2</sup>, 1.5 m average depth) in two locations (Vodňany and Horažďovice, Czech Republic) with varying hydrographic characteristics were chosen. Water was supplied from channel of local river Blanice in Vodňany or river Otava in Horažďovice. The diet consisted of naturally occurring food (winter and spring zooplankton and benthos). Temperature was recorded every 2 h with an automatic temperature sensor (Minikin T, Environmental Measuring Systems, Brno, Czech Republic) in each pond 1 meter below water surface. Water quality and pond control (oxygen =  $69.0 \pm 7.5$  %, pH =  $6.89 \pm 0.35$ , NH<sup>4+</sup> =  $0.26 \pm 0.1$  mg l<sup>-1</sup>, and NO<sub>3</sub> =  $0.48 \pm 0.2$  mg l<sup>-1</sup>) was recorded once a week. After 163-165 days, fish were harvested using a net cage in the outlet channel according to Polícar, Blaha, Kristan & Stejskal (2011b) and Stejskal, Kouril, Polícar, Hamackova & Musil (2009). The survival rate (SR %) was calculated as  $SR = (SF_2/SF_1) \times 100$ , where SF<sub>2</sub> is number of harvested fish, and SF<sub>1</sub> is number of stocked fish. A sample of 50 representative juveniles was collected from each pond for biometric measurements. Fish were anaesthetized with clove oil before handling according to Kristan, Stara, Turek, Polícar & Velisek (2012). Specific growth rate (SGR) and condition were calculated by the formulae:  $SGR = 100 t^{-1} \ln(W_f/W_i)$ , where W<sub>i</sub> and W<sub>f</sub> are initial and final body weight in g, and t is the growing period in days, and Fulton's condition coefficient  $K = (W/TL^3) \times 100$ , where W is final body weight and TL is final total length.

Fish of the RAS group were stocked into a recirculating aquaculture system at USB FFPW comprising five cylindrical 350 l tanks (~8 fish l<sup>-1</sup>, 3000 fish per tank) fitted with mechanical (Ratz Aqua Polymer Technik, Remscheid, Germany) and biological filters (Nexus 310, Evolution Aqua, Wigan, UK). Water temperature and quality parameters (oxygen level =  $80.0 \pm 5.5$  %, pH =  $6.94 \pm 0.22$ , NH<sup>4+</sup> =  $0.12 \pm 0.05$  mg l<sup>-1</sup> and NO<sub>3</sub> =  $0.33 \pm 0.3$  mg l<sup>-1</sup>) were measured daily. Fish were fed a commercial diet (Inicio 917, size 1.1 mm BioMar Ltd) at 1% of current biomass. All fish were collected and weighed at 30-33 day intervals. A sample of 30 representative juveniles was collected from each tank for biometric measurements. The SR, SGR, and K were calculated as for the P group assessment. The feed conversion ratio (FCR) was calculated as  $FCR = F/(W_f - W_i)$ , where F is total feed consumption, and W<sub>i</sub>, W<sub>f</sub> are initial and final body weight, respectively.

At the conclusion of the trial, an economic analysis of culture methods was conducted in cooperation with an economist of the USB FFPW, taking into consideration total costs (stocking material, salaries, feed, overheads, and depreciation), revenue, and profit or loss of each production group.

## Statistical Analysis

Data were expressed as mean  $\pm$  SD. First, datasets were checked for normal distribution using the Kolmogorov-Smirnov test and homoscedasticity of variance (Bartlett's test). Data meeting normality criteria were then analysed with single-factor ANOVA. When a significant ( $p < 0.05$ ) difference was found, a Tukey's multiple comparison tests was used. Statistical assessment of all data was carried out with Statistica 7.0 (StatSoft, Inc., Czech Republic).

## Results

Pond temperatures fluctuated throughout the experimental period with the lowest value being 0.9°C. The mean temperature was  $4.2 \pm 2.6^\circ\text{C}$  (range 0.9-13.1 °C) in the ponds and a stable  $20.8 \pm 0.4^\circ\text{C}$  in the RAS (Fig. 1).

The mean survival rate in the RAS was significantly higher ( $97.8 \pm 0.5\%$ ) compared to  $10.9 \pm 11.4\%$  in P group (Table 1).

The SGR at the conclusion of the trial was significantly higher ( $p < 0.05$ ) in RAS at  $0.49 \pm 0.08\% \text{ d}^{-1}$  than  $-0.12 \pm 0.10\% \text{ d}^{-1}$  in P group (Table 2). The mean final TL of juveniles cultured in RAS was  $70.1 \pm 5.2 \text{ mm}$ , compared  $61.4 \pm 1.8 \text{ mm}$  for P group. Significantly higher ( $p < 0.05$ ) body weight was recorded in RAS ( $5.59 \pm 0.89 \text{ g}$ ) compared to P group ( $2.07 \pm 0.53 \text{ g}$ ) (Table 1, Fig. 2).

In P group, Fulton's condition coefficient was significantly ( $p < 0.05$ ) lower at the end of the culture period than at the beginning, while, in the RAS group, final  $K$  was higher (Table 1). The feed conversion ratio calculating in the RAS group only, was  $\text{FCR} = 1.69 \pm 0.07$ .

Results of the economic study are shown in Table 2. The total production costs of equal-sized fish groups were lower for P group (€1,301) compared to rearing in RAS group (€2,364). Revenue of €2,714 was obtained from RAS compared to €304 for P group. Production profit of €350 was obtained in RAS group compared to a production loss of -€997 in the P group.

## Discussion

The ability of fish to survive the first winter is an important determinant of success in fish production from the perspective of having enough juveniles for on-growing to market size category. Survival of grass carp juveniles in RAS was significantly higher than in ponds. The low pond survival rate of grass carp was probably influenced primarily by low temperatures (Hurst, 2007; Binder *et al.*, 2015). Grass carp tolerate a range of water temperatures from 0 to 33°C (Cudmore & Mandrak, 2004). Similar survival rate achieved Jones *et al.* (2017) in different Canadian lakes.

Limited food availability also plays a major role in survival (Ludsin & DeVries, 1997; Post, Kitchell & Hodgson, 1998). Grass carp feeding activity begins at 7-8°C, intensive feeding occurs only when water temperature is at least 20°C (Cudmore & Mandrak, 2004). Chilton & Muoneke (1992) stated that grass carp rarely fed at temperatures below 3°C and, while in their over-winter habitat, they do not feed at all (Fischer & Lyakhoich, 1973). Under pond conditions, fish not only lose weight, but are in relatively poor condition when water temperatures begin to rise (David, 2006). The weakened fish are vulnerable to bacterial and fungi contamination and disease (Binder *et al.*, 2015) and are not suitable for further aquaculture production. According to available published data the immune system is non-functional in grass carp for most of the

overwintering period. Water temperature only reached 14 °C at the end of the culture period of this study. Predation by birds is an additional problem with pond culture during winter and spring (Adamek, Kortan & Flašhans, 2007; Kortan, Adamek, Flašhans & Piackova, 2008). Grass carp may be attacked by mammalian or even insect predators-dragonfly larvae (Akpona *et al.*, 2015). Injured juveniles may also contract secondary bacterial and fungi infection and disease (Binder *et al.*, 2015).

Rearing in RAS positively affected survival rate and health and provided protection against predators. David (2006) also reported these benefits in the culture of European catfish *Silurus glanis* in RAS during winter. Similar results were observed in grass carp by Du *et al.* (2006) who reported SGR of 0.67% at a feeding rate 1% of biomass and a temperature of 25°C. Du *et al.* (2006) recommended an optimum feeding rate of 1.97% body weight d<sup>-1</sup> for fish around 3 g. Shireman, Colle & Rottmann (1977) recommended 2.1 fish per litre for optimal growth rate. In this experiment, the high density of fish (8 fish per litre) was not a problem for wintering storage time in RAS. The daily feeding rate was only 1% in comparison with *ad libitum* in study Shireman, Colle & Rottmann (1977). The condition of fish reared in RAS was higher than pond reared fish. Similar results were obtained by Cai, Fang, Johnson, Lin, Tu, Liu & Huang (2014), who demonstrated *K* of 1.4–1.6 in RAS cultured grass carp of TL 130 mm. In this study, the feed conversion ratio was similar (FCR = 1.69 ± 0.07) and compared to Ming, Ye, Zhang, Xu & Xie (2015), who reported FCR of 1.7–2.1 in grass carp.

The results of economic analysis showed a profit in the RAS of €350, compared to loss of -€997 for the pond group (Table 2), stressing the importance of including RAS in the grass carp production cycle.

This seems to be the first study comparing overwintering of grass carp juveniles in ponds with their maintenance in RAS. On the basis of better obtained production data (e.g. survival rate, Fulton's condition coefficient) and economic analysis, the recirculating aquaculture system showed potential to increase the production and profitability of grass carp juveniles for on-growing culture in Central Europe which can significantly increase production of this species in mentioned region.

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## References

- Able, K.W. & Curran, M.C. (2008) Winter Mortality in Some Temperate Young-of-the-Year Fishes. *Bulletin New Jersey Academy of Science*, 53(2):1–5
- Adamek, Z., Kortan, J., & Flašhans, M. (2007). Computer-assisted image analysis in the evaluation of fish wounding by cormorant [*Phalacrocorax carbo sinensis* (L.)] attacks. *Aquaculture International*, 15, 211–216. <http://dx.doi.org/10.1007/s10499-007-9087-0>
- Akpona, A.H., Djagoun, C.A.M.S., Harrington, L.A., Kabré, A.T., Mensah, G.A., & Sinsin, B. (2015). Conflict between spotted-necked otters and fishermen in Hlan River, Benin. *Journal for Nature Conservation*, 27, 63–71. <http://dx.doi.org/10.1016/j.jnc.2015.06.007>
- Blecha, M., Křišťan, J., & Polícar, T. (2016). Adaptation of intensively reared pikeperch (*Sander lucioperca*) juveniles to pond culture and subsequent re-adaptation to a recirculation aquaculture system. *Turkish Journal of Fisheries and Aquatic Sciences*, 16, 15–18. [http://dx.doi.org/10.4194/1303-2712-v16\\_1\\_02](http://dx.doi.org/10.4194/1303-2712-v16_1_02)



- Binder, T.R., O'Connor, C.M., McConnachie, S.H., Wilsona, S.M., Nannini, M.A., Wahl, D.H., & Cooke, S.J. (2015). Is winter worse for stressed fish? The consequences of exogenous cortisol manipulation on over-winter survival and condition of juvenile largemouth bass. *Comparative biochemistry and physiology. Part A, Molecular & integrative physiology*, 187, 97–102. <http://dx.doi.org/10.1016/j.cbpa.2015.05.008>
- Brožová, M. (2005). Fish - Annual Report of Ministry of Agriculture. *Ministry of Agriculture of the Czech Republic*, 40p – in Czech only.
- Cai, L., Fang, M., Johnson, D., Lin, S., Tu, Z., Liu, G., & Huang, Y. (2014). Interrelationships between feeding, food deprivation and swimming performance in juvenile grass carp. *Aquatic Biology*, 20, 69–76. <http://dx.doi.org/10.3354/ab00546>
- Cudmore, B. & Mandrak, N.E. (2004). Biological Synopsis of Grass Carp (*Ctenopharyngodon idella*). *Canadian Manuscript Report of Fisheries and Aquatic Sciences*. 44p.
- David, J.A. (2006). Water quality and accelerated winter growth of European catfish using an enclosed recirculating system. *Water and Environment Journal*, 20, 233–239. <http://dx.doi.org/10.1111/j.1747-6593.2006.00021.x>
- Dibble, E.D. & Kovalenko, K. (2009). Ecological impact of grass carp: A review of the available data. *Journal of Aquatic Plant Management*, 47, 1–15.
- Du, Z.Y., Liu, Y.J., Tian, L.X., He, J.G., Cao, J.M., & Liang, G.Y. (2006). The influence of feeding rate on growth, feed efficiency and body composition of juvenile grass carp (*Ctenopharyngodon idella*). *Aquaculture International*, 14, 247–257. <http://dx.doi.org/10.1007/s10499-005-9029-7>
- FAO (2016). Food and Agriculture Organization of the United Nations. [http://www.fao.org/fishery/culturedspecies/Ctenopharyngodon\\_idellus/en](http://www.fao.org/fishery/culturedspecies/Ctenopharyngodon_idellus/en) (accessed July, 2017).
- Fischer, Z. & Lyakhnovich, V.P. (1973). Biology and energetics of grass carp (*Ctenopharyngodon idella*, Val.) *Polish Archives of Hydrobiology*, 20(4), 309–318.
- Fu, J., Shen, Y., Xu, X., & Li, J. (2016). Genetic parameter estimates for growth of grass carp, *Ctenopharyngodon idella*, at 10 and 18 months of age. *Aquaculture*, 450, 342–348. <http://dx.doi.org/10.1016/j.aquaculture.2015.08.018>
- Hurst, T.P. (2007). Causes and consequences of winter mortality in fishes. *Journal of Fish Biology*, 71, 315–345. <http://dx.doi.org/10.1111/j.1095-8649.2007.01596.x>
- Chilton, E. W. & M. I. Muoneke, M.I. (1992). Biology and management of grass carp (*Ctenopharyngodon idella*, Cyprinidae) for vegetation control: a North American perspective. *Reviews in Fish Biology and Fisheries*, 2, 283–320.
- Jones, L.A., Drake, D.A.R., Mandrak, N.E., Jerde, C.L., Wittmann, M.E., Lodge, D.M., van der Lee, A.S., Johnson, T.B., and Koops, M.A. 2017. Modelling Survival and Establishment of Grass Carp, *Ctenopharyngodon idella*, in the Great Lakes Basin. *Canadian Science Advisory Secretariat*, 52.
- Kortan, J., Adamek, Z., Flajshans, M., & Piackova, V. (2008). Indirect manifestation of cormorant (*Phalacrocorax carbo sinensis* (L.)) predation on pond fish stock. *Knowledge and Management of Aquatic Ecosystems*, 389, 345–349. <http://dx.doi.org/10.1051/kmae:2008006>
- Kristan, J., Stara, A., Turek, J., Policar, T., & Velisek, J. (2012). Comparison of the effects of four anaesthetics on haematological and blood biochemical profiles in pikeperch (*Sander lucioperca* L.). *Neuroendocrinology Letters*, 33, 66–71.
- Krupauer, V. (1971). Morphology of grass carp (*Ctenopharyngodon idella* Val.) and acclimatization in ponds during first three years. *USB RIFH Vodnany* 9, 49–97 – in Czech only.
- Lin, W.L., Zeng, Q.X., Zhu, Z.W., & Song, G.S. (2012). Relation between protein characteristics and TPA texture characteristics of crisp grass carp (*Ctenopharyngodon idellus* C. et V) and grass carp (*Ctenopharyngodon idellus*). *Journal of Texture Studies*, 43, 1–11. <http://dx.doi.org/10.1111/j.1745-4603.2011.00311.x>
- Lin, W.L., Yang, X.Q., Li, L.H., Hao, S.X., Wang, J.X., Huang, H., Wei, Y., & Wu, Y.Y. (2016). Effect of ultrastructure on changes of textural characteristics between crisp grass carp (*Ctenopharyngodon idellus* C. Et V) and grass carp (*Ctenopharyngodon idellus*) inducing heating treatment. *Journal of Food Science*, 81, 404–411. <http://dx.doi.org/10.1111/1750-3841.13189>
- Ludsin, S.A., & De Vries, D.R. (1997). First-year recruitment of largemouth bass: the interdependency of early life stages. *Ecological Applications*, 7, 1024–1038.
- Ming, J.H., Ye, J.Y., Zhang, Y.Z., Xu, P., & Xie, J. (2015). Effects of dietary reduced glutathione on growth performance, non-specific immunity, antioxidant capacity and expression levels of IGF-I and HSP70 mRNA of grass carp (*Ctenopharyngodon idella*). *Aquaculture*, 438, 39–46. <http://dx.doi.org/10.1016/j.aquaculture.2014.12.038>
- Murphy, J.E., Beckmen, K.B., Johnson, J.K., Cope, R.B., Lawmaster, T., & Beasley, V.R. (2002). Toxic and feeding deterrent effects of native aquatic macrophytes on exotic grass carp (*Ctenopharyngodon idella*). *Ecotoxicology*, 11, 243–254. <http://dx.doi.org/10.1023/A:1016344103565>
- Pípalová, I. (2006). A review of grass carp use for aquatic weed control and its impact on water bodies. *Journal of Aquatic Plant Management*, 44, 1–12.
- Policar, T., Blecha, M., Křišťan, J., Mráz, J., Velišek, J., Stará, A., Stejskal, V., Malinovskyi, O., Svačina, P., & Samarín, A.M. (2016). Comparison of production efficiency and quality of differently cultured pikeperch (*Sander lucioperca* L.) juveniles as a valuable product for ongrowing culture. *Aquaculture International*, 24, 1607–1626. <http://dx.doi.org/10.1007/s10499-016-0050-9>
- Policar, T., Podhorec, P., Stejskal, V., Kozák, P., Švinger, V. & Alavi, S.M.H. (2011a). Growth and survival rates, puberty and fecundity in captive common barbel (*Barbus barbus* L.) under controlled conditions. *Czech Journal of Animal Science*, 56, 433 – 442.
- Policar, T., Blaha, M., Kristan, J., & Stejskal, V. (2011b). High-quality and stable production of pond-cultured pikeperch (*Sander lucioperca*) juveniles under pond conditions. *Edition of Methodics, USB FFPW*, 110p.

- Polcar, T., Stejskal, V., Kristan, J., Podhorec, P., Svinger, V., & Blaha, M. (2013). The effect of fish size and stocking density on the weaning success of pond-cultured pikeperch *Sander lucioperca* L. juveniles. *Aquaculture International*, 21, 869–882. <http://dx.doi.org/10.1007/s10499-012-9563-z>
- Post, D.M., Kitchell, J.F., & Hodgson, J.R. (1998). Interactions among adult demography, spawning date, growth rate, predation, overwinter mortality, and the recruitment of largemouth bass in a northern lake. *Canadian Journal of Fisheries and Aquatic Sciences*, 55, 2588–2600. <http://dx.doi.org/10.1139/cjfas-55-12-2588>
- Rümmler, F., Pfeifer, M., Jährling, R., Rank, H., Weichler, F., & Schiewe, S. (2011). Emissionen der Warmwasserfischzucht. *Schriftenreihe des Lfulg*, 4, 101p – in Germany only.
- Shireman, J.V., Colle, D.E., & Rottmann R.W. (1977). Intensive culture of grass carp, *Ctenopharyngodon idella*, in circular tanks. *Journal of Fish Biology*, 11, 267–272. <http://dx.doi.org/10.1111/j.1095-8649.1977.tb04119.x>
- Schoonbee, H.J. (1991). Biological control of fennel-leaved pondweed, *Potamogeton pectinatus* (Potamogetonaceae), in South Africa. *Agriculture, Ecosystems and Environment*, 37, 231–237. [http://dx.doi.org/10.1016/0167-8809\(91\)90152-N](http://dx.doi.org/10.1016/0167-8809(91)90152-N)
- Silva, A.F., Cruz, C., Pitelli, R.L.C.M., & Pitelli, R.A. (2014). Use of grass carp (*Ctenopharyngodon idella*) as a biological control agent for submerged aquatic macrophytes. *Planta Daninha* 32, 765–773.
- Stejskal, V., Kouril, J., Polcar, T., Hamackova, J., & Musil, J. (2009). Growth pattern of all-female perch (*Perca fluviatilis* L.) juveniles—is monosex perch culture beneficial? *Journal of Applied Ichthyology*, 25, 432–437. <http://dx.doi.org/10.1111/j.1439-0426.2009.01253.x>

**Table 1.** The number of stocked and harvested fish, survival rate, initial and final weight, total length, Fulton’s condition coefficient *K*, and specific growth rate of grass carp overwintered in ponds and RAS.

	Stocked fish (n)	Harvested fish (n)	Survival rate (%)	Initial		Final		SGR	
				W (g)	TL (mm)	W (g)	TL (mm)	<i>K</i>	(% d <sup>-1</sup> )
Ponds	15 000	1642	10.9 ± 11.4	2.49 ± 0.41	59.9 ± 1.7	2.07 ± 0.53	61.4 ± 1.8	0.90 ± 0.31	-0.12 ± 0.10
				2.49 ± 0.33	59.3 ± 2.1	5.59 ± 0.89	70.1 ± 5.2	1.62 ± 0.14	0.49 ± 0.08
RAS	15 000	14 669	97.8 ± 0.5						

**Table 2.** Economic comparison of juvenile grass carp overwintering practices.

	Ponds €	Pond characteristics	RAS €	RAS characteristics
Costs	555	15 000 fish	555	15 000 fish
	483	0.5 h per day	965	1 h per day
	0		229	80
	156	15% of costs	262	15% of costs
	107		422	
	1301	Total production 1642 fish	2364	Total production 14 669 fish
Revenue	304	Price per fish € 0.185	2714	Price per fish € 0.185
Profit/loss	€ -997		€ 350	

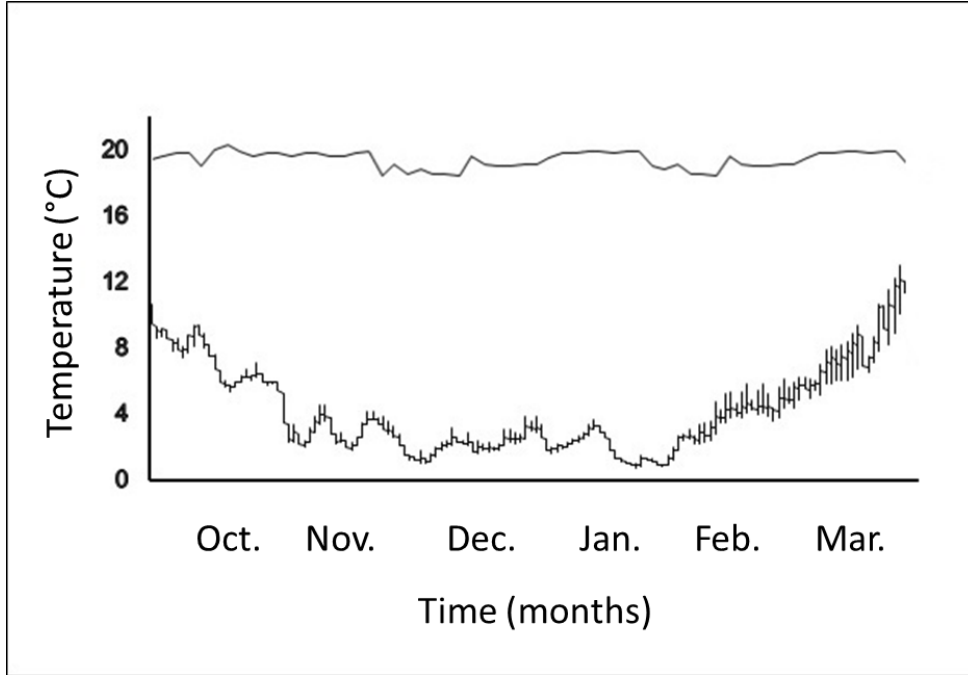


Figure 1. Temperature in ponds and RAS during the juvenile grass carp rearing trial.

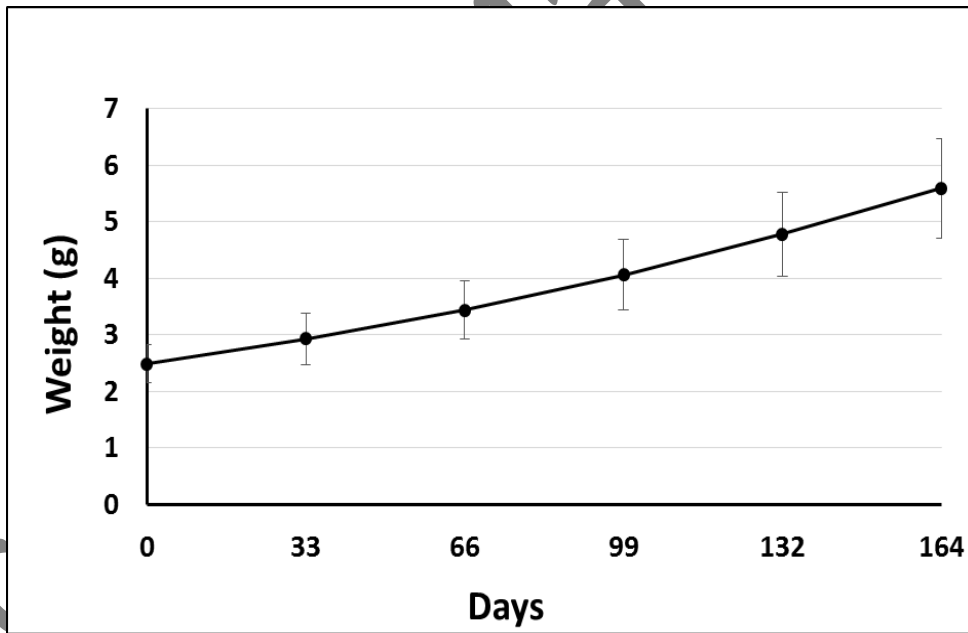


Figure 2. Growth rate in recirculation aquaculture system.